

Northumbria Research Link

Citation: Wong, Max and Lodge, John (2012) A metabolomic investigation of the effects of vitamin E supplementation in humans. *Nutrition & Metabolism*, 9 (1). p. 110. ISSN 1743-7075

Published by: BioMed Central

URL: <http://dx.doi.org/10.1186/1743-7075-9-110> <<http://dx.doi.org/10.1186/1743-7075-9-110>>

This version was downloaded from Northumbria Research Link:
<https://nrl.northumbria.ac.uk/id/eprint/12113/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)

RESEARCH

Open Access

A metabolomic investigation of the effects of vitamin E supplementation in humans

Max Wong^{1,3} and John K Lodge^{2,3*}

Abstract

Background: Vitamin E is a nutrient with both antioxidant and non-antioxidant activities and has been shown to modulate the function of a number of cell types *in vitro* and in human studies. However studies have also shown vitamin E to have detrimental interactions and therefore it is important to establish the extent to which this nutrient influences metabolism. Metabolomics can potentially identify nutrient-metabolism interactions and therefore the aim of this study was to use a non-targeted metabolomic approach to identify changes to the plasma metabolome following vitamin E supplementation in humans.

Methods: A relatively homogenous healthy adult male population ($n = 10$) provided a fasting blood sample immediately before and after a 4-week vitamin E supplementation regime (400 mg/d of *RRR*- α -tocopheryl acetate) on top of their habitual diet. Plasma samples were analysed for vitamin E and clinical markers. Plasma underwent non-targeted metabolite profiling using liquid chromatography/mass spectroscopy and data was processed using multivariate statistical analysis.

Results: Plasma vitamin E concentrations were significantly increased following supplementation ($p < 0.001$). A partial least squares-discriminant analysis (PLS-DA) model was able to discriminate between samples taken pre and post vitamin E supplementation (goodness of fit $R^2Y = 0.82$, predictive ability $Q^2 = 0.50$). Variable influence on projection and PLS-DA loadings highlighted a number of discriminating ions that were confirmed as discriminatory through pairwise analysis. From database searches and comparison with standards these metabolites included a number of lysophosphatidylcholine species (16:0, 18:0, 18:1, 18:2, 20:3 and 22:6) that were increased in intensity post supplementation by varying degrees from 4% to 29% with the greatest changes found for lysoPC 22:6 and 20:3.

Conclusions: Although a small scale study, these results potentially indicate that vitamin E supplementation influences phospholipid metabolism and induces lysoPC generation; a general pro-inflammatory response. Moreover the study identifies novel areas of vitamin E interactions and highlights the potential of metabolomics for elucidating interactions between nutrients and metabolic pathways in nutritional research.

Keywords: Metabolite profiling, LC/MS, Vitamin E, Lysophospholipids

Background

It is well established that nutrients interact with metabolic pathways at various levels and have multiple targets. With the increasing use of '-omic' technologies in nutritional research [1] it is now becoming possible to further explore the influence of single nutrients or dietary change on physiological processes.

Vitamin E has been widely researched owing to its antioxidant [2] and non-antioxidant functions [3] and its potential as a cardioprotective agent. Vitamin E appears to have wide ranging effects on cellular systems with the ability to inhibit protein kinase C, activating diacylglycerol kinase (decreasing diacylglycerol availability) and protein phosphatase 2A [4], regulating specific gene expression [5,6]; in this way vitamin E can influence a number of biological functions and metabolism. Interestingly, α -TTP knock out mice (model of vitamin E deficiency) had lower glucose levels, improved glucose tolerance and increased insulin excretion [7] indicating a

* Correspondence: john.lodge@northumbria.ac.uk

²School of Life Sciences, Northumbria University, Ellison Building, Newcastle-Upon-Tyne, Tyne & Wear NE1 8ST, UK

³Faculty of Health and Medical Sciences, University of Surrey, Guildford, Surrey GU2 7XH, UK

Full list of author information is available at the end of the article

role for α -tocopherol regulation in glucose control. Vitamin E supplementation has been shown to influence platelet [8] and mononuclear cell functions [8,9], and reduce indices of oxidative stress and inflammation [10] in humans. However vitamin E may also have potential detrimental actions as high dose (>400 mg/d) supplementation was found to increase mortality in a meta-analysis [11] and high dose supplementation has been proposed to potentially interfere with drug metabolism [12]. As nutrients generally have multiple targets and interactions, it is likely that vitamin E has alternate functions and can further influence metabolic pathways.

