

Northumbria Research Link

Citation: Panagiotou, Marios, Vogiatzis, Ioannis, Jayasekera, Geeshath, Louvaris, Zafeiris, MacKenzie, Alison, McGlinchey, Neil, Baker, Julien, Church, Alistair, Peacock, Andrew and Johnson, Martin (2018) Validation of impedance cardiography in pulmonary arterial hypertension. *Clinical Physiology and Functional Imaging*, 38 (2). pp. 254-260. ISSN 1475-0961

Published by: Wiley-Blackwell

URL: <https://doi.org/10.1111/cpf.12408> <<https://doi.org/10.1111/cpf.12408>>

This version was downloaded from Northumbria Research Link: <http://nrl.northumbria.ac.uk/30047/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)



Northumbria
University
NEWCASTLE

Validation of impedance cardiography in pulmonary arterial hypertension

Marios Panagiotou, Ioannis Vogiatzis, Geeshath Jayasekera, Zafeiris Louvaris, Alison Mackenzie, Neil McGlinchey, Julien S. Baker, Alistair C. Church, Andrew J. Peacock, Martin K. Johnson

Summary

Background

Non-invasive methods of measuring cardiac output are highly desirable in pulmonary arterial hypertension (PAH). We therefore sought to validate impedance cardiography (ICG) against thermodilution (TD) and cardiac magnetic resonance (CMR) in the measurement of cardiac output in patients under investigation for PAH.

Methods

A prospective, cross-sectional study was performed to compare single-point measurements of cardiac output obtained by impedance cardiography (COICG) technology (PhysioFlow®) with (i) contemporaneous TD measurements (COTD) at rest and steady-state exercise during right heart catheterization and (ii) CMR measurements (COCMR) at rest obtained within 72 h.

Results

Paired COICG and COTD measurements were obtained in 25 subjects at rest and 16 subjects at exercise. COCMR measurements were obtained in 16 subjects at rest. There was unsatisfactory correlation and agreement between COICG and COTD at rest ($r = 0.42$, $P = 0.035$; bias: 1.21 l min^{-1} , 95% CI: -2.33 to 4.75 l min^{-1}) and exercise ($r = .65$, $P = .007$; bias: 1.41 l min^{-1} ; 95% CI: -3.99 to 6.81 l min^{-1}) and in the change in COICG and COTD from rest to exercise ($r = 0.53$, $P = 0.033$; bias: 0.76 l min^{-1} , 95% CI: -3.74 to 5.26 l min^{-1}). There was also a lack of correlation and unsatisfactory agreement between resting COICG and COCMR ($r = 0.38$, $P = 0.1$; bias: 1.40 l min^{-1} , 95% CI: -2.48 to 5.28 l min^{-1}). In contrast, there was close correlation and agreement between resting COTD and COCMR ($r = 0.87$, $P < 0.001$; bias: -0.16 l min^{-1} , 95% CI: -1.97 to 1.65).

Conclusions

In a representative population of patients under investigation for PAH, ICG showed insufficient qualitative and quantitative value in the measurement of resting and exercise cardiac output when compared with TD and CMR.

Introduction

Pulmonary arterial hypertension (PAH) is characterized by increased pulmonary vascular resistance resulting in progressing right ventricular failure, left ventricular underfilling and reduced cardiac output (Galiè et al., 2015). Cardiac output and cardiac index either at baseline or following therapeutic interventions have been identified among the most robust indicators of right ventricular function and prognosis in PAH (D'Alonzo et al., 1991; McLaughlin et al., 2002; Humbert et al., 2010). Accordingly, normalization of cardiac index has become an important treatment goal endpoint in PAH (McLaughlin et al., 2013; Galiè et al., 2015).