Metabolomics offers the opportunity to investigate global changes to metabolites in biological samples and there is much interest in this technology for nutritional studies [1,13]. Potential opportunities include a further understanding of the interactions of whole diet or individual nutrients on metabolic pathways, the metabolic interactions between diet and disease, and developing biomarkers for dietary exposure and/or disease [14,15]. Metabolomic technology has been applied to a number of nutritional studies (reviewed in [1,16]), allowing investigators to compare the influence of dietary patterns on metabolite profiles [17], monitor nutrient metabolism or identify novel metabolic pathways influenced by the nutrient [18]. Interestingly metabolomics has previously been used in animal models to investigate vitamin E metabolism. Following pregnane X receptor activation in knockout mice there was an attenuation of α -tocopherol metabolism but production of a novel γ -tocopherol metabolite [19]. Novel vitamin E metabolites were also found by Johnson et al. in human and mouse models by LC/MS [20]. In a mouse model of neuronal ceroid lipofuscinosis, vitamin E supplementation reversed metabolic abnormalities associated with the phenotype [21]. In a rat liver model of vitamin E deficiency, the authors found metabolic changes associated with amino acids, glucose and purines that suggested shifts in energy metabolism with deficiency [22]. In the only human metabolic study involving vitamin E, metabolomics was successfully used to determine 'predictors of response' to vitamin E treatment in subjects with Non-Alcoholic Steatohepatitis [23]. These studies demonstrate that vitamin E can influence metabolic pathways and so it is of interest to investigate other potential interactions in a human supplementation study.

We have previously highlighted that metabolomics can be used to provide insights into how vitamin E can influence the metabolome [16] and in the present study we have expanded on these observations to identify subtle but distinct changes to human plasma metabolome following vitamin E supplementation, the main effects of which include significant increases in the intensities of a number of lysophospholipid species.

Methods

Materials

All solvents were purchased from Fisher Scientific (Loughborough, UK). Acetonitrile and methanol were LC/MS grade, formic acid was laboratory reagent grade. Water was purified using an Elga PureLab Ultra system (Elga, High Wycombe, UK). Leucine enkephalin and lysophosphatidylcholine pure standards (analytical grade) were purchased from Sigma Aldrich (Poole, UK). Gelatine capsules containing 400 mg of natural (*RRR*) α -tocopheryl acetate (purity 98.9%) in vitamin E-stripped corn oil were purchased from Eurocaps Ltd. (Gwent, UK).

Human study

For this study 10 male subjects were recruited from within the University of Surrey and surrounding area through advertisement. Their average age and BMI were 32 ± 9 years and 24 ± 4 kg/m² respectively. The subjects were outwardly healthy, normolipidaemic (total cholesterol 4.76 ± 1.3 mmol/L and triacylglycerols 0.95 ± 0.6 mmol/L) and were not taking any prescription medication or dietary supplements. The study was a one-way supplementation trial with subjects taking a capsule containing 400 mg of α -tocopheryl acetate every day for 4 weeks on top of their habitual diet (no dietary standardisation was performed). At the start and end of the trial, subjects visited the clinical investigation unit at the University of Surrey in a fasted state where they provided a blood sample (20 mL, EDTA). Under identical conditions for each collection, plasma was harvested by centrifugation, immediately aliquoted (0.5 mL), frozen and stored at -80°C prior to analysis. The University of Surrey Ethics Committee granted approval for this study.

Sample preparation

For protein precipitation, 100 μL of freshly thawed human plasma was vortex mixed with 400 μL of chilled (-80°C) methanol for 1 min then left on ice for 10 min. The mixture was then centrifuged at 13,000 rpm for 10 minutes and the supernatant transferred to autosampler vials (Waters, Manchester, UK) for LC/MS analysis. For metabolite identification, pure standards (1 ng/ μL injected) were dissolved in methanol and analysed under the same conditions as for the plasma samples. Plasma samples were also spiked with pure standards.