Thermodilution (TD) (Ganz et al., 1971) remains the reference standard for the measurement of cardiac output in PAH; it is reliable, detects directional changes and correlates well with the Fick technique in patients with precapillary pulmonary hypertension even in the presence of low cardiac output and

significant tricuspid regurgitation (Hoepfer et al., 1999). More recently, cardiac magnetic resonance (CMR) has offered a non-invasive method of assessment of the right ventricular function (Hundley et al., 1995) and has been successfully validated against the Fick method in the measurement of cardiac output in PAH (Mauritz et al., 2008). However, both TD and CMR have certain limitations. Thermodilution is invasive and subject to technical errors (Stevens et al., 1985; Nishikawa & Dohi, 1993), whereas CMR is restricted by a number of factors including higher cost and limited availability, time-consuming analysis, incompatibility with infusion pumps and requirement of breath holding, which may be difficult for patients (Peacock & Vonk Noordegraaf, 2013).

Impedance cardiography (ICG) relies on the principle that variations in the bioimpedance to an alternating current flow during cardiac ejection result in a specific waveform from which stroke volume can be calculated (Charloux et al., 2000). Impedance cardiography is attractive due to its potential for non-invasive measurements of cardiac output at rest and activity on a beat-to-beat basis (Bour & Kellett, 2008; Tang & Tong, 2009); however, it has not been adequately validated in PAH to date. We therefore sought to investigate the qualitative and quantitative validity of ICG against TD and CMR in the measurement of resting and exercise cardiac output in patients under investigation for PAH.

Materials and methods

Study design

A prospective, cross-sectional study was performed. All consecutive patients who underwent right heart catheterization for investigation of PAH in the Scottish Pulmonary Vascular Unit between December 2014 and November 2015 were eligible. Approval from the West of Scotland Research Ethics Committee (14/WS/1075) and written consent were obtained.

Single-point measurements of cardiac output were obtained using impedance cardiography (COICG) and compared with (i) contemporaneous TD measurements (COTD) at rest and steady-state exercise during right heart catheterization and (ii) CMR measurements (COCMR) at rest obtained within 72 h. All measurements were taken in a blinded fashion, and all COICG and COCMR measurements were taken by single operators (MP and GJ, respectively).

Measurements of COICG and COTD at exercise were obtained in patients who consented to exercise in the supine position during right heart catheterization. Subjects either performed straight leg raising or cycled on an electronically braked ergometer (Angio 917900, Lode B.V., Groningen, the Netherlands) secured to the catheterization table. Cycling rate was maintained at 60 revolutions·min⁻¹ and workload kept constant matching 50% of patient's maximal oxygen uptake (VO₂ max) measured the previous day during an upright cycle cardiopulmonary exercise test. In order to reach physiological steady state, measurements were taken during the sixth minute of exercise.

Right heart catheterization & thermodilution

Diagnostic right heart catheterization was performed according to current guidelines (Galiè et al., 2015). A balloon-tipped, double-lumen, fluid-directed 7-Fr Swan Ganz catheter was advanced through an 8-F introducer sheath inserted into the right internal jugular vein. The positioning of the catheter was confirmed with the presence of pulmonary artery pressure waveform from its distal port and fluoroscopic visualization of its distal tip in the pulmonary artery. Cardiac output was measured with the TD technique (Ganz et al., 1971) following the injection of 10 ml sterile, ice-cold isotonic (0.9%) saline through the

proximal (right atrial) lumen of the Swan–Ganz catheter. The drop in temperature was measured at the distal thermistor. The final value used was the average of three measurements agreeing within 10%.

Impedance cardiography

Cardiac output obtained by ICG measurements at rest and exercise were obtained using a commercially available technology (PhysioFlow® Enduro; Manatec Biomedical, Macheren, France), a detailed description of which can be found elsewhere (Charloux et al., 2000; Richard et al., 2001). In brief, PhysioFlow uses variations in the transthoracic impedance to a high-frequency (75 kHz), low-amperage (1.8 mA) alternating current across the thorax during cardiac ejection to calculate stroke volume. Its basic equation for calculating stroke volume (Signal-Morphology-ICG™) has resulted from the introduction of a novel approach to ICG in order to overcome the confounding variables of basal thoracic impedance (including thorax morphology, fluid and gas content, blood resistivity and skin thickness) and interelectrode distance. This is achieved by relying only on change in impedance (dZ) to establish stroke volume, independent of baseline impedance (Z_0) (Charloux et al., 2000; Tan et al., 2006; Tonelli et al., 2011).

In the PhysioFlow algorithm, an initial evaluation of stroke volume index is computed at rest during 24 consecutive heart beats (autocalibration procedure) by using the largest impedance variation during systole ($Z_{\max} - Z_{\min}$), the largest rate of variation in the impedance signal (contractility index, dZ/dt_{\max}), the thoracic fluid inversion time (time in ms between the start of the QRS complex and the first nadir after the peak of the ejection velocity on the first mathematical derivative, a surrogate of the left ventricular ejection time), heart rate and pulse pressure (systolic minus diastolic arterial pressure). Cardiac output ($l \text{ min}^{-1}$) is then calculated by multiplying the stroke volume index by body surface area and heart rate [R-R interval determined on the electrocardiographic (ECG) first derivative] (Charloux et al., 2000; Bour & Kellett, 2008; Tonelli et al., 2011).

Cardiac output obtained by ICG measurements required skin preparation with shaving, scraping and cleaning for the application of six transcutaneous electrodes (PhysioFlow PF50; Manatec Biomedical, Macheren, France) and performed by a single investigator (MP) as instructed/trained by the manufacturer. Two emitting electrodes were applied at the left base of the neck, above the supraclavicular fossa and two sensing electrodes along the xiphoid. In the exercising patients, the two sensing electrodes were positioned at the left paravertebral area, at the level of xiphoid. One electrode at the mid-sternum and one at the left lateral chest wall were used to provide a single electrocardiogram lead. Subjects' sex, age, height, weight and supine systolic and diastolic blood pressure were logged in the PhysioFlow software and autocalibration was initiated after a period of 5 min during which subjects remained still and relaxed on the catheterization table. Signal quality was verified by visualizing the electrocardiographic tracing and its first derivative ($dECG/dt$) and the impedance waveform (ΔZ) and its first derivative (dZ/dt) (Charloux et al., 2000) and approved by the manufacturer following manual review in a blinded fashion. Cardiac output obtained by ICG measurements were obtained thereafter in 10-s intervals throughout right heart catheterization and downloaded to a purpose-specific notebook computer (Lenovo; North York, Canada). Manual review of the data and artefact detection was performed retrospectively. Data thought to be physiologically implausible, that is stroke volume for a given subject greater than their mean stroke volume $\pm 20\%$ (at unchanged heart rate, i.e. variation $\leq 5 \text{ beats min}^{-1}$), were deleted from the series. The final value of COICG was the average of all values obtained during TD series.

Cardiac magnetic resonance

Phase-contrast CMR was performed on a 1.5-T MAGNETOM Avanto MRI scanner (Siemens, Munich, Germany) during simultaneous electrocardiogram recording and continuous breathing. A velocity-encoded, k-space-segmented, gradient-echo sequence was used to generate 30 matched pairs of phase and

magnitude images (echo time/repetition time/flip angle/slice thickness/temporal resolution/image matrix/field of view/in-plane resolution/velocity encoding range: TE 2.18/TR 29.9/30°/5 mm/dependent on heart rate/256 × 256/320 mm × 240 mm/1.25 mm × 1.25 mm/0–150 cm s⁻¹). Retrospective electrocardiogram gating was used to ensure complete cardiac cycle coverage. The average acquisition time was 2–3 min, depending on heart rate. Left ventricular stroke volume was measured 2–4 cm above the aortic valve and distal to the coronary arterial ostia (Mauritz et al., 2008). Imaging analysis was performed using the Argus software (Siemens, Erlangen, Germany).

Statistical analysis

Data are presented as means ± standard deviation (SD). Pearson's correlation (*r*) was calculated as measures of raw associations between measurements. *P* values <0.05 were considered significant. The agreement between the three techniques was analysed in a pairwise manner using the Bland–Altman analysis (Bland & Altman, 1986). All analyses were performed using the SPSS statistical software (v. 20, IBM SPSS Statistics, Chicago, IL, USA).

Results

Patient characteristics

Twenty-eight patients were studied between November 2014 and December 2015. Due to failed PhysioFlow autocalibration in three patients, the final analysis included 25 patients; all of whom had paired COICG and COTD measurements at rest; 16 of those patients also had paired measurements of COICG and COTD at exercise (five patients performed straight leg raise and 11 patients cycled) and 16 had measurements of COCMR at rest within 72 h.