Liquid Chromatography Mass Spectrometry (LC/MS)

Chromatography was performed on a Waters Acquity UPLCTM system (Waters, Manchester, UK) using an Acquity BEH C₁₈ column (1.7 μm 2.1 \times 100 mm) for sample separation following a 5 μL injection volume using partial loop mode. The column oven was maintained at 40°C and the autosampler at 4°C . The mobile

phase consisted of: (A) 0.1% formic acid in water, and (B) acetonitrile with 0.1% formic acid. The gradient program began with 100% (A) for 0.5 min, then proceeded with a linear gradient to 100% (B) over 8.5 min, then returned to initial conditions (100% A) and maintained for 1 min. The total run time was 10 min with a flow rate of 0.6 ml/min.

Mass Spectrometry was performed on a Waters Micromass QToF Premier™ (Waters, Manchester, UK) operating in both positive and negative ion electrospray modes. The source temperature was set to 120°C, the desolvation temperature was 450°C, with nebulisation gas set to 800 L/hr. The collision energy was set at 4 eV. The capillary and cone voltages were set at 3.1 kV and 38 V respectively in positive ionisation mode, and 2.9 kV and 35 V respectively in negative ionisation mode. A LockSpray™ interface was used to ensure mass accuracy. For this leucine-enkephalin (556.2771 m/z in positive ionisation mode) was infused at a concentration of 200 µg/µL at a flow rate of 0.01 ml/min. Data was collected in centroid mode over the range 100 to 1000 m/z with an acquisition rate of 0.2 s, interscan delay of 0.02 s, with dynamic range enhancement activated. MS/MS experiments were performed by ramping the collision energy from 10–50 V on selected masses of interest while all other parameters remained the same. Each sample was injected 6 times. The sample list was randomised and also contained a quality control sample and a metabolite test mix sample which were analysed following every 20 injections in order to monitor retention time and mass drift within the total analytical period.

Other analysis

Plasma vitamin E concentrations were analysed by HPLC incorporating electrochemical detection as previously described [24]. Plasma total cholesterol, triacylglycerols, glucose and nonesterified fatty acids were measured using enzymatic test kits (Instrumentation Laboratory, UK) on an automated clinical analyser (ILab650, Instrumentation Laboratory, UK).

Data analysis

The acquired LC/MS data was analysed and visualised using MassLynx and MarkerLynx (both Version 4.1, Waters), using a noise elimination level set at 6 and 15 masses per retention time. Background ions from associated blank injections were excluded from each sample. The mass and retention time windows were 0.05 Da and 0.1 min respectively and signal drift was corrected automatically by the software. The peak information was then transferred to a multivariate analysis package (SIMCA-P+, Ver 12, Umetrics AB, Sweden). Data were initially analysed by Principal Component Analysis (PCA) to identify any outliers. All variables were set to

Pareto scaling. A 2-component model was generated for Partial Least Squares Discriminant Analysis (PLS-DA) and the predictive ability (Q^2) of the model assessed by internal cross validation (seven-group venetian blind cross-validation). A predictive set was established by randomly removing 30% of the data to form a training set and testing the predictive ability of the model against the remaining 'test' data through averaging seven iterations of the data. The model was able to correctly classify 72% of samples (in both groups) in a predictive set. PLS-DA loadings and variable influence on projection (VIP) plots were used to identify the most important discriminatory species from the total list of features detected. For the pairwise analysis of signal intensities, Wilcoxon Matched Pairs tests were performed using Prism (Ver 5, GraphPad Software Inc). A Bonferroni correction, to take into account multiple comparisons, was applied using the number of discriminating ions from the VIP plot as the number of variables in the calculation, giving an alpha level of 0.002. Only signals that were significantly different with treatment ($p < 0.05$), or a trend for significance ($p < 0.1$) following pair-wise comparisons, underwent putative identification.