Clinical and physiological patient characteristics are presented in Tables 1 and 2 and measurements of COICG, COTD and COCMR in Table 3. None of the patients had significant cardiac shunt. Echocardiographic evidence of tricuspid valve regurgitation was present in all patients: 14, trace to mild; seven mild to moderate; and three moderate/severe to severe. Atrial fibrillation was present in four patients.

Impedance cardiography versus cardiac magnetic resonance

There was no correlation between COICG and COCMR (*P* = 0.1), and the agreement between ICG and CMR was poor (bias: 1.40 l min⁻¹, 95% CI: -2.48 to 5.28 l min⁻¹; Fig. 1, Table 3).

Thermodilution versus cardiac magnetic resonance

There was a strong correlation (*r* = 0.87, *P* < 0.01) and good agreement between COTD and COCMR (bias: -0.16 l min⁻¹, 95% CI: -1.97 to 1.65; Fig. 1, Table 3).

All the above results remained essentially unchanged when the subset of PAH patients (*n* = 14) was examined separately (data not shown).

Discussion

This study tested ICG against TD and CMR in the measurement of cardiac output in patients under investigation for PAH. Whereas there was a close correlation and agreement between TD and CMR, ICG technology showed insufficient correlation and significant overestimation of cardiac output with at least

twice as wide limits of agreement compared with TD and CMR. These findings apply for both rest and exercise conditions and the change from rest to exercise and did not change when the subset of PAH patients was examined separately.

A previous validation of the PhysioFlow technology against TD (Tonelli et al., 2011) in 39 patients with suspected or known pulmonary hypertension of various aetiologies showed stronger correlations for resting cardiac output ($r = 0.94$) and considerably smaller bias (0.3 l min^{-1}) but still wide limits of agreement (-2.2 to 2.8 l min^{-1}). Elsewhere, relative changes in measurements of cardiac index by PhysioFlow during incremental exercise were able to help identify subjects with severe PAH, but the incremental value of PhysioFlow over peak oxygen uptake was relatively modest and measurements were not possible in a quarter of patients (Ferreira et al., 2012). Nonetheless, change in cardiac index measured by PhysioFlow has been associated with 6-min walk distance and heart rate recovery in 30 patients with pulmonary hypertension of various aetiologies (Tonelli et al., 2013).

A different ICG device has shown strong correlation with Fick ($r = 0.84$) and TD ($r = 0.80$) in the measurement of resting cardiac output and strong agreement with both methods in the measurement of resting cardiac index in 39 subjects with precapillary pulmonary hypertension (bias: $-0.13 \text{ l min}^{-1} \text{ m}^{-2}$, 95% CI: -1.05 to $0.79 \text{ l min}^{-1} \text{ m}^{-2}$ versus Fick; bias: $-0.23 \text{ l min}^{-1} \text{ m}^{-2}$, 95% CI: -1.49 to $0.63 \text{ l min}^{-1} \text{ m}^{-2}$ versus TD) (Yung et al., 2004). In another study of 65 patients with precapillary pulmonary hypertension, whole-body ICG showed good correlation with TD ($r = 0.715$) and Fick ($r = 0.653$) and satisfactory agreement with both methods in the measurement of resting cardiac output (bias: 0.50 , 95% CI: -1.61 to 2.61 l min^{-1} versus TD; bias: -0.54 , 95% CI: -2.57 to 1.49 l min^{-1} versus Fick) (Taniguchi et al., 2013). Finally, transthoracic bioreactance cardiography, a similar technology to ICG, showed acceptable results versus TD ($r = 0.60$, bias: -0.81 , 95% CI: -3.54 to 1.92) and Fick ($r = 0.54$, bias: 0.02 , 95% CI: -2.41 to 2.44) in the measurement of resting cardiac output in 50 patients with pulmonary hypertension of various aetiologies (Rich et al., 2013). Therefore, previous evidence on the validity and reliability of ICG and similar technologies in precapillary pulmonary hypertension remains limited to small, individual studies with some encouraging but preliminary results.