Results and discussion

In this study LC/MS based metabolomics has been applied to plasma samples obtained pre and post vitamin E supplementation. Metabolomics is being increasingly used in nutritional studies [16], as it is of importance to further understand how diet influences metabolic pathways and to develop biological markers of dietary exposure [14]. Vitamin E supplementation has been shown to have varied responses in a number of *ex vivo* and *in vitro* studies and with the development of more sophisticated research tools such as metabolomics, our aim was to use this technology to identify any effects of vitamin E supplementation on human metabolism. Vitamin E plasma concentrations were relatively high at baseline, suggesting the subjects were replete in vitamin E, but values significantly increased from 31.53 ± 4.6 µmol/L before supplementation to 46.65 ± 5.16 µmol/L post supplementation ($p < 0.001$), which represents a 1.5 fold increase in concentration. This is in line with what is expected from similar vitamin E interventions [25] as it is known that due to hepatic regulation plasma concentrations become saturated and cannot be raised more than 2–3 fold [26]. There were no changes to plasma concentrations of glucose, total cholesterol and triacylglycerol following supplementation (data not shown).

Classification of samples

The LC/MS approach detected approximately 2500 ion species in plasma samples, and there was no significant

difference between the total number of species detected pre (2551 ± 176) and post (2556 ± 142) supplementation. The data were firstly analysed by unsupervised Principal Component Analysis (PCA) to check for overall data quality (Figure 1a). We found no data points outside the Hotelling confidence intervals and so no data points were investigated further. PCA was unable to discriminate between sample groups (Figure 1a), however this is not uncommon as PCA may not be sensitive enough to discriminate between samples sets when there is large variation and only modest differences between groups, as commonly seen in nutritional studies [27].

To further explore the data a two-component PLS-DA model was established (Figure 1b), and was able to discriminate between treatment (model parameters; $R^2Y = 0.82$, $Q^2 = 0.50$), with samples post supplementation clustering mainly to the left along the first latent variable axis. To identify discriminatory species the data was explored through a PLS-DA loadings plot and a VIP plot of the first latent variable (Figure 2a and b respectively). This generated a list of top discriminatory species above a VIP score of 7.5. We then analysed single ion chromatograms and associated molecular ions of the species in the original data in order to check for adducts and remove false positive signals, and performed pairwise comparisons of ion intensities between sample class to confirm differences between treatment groups. From this analysis out of the approximately 2500 features initially detected, only seven discriminatory signals remained for further identification.

Identification of signals discriminatory for vitamin E supplementation

To identify these discriminatory species, elemental composition analysis was performed using Masslynx to

obtain potential formulae. Masses were then entered into metabolite databases (Human Metabolite Database, www.hmdb.ca; METLIN, metlin.scripps.edu), and formulae compared with possible elemental composition taking into account potential adducts from molecular ion analysis. To confirm identities pure compounds were purchased when possible and analysed under identical conditions to that of the plasma samples, including spiking of original samples. Molecular ion and fragmentation patterns of the standards and the associated peak in original samples, retention time and sample spiking confirmed the assignments. An example of such an assignment strategy is shown in Figure 3 for the species at 7.78 min, 524.37 m/z in plasma. A positive ion mode TIC of the region around the peak in a representative plasma sample is shown in Figure 3a. A spectrum of the peak at 7.78 min in positive mode (Figure 3b) revealed a major signal at 524.372 m/z and in negative mode (Figure 3c) at 568.36 and 508.34 m/z. The top match for this signal from metabolite databases was found to be the $[M + H]^+$ adduct of lysophosphatidylcholine (C18:0). MS/MS fragmentation patterns of the peak at 7.78 min in positive mode (Figure 3d) revealed major signals at 184.07 and 104.10 m/z characteristic of the phosphocholine moiety [28] and in negative mode (Figure 3e) at 283.25 m/z characteristic of the $[M - H]^-$ adduct of stearic acid. A comparison of this analysis with that of a pure standard of lysophosphatidylcholine (C18:0) (Figure 3f-j) revealed identical mass spectroscopy behaviour. Using the same approach, six lysophosphatidylcholine (LysoPC) species were assigned (Table 1) that were increased in intensity post supplementation (pairwise analysis, Figure. 4). Similar assignment strategies were used for the confirmation of LysoPC in previous studies [29,30].