The reasons behind the lack of accuracy of the ICG in the present population are not clear. As previously stated, the PhysioFlow algorithm should not be confounded by determinants of basal thoracic impedance such as thoracic morphology, fluid and gas content, perspiration, blood resistivity, skin and subcutaneous adiposity thickness and interelectrode distance (Charloux et al., 2000). However, motion artefacts induced by exercise and exaggerated ventilatory responses to exercise might have interfered with the ICG signal (Edmunds et al., 1982; Warburton et al., 1999). Furthermore, measurement of stroke volume in the PhysioFlow algorithm requires calculation of the ventricular ejection time through measurement of the 'thoracic flow inversion time' (Charloux et al., 2000). The latter depends on the pulse pressure and heart rate that are typically pathological in PAH patients; this potential source of error can be amplified by dissociation between electrical and cardiac mechanical events in the lieu of higher pulmonary vascular pressures (Ferreira et al., 2012).

Also, the main pulmonary artery in PAH becomes dilated and frequently exceeds the aortic diameter, which might have introduced an overestimation of the largest impedance variation during systole ($Z_{\text{max}} - Z_{\text{min}}$) during the autocalibration procedure (Ferreira et al., 2012). Finally, very low cardiac output, significant regurgitant volumes, haemodynamically significant cardiac shunts and cardiac arrhythmias may hamper ICG tracings (Bour & Kellett, 2008), but mean cardiac in the present population was preserved and none of the patients had significant cardiac shunt. However, three patients had moderate/severe-to-severe tricuspid regurgitation and four had atrial fibrillation. Whether these factors had collectively accounted for the significant minority of patients with invalid ICG signal (three of 28 patients, 11%) is unknown.

The present study is limited by the small and inhomogeneous population. However, the present population is well representative of patients who undergo investigation for PAH in clinical practice. Also, CMR was not contemporaneous to ICG and TD, but it was performed in a blinded fashion within as little as 72 h to minimize the possibility of significant interval changes in cardiac output; this is supported by the good agreement between CMR and TD.

Conclusion

In a representative population of patients under investigation for PAH, ICG technology showed insufficient correlation and significant overestimation of cardiac output with at least twice as wide limits of agreement compared with TD and CMR. These findings do not support the quantitative and qualitative validity of ICG in the current population. Further research is warranted to allow for the routine use of ICG and similar technologies in the non-invasive assessment of central haemodynamics in PAH.

Acknowledgments

Dr. Marios Panagiotou is the recipient of a European Respiratory Society PAH Long-Term Research fellowship n 2014-3106 supported by an unrestricted grant by GSK.

Conflict of interest

The authors declare no conflict of interest.

Bland JM, Altman DG. Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet* (1986); 1: 307–310.

Bour J, Kellett J. Impedance cardiography: a rapid and cost-effective screening tool for cardiac disease. *Eur J Intern Med* (2008); 19: 399–405.

Charloux A, Lonsdorfer-Wolf E, Richard R, et al. A new impedance cardiograph device for the non-invasive evaluation of cardiac output at rest and during exercise: comparison with the “direct” Fick method. *Eur J Appl Physiol* (2000); 82: 313–320.

D'Alonzo GE, Barst RJ, Ayres SM, et al. Survival in patients with primary pulmonary hypertension. Results from a national prospective registry. *Ann Intern Med* (1991); 115: 343–349.

Edmunds AT, Godfrey S, Tooley M. Cardiac output measured by transthoracic impedance cardiography at rest, during exercise and at various lung volumes. *Clin Sci (Lond)* (1982); 63: 107–113.

Ferreira EM, Ota-Arakaki JS, Barbosa PB, et al. Signal-morphology impedance cardiography during incremental cardiopulmonary exercise testing in pulmonary arterial hypertension. *Clin Physiol Funct Imaging* (2012); 32: 343–352.

Galiè N, Humbert M, Vachiery JL, et al. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: the Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS) Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Respir J* (2015); 46: 903–975.