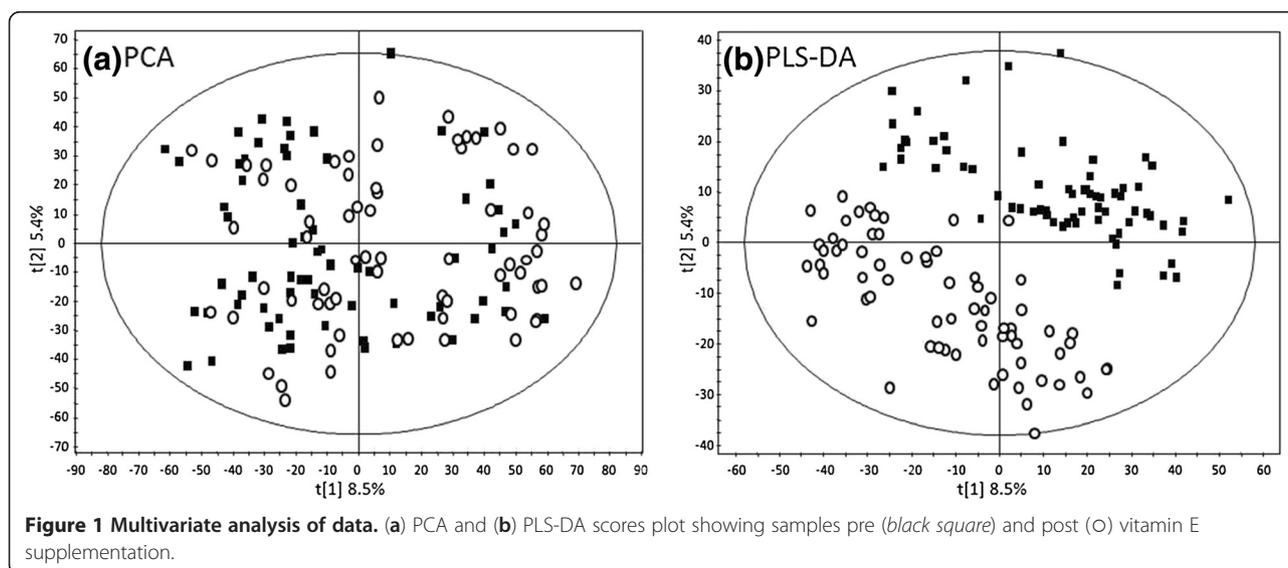
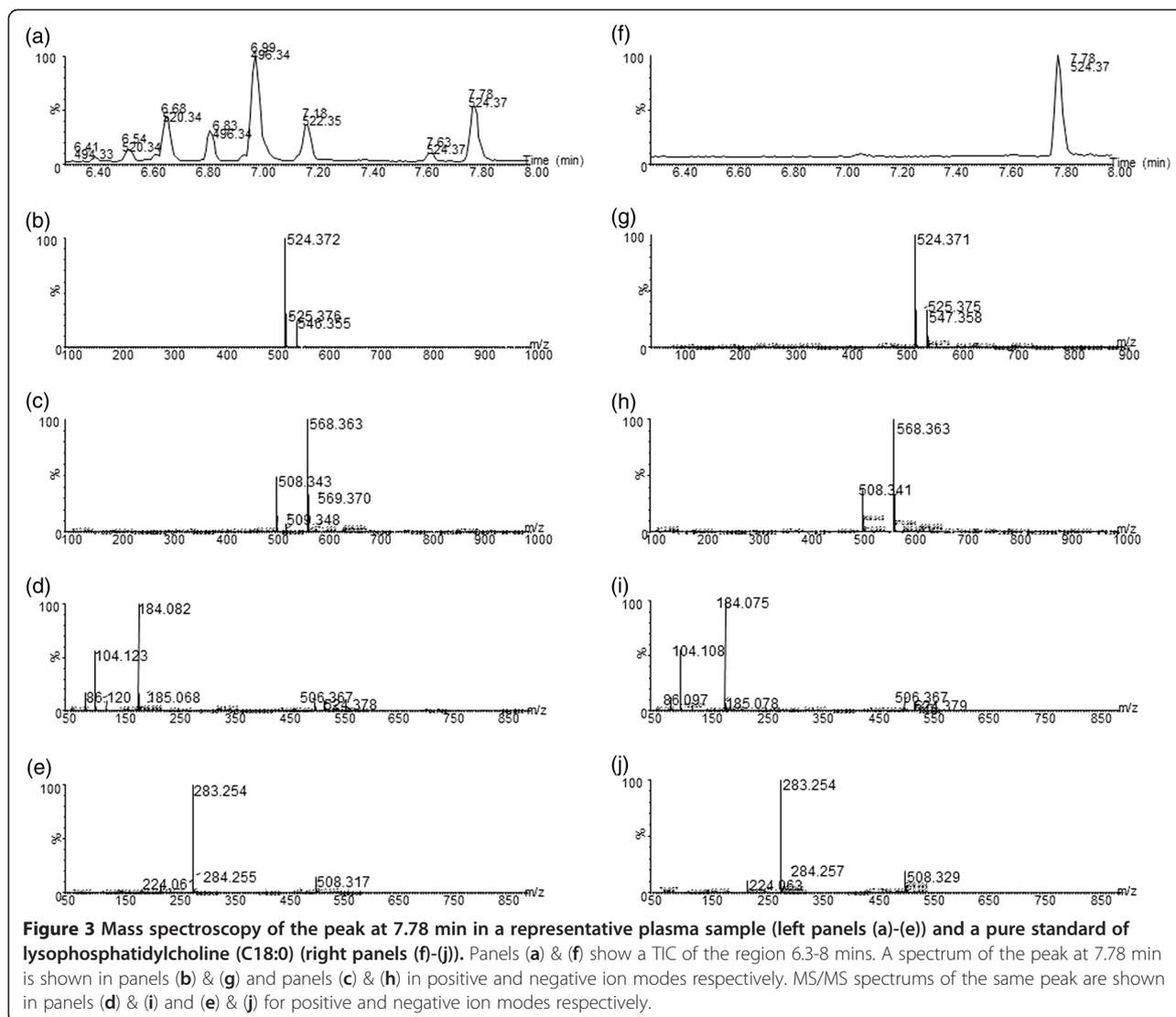