- Ganz W, Donoso R, Marcus HS, et al. A new technique for measurement of cardiac output by thermodilution in man. *Am J Cardiol* (1971); 27: 392–396.
- Hoepfer MM, Maier R, Tongers J, et al. Determination of cardiac output by the Fick method, thermodilution, and acetylene rebreathing in pulmonary hypertension. *Am J Respir Crit Care Med* (1999); 160: 535–541.
- Humbert M, Sitbon O, Chaouat A, et al. Survival in patients with idiopathic, familial, and anorexigen-associated pulmonary arterial hypertension in the modern management era. *Circulation* (2010); 122: 156–163.
- Hundley WG, Li HF, Hillis LD, et al. Quantitation of cardiac output with velocity-encoded, phase-difference magnetic resonance imaging. *Am J Cardiol* (1995); 75: 1250–1255.
- Mauritz GJ, Marcus JT, Boonstra A, et al. Non-invasive stroke volume assessment in patients with pulmonary arterial hypertension: left-sided data mandatory. *J Cardiovasc Magn Reson* (2008); 10: 51.
- McLaughlin VV, Shillington A, Rich S. Survival in primary pulmonary hypertension: the impact of epoprostenol therapy. *Circulation* (2002); 106: 1477–1482.
- McLaughlin VV, Gaine SP, Howard LS, et al. Treatment goals of pulmonary hypertension. *J Am Coll Cardiol* (2013); 62: D73–D81.
- Nishikawa T, Dohi S. Errors in the measurement of cardiac output by thermodilution. *Can J Anaesth* (1993); 40: 142–153.
- Peacock AJ, Vonk Noordegraaf A. Cardiac magnetic resonance imaging in pulmonary arterial hypertension. *Eur Respir Rev* (2013); 22: 526–534.
- Rich JD, Archer SL, Rich S. Noninvasive cardiac output measurements in patients with pulmonary hypertension. *Eur Respir J* (2013); 42: 125–133.
- Richard R, Lonsdorfer-Wolf E, Charloux A, et al. Non-invasive cardiac output evaluation during a maximal progressive exercise test, using a new impedance cardiograph device. *Eur J Appl Physiol* (2001); 85: 202–207.
- Stevens JH, Raffin TA, Mihm FG, et al. Thermodilution cardiac output measurement. Effects of the respiratory cycle on its reproducibility. *JAMA* (1985); 253: 2240–2242.
- Tan KH, Lai FO, Hwang NC. Measurement of cardiac output using Physio Flow with different positions of electrode placement. *Singapore Med J* (2006); 47: 967–970.
- Tang WH, Tong W. Measuring impedance in congestive heart failure: current options and clinical applications. *Am Heart J* (2009); 157: 402–411.
- Taniguchi Y, Emoto N, Miyagawa K, et al. Noninvasive and simple assessment of cardiac output and pulmonary vascular resistance with whole-body impedance cardiography is useful for monitoring patients with pulmonary hypertension. *Circ J* (2013); 77: 2383–2389.
- Tonelli AR, Alnuaimat H, Li N, et al. Value of impedance cardiography in patients studied for pulmonary hypertension. *Lung* (2011); 189: 369–375.
- Tonelli AR, Alkukhun L, Arelli V, et al. Value of impedance cardiography during 6-minute walk test in pulmonary hypertension. *Clin Transl Sci* (2013); 6: 474–480.

Warburton DE, Haykowsky MJ, Quinney HA, et al. Reliability and validity of measures of cardiac output during incremental to maximal aerobic exercise. Part II: novel techniques and new advances. *Sports Med* (1999); 27: 241–260.

Yung GL, Fedullo PF, Kinninger K, et al. Comparison of impedance cardiography to direct Fick and thermodilution cardiac output determination in pulmonary arterial hypertension. *Congest Heart Fail* (2004); 10: 7–10.