Figure 1 Multivariate analysis of data. (a) PCA and (b) PLS-DA scores plot showing samples pre (black square) and post (O) vitamin E supplementation.



2 diabetes [40] does link these various lines of evidence and warrants further study.

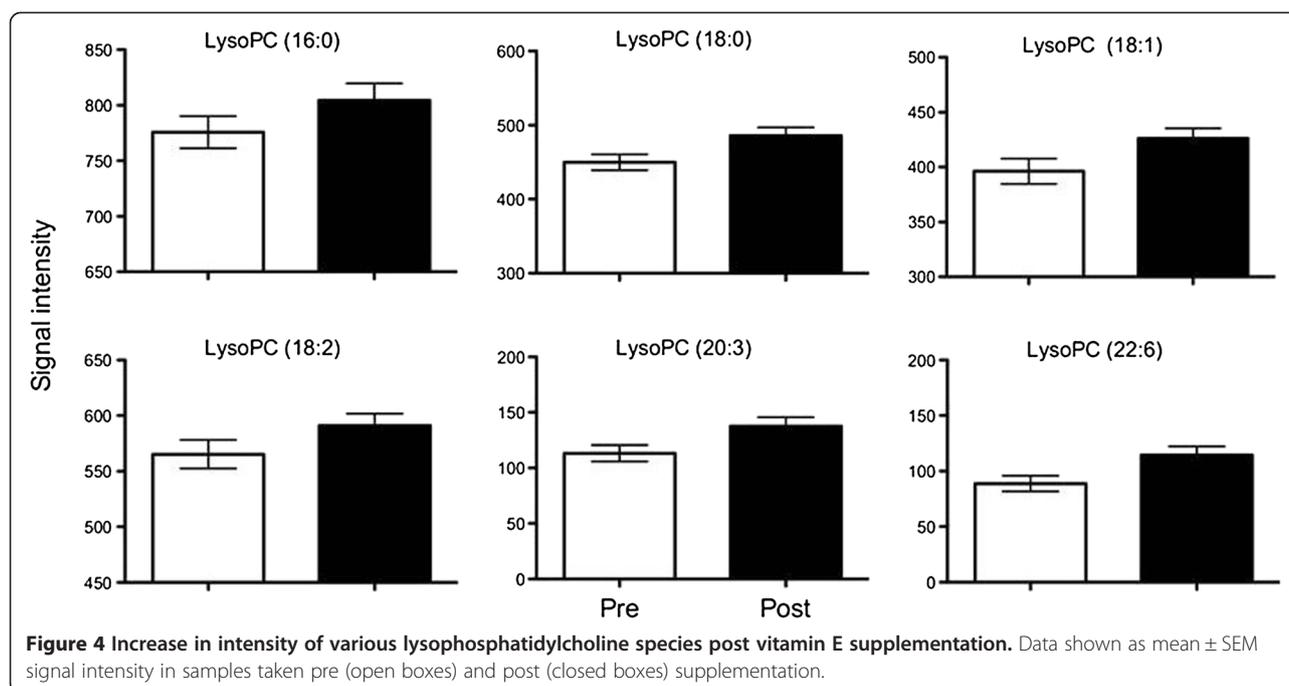
Potential mechanisms of raised lysophosphatidylcholines

There are a number of potential mechanisms by which vitamin E supplementation could increase levels of lysoPC. Vitamin E has been shown to influence gene expression of a number of systems and modulate the activities of a number of enzymes involved in signal transduction, including PLA₂ [41]. Depending on the model used vitamin E treatment has been shown to activate [37,42,43], inhibit [44-46] or have no effect [47] on PLA₂ activity, but the weight of evidence overall is suggestive of an increase in PLA₂ activity following vitamin E exposure in cell/animal models. Because of the difference in models used and lack of human data it is not possible to relate PLA₂ activity with vitamin E levels. Vitamin E can form complexes with lysoPC which help

Table 1 Major discriminatory species identified from the PLS-DA loadings and VIP plot

VIP	<i>m/z</i> [M + H] ⁺	RT (min)	<i>P</i> value*	Assignment
18.02	524.371	7.78	0.037	LysoPC (18:0)
17.93	520.340	6.60	0.052	LysoPC (18:2)
16.02	496.339	6.93	0.085	LysoPC (16:0)
15.67	522.355	7.11	0.045	LysoPC (18:1)
13.32	546.356	6.88	0.018	LysoPC (20:3)
12.73	568.340	6.56	0.023	LysoPC (22:6)
9.59	235.181	2.61	0.012	NA

* from pairwise comparisons pre vs post, with an adjusted Bonferroni *p* value (<0.002) no significance remained. NA, no assignment from metabolite databases. All signals were increased in intensity post vitamin E supplementation.



to maintain membrane structure and although vitamin E is thought to stabilise membranes through its interaction with phospholipids and other membrane components, there is evidence that it can also destabilise [48]. An increase in vitamin E content of biological membranes, following supplementation, may potentially disrupt the membrane causing displacement of membrane phospholipids, which are metabolised releasing lysoPC. However it is not known if a 1.5 fold increase in vitamin E content (as found in the current study) could have these effects on biological membranes. Alternatively the result may not necessarily be specific to vitamin E but could be representative of an acute phase response to the intervention, or a similar phenomenon, as other studies have found lysoPC to be discriminating species. For example, a metabolomic study found a number of lysoPC species increased during an oral glucose tolerance test [18]. LysoPC have also been associated with a number of disease states and found to be increased in subjects with hepatitis B-induced deterioration of liver function [29], rheumatoid arthritis [49], type 2 diabetes [50] and obesity [51], whereas lysoPC were significantly decreased in subjects with colorectal cancer [52], and in sepsis patients [53]. Therefore, as lysoPC appear to be influenced by a number of physiological conditions further work is necessary to elucidate the mechanism of this effect and the identification of other species influenced by vitamin E supplementation. It would be of interest to pursue and validate these observations in a more comprehensive study of vitamin E supplementation.

Conclusions

Overall the data in this study suggest that vitamin E supplementation influences phospholipid metabolism, increasing the intensity of a variety of lysophosphatidylcholine species in plasma in the order 22:6 > 20:3 > 18:0 ~ 18:1 > 18:2 ~ 16:0. This is one of only a few reports using metabolomics to investigate single nutrient interactions in a human intervention study and highlights the potential of metabolomic technology in nutrition research in defining alternative functions for nutrients and elucidating how nutrients interact with metabolic pathways, in this case vitamin E.