Table 1 Clinical patient characteristics^a

Characteristics	ICG versus TD at rest (n = 25)	ICG versus TD versus CMR at rest (n = 16)	ICG versus TD at exercise (n = 16)
Age, year	63.8 ± 11.9	59.6 ± 11.5	62.1 ± 12.3
Sex (male:female)	13:12	8:8	8:8
BMI, kg m ⁻²	28.1 ± 5.5	28.4 ± 5.7	28.1 ± 5.3
WHO FC, II:III:IV	5:17:3	4:11:1	4:12:0
6MWD, m	268.3 ± 134.8	285.1 ± 140.2	325.9 ± 124.4
Diagnosis			
IPAH	7	5	4
CTD	5	3	4
PoPH	2	2	1
PH due to left heart disease	3	1	2
PH due to lung respiratory diseases and/or hypoxia	4	2	1
CTEPH	1	1	1
No PH	3	2	3

ICG, impedance cardiography; TD, thermodilution; CMR, cardiac magnetic resonance; BMI, body mass index; WHO FC, World Health Organization functional class; 6MWT, six-minute walk distance; IPAH, idiopathic pulmonary arterial hypertension; CTD, connective tissue disease-associated PAH; PoPH, portopulmonary arterial hypertension; PH, pulmonary hypertension; CTEPH, chronic thromboembolic pulmonary hypertension.

^aData are presented as means ± SD.

Table 2 Physiological patient characteristics at right heart catheterization^a

Characteristics	ICG versus TD at rest (n = 25)	ICG versus TD versus CMR at rest (n = 16)	ICG versus TD at exercise (n = 16)
mean PAP, mm Hg	35.3 ± 12.3	35.2 ± 12.4	Rest: 31.9 ± 10.9 Exercise: 52.4 ± 17.6
PAOP, mm Hg	6.5 ± 5.2	6.8 ± 6.3	Rest: 5.5 ± 3.5 Exercise: 11.6 ± 6.7
CO, l min ⁻¹	4.7 ± 1.7	4.8 ± 1.9	Rest: 5.2 ± 1.8 Exercise: 11.9 ± 3.1
CI, l min ⁻¹ m ⁻²	2.5 ± 0.9	2.6 ± 1.0	Rest: 2.7 ± 0.9 Exercise: 4.2 ± 1.6
PVR, Wood units	7.4 ± 6.2	7.3 ± 6.9	Rest: 5.7 ± 3.7 Exercise: 5.6 ± 4.2

PAP, pulmonary artery pressure; PAOP, pulmonary artery occlusion pressure; CO, cardiac output; CI, cardiac index; TD, Thermodilution; PVR, pulmonary vascular resistance; CMR, cardiac magnetic resonance.

^aData are presented as means ± SD.

Table 3 Numerical values, Pearson's correlation coefficient (r) and Bland–Altman analyses for cardiac output^a

	Modality (n)	Mean CO, l min ⁻¹	Correlation coefficient (P value)	Bias, l min ⁻¹	Lower and upper limits, l min ⁻¹
CO _{ICG} versus CO _{TD}	Rest (25)	5.93 ± 1.66 versus 4.72 ± 1.70	0.42 (0.035)	1.21	-2.33 to 4.75
CO _{ICG} versus CO _{CMR}	Rest (16)	6.40 ± 1.78 versus 5.01 ± 1.77	0.38 (0.1)	1.40	-2.48 to 5.28
CO _{ICG} versus CO _{TD}	Exercise (16)	9.31 ± 3.44 versus 7.91 ± 3.07	0.65 (0.007)	1.41	-3.99 to 6.81
DCO _{ICG} versus DCO _{TD}	Rest to exercise (16)	3.51 ± 2.69 versus 2.75 ± 1.77	0.53 (0.033)	0.76	-3.74 to 5.26
CO _{TD} versus CO _{CMR}	Rest (16)	4.85 ± 1.88 versus 5.01 ± 1.77	0.87 (<0.001)	-0.16	-1.97 to 1.65

CO, cardiac output; CO_{ICG}, impedance cardiography-measured cardiac output; CO_{TD}, thermodilution-measured cardiac output; CO_{CMR}, cardiac magnetic resonance imaging-measured cardiac output; DCO_{ICG} and DCO_{TD}, changes in CO_{ICG} and CO_{TD} from rest to exercise.

^aData are presented as means ± standard deviation.

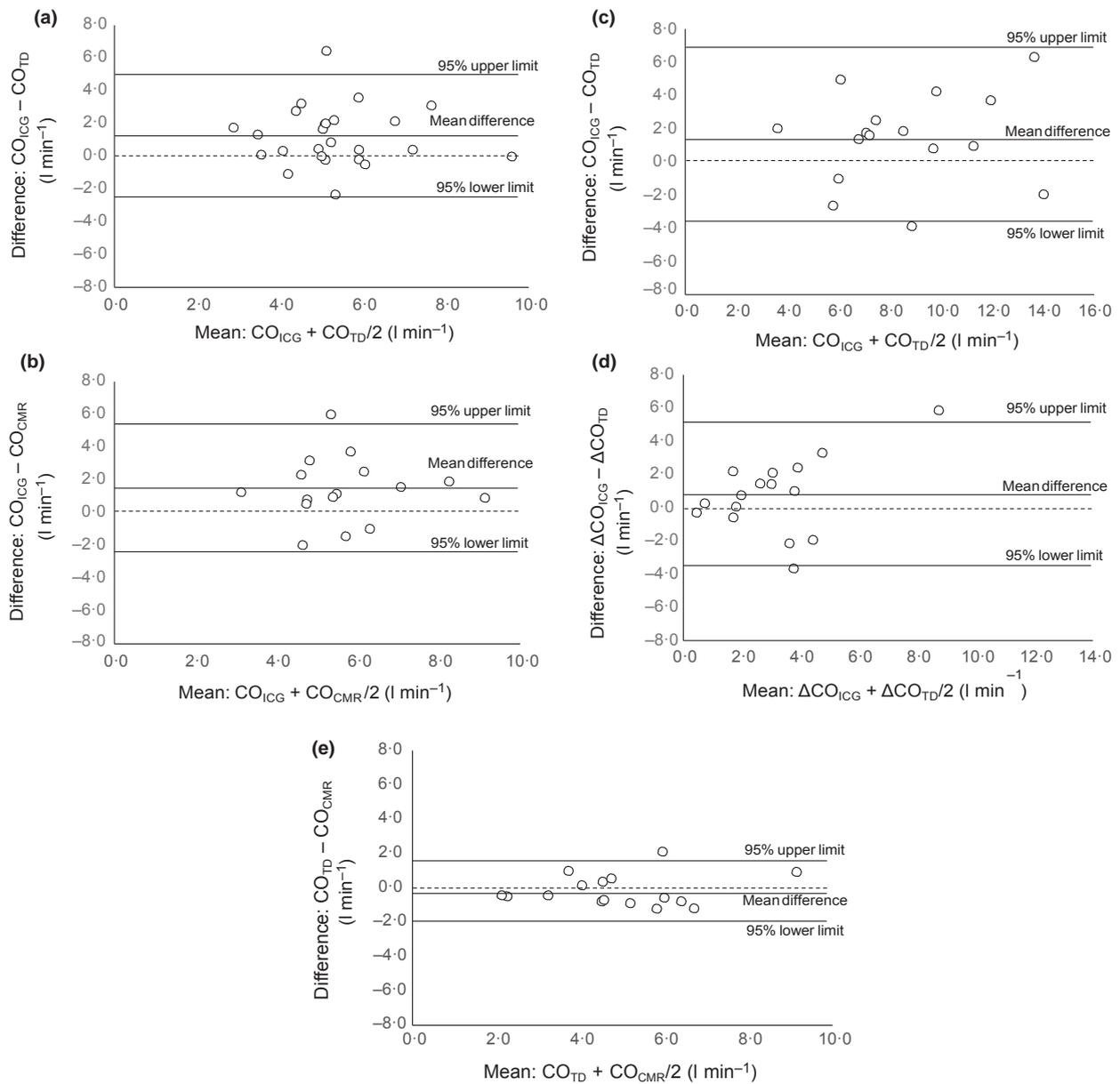


Figure 1 Bland–Altman plots comparing cardiac output measured by impedance cardiography (CO_{ICG}), thermodilution (CO_{TD}) and cardiac magnetic resonance (CO_{CMR}): (a) CO_{ICG} versus CO_{TD} at rest; (b) CO_{ICG} versus CO_{CMR} at rest; (c) CO_{ICG} versus CO_{TD} at exercise; (d) change in CO_{ICG} versus change in CO_{TD} from rest to exercise; (e) CO_{TD} versus CO_{CMR} at rest.