Abbreviations

BMI: Body mass index; EDTA: Ethylene diamine tetra acetic acid; HPLC: High pressure liquid chromatography; LC/MS: Liquid Chromatography Mass Spectroscopy; PCA: Principal component analysis; PLS-DA: Partial least squares-discriminant analysis; lysoPC: Lysophosphatidylcholine; Q^2 : Model predictive ability; QTOF: Quadrupole time of flight spectrometer; R^2Y : Model goodness of fit; VIP: Variable influence on projection.

Competing interests

The authors declare no competing interest.

Authors' contributions

JL conceived and designed the study and prepared the manuscript. MW carried out all the experimental work and statistical analysis and helped to draft the manuscript. All authors read and approved the final manuscript.

Acknowledgments

We would like to thank the subjects who took part in this study and to the undergraduate project students that helped co-ordinate the human trial. We thank the University of Surrey for pump-priming funding for this study.

Author details

¹Department of Paediatrics, University of Cambridge, Addenbrooke's Hospital, Cambridge CB2 0QQ, UK. ²School of Life Sciences, Northumbria

University, Ellison Building, Newcastle-Upon-Tyne, Tyne & Wear NE1 8ST, UK.
³Faculty of Health and Medical Sciences, University of Surrey, Guildford, Surrey GU2 7XH, UK.

Received: 17 October 2012 Accepted: 15 December 2012
Published: 19 December 2012

References

1. Rezzi S, Ramadan Z, Fay LB, Kochhar S: **Nutritional metabonomics: applications and perspectives.** *J Proteome Res* 2007, **6**:513–525.
2. Burton GW, Traber MG: **Vitamin E: antioxidant activity, biokinetics and bioavailability.** *Annu Rev Nutr* 1990, **10**:357–382.
3. Rimbach G, Minihane AM, Majewicz J, Fischer A, Pallauf J, Virgli F, Weinberg PD: **Regulation of cell signalling by vitamin E.** *Proc Nutr Soc* 2002, **61**:415–425.
4. Azzi A, Breyer I, Feher M, Pastori M, Ricciarelli R, Spycher S, Staffieri M, Stocker A, Zimmer S, Zingg JM: **Specific cellular responses to alpha-tocopherol.** *J Nutr* 2000, **130**:1649–1652.
5. Rimbach G, Moehring J, Huebber P, Lodge JK: **Gene-regulatory activity of alpha-tocopherol.** *Molecules* 2010, **15**:1746–1761.
6. Azzi A, Gysin R, Kempna P, Munteanu A, Villacorta L, Visarius T, Zingg JM: **Regulation of gene expression by alpha-tocopherol.** *Biol Chem* 2004, **385**:585–591.
7. Birringer M, Kuhlow D, Pfluger PT, Landes N, Schulz TJ, Glaubitz M, Florian S, Pfeiffer A, Schuelke M, Brigelius-Flohe R, Ristow M: **Improved glucose metabolism in mice lacking alpha-tocopherol transfer protein.** *Eur J Nutr* 2007, **46**:397–405.
8. Williams JC, Forster LA, Tull SP, Ferns GA: **Effects of vitamin E on human platelet and mononuclear cell responses in vitro.** *Int J Exp Pathol* 1999, **80**:227–234.
9. Devaraj S, Li D, Jialal I: **The effects of alpha-tocopherol supplementation on monocyte function. Decreased lipid oxidation, interleukin 1 beta secretion and monocyte adhesion to endothelium.** *J Clin Invest* 1996, **98**:756–763.
10. Devaraj S, Tang R, Adams-Huet B, Harris A, Seenivasan T, de Lemos JA, Jialal I: **Effect of high-dose alpha-tocopherol supplementation on biomarkers of oxidative stress and inflammation and carotid atherosclerosis in patients with coronary artery disease.** *Am J Clin Nutr* 2007, **86**:1392–1398.
11. Miller ER 3rd, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, Guallar E: **Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality.** *Ann Intern Med* 2005, **142**:37–46.
12. Brigelius-Flohe R: **Adverse effects of vitamin E by induction of drug metabolism.** *Genes Nutr* 2007, **2**:249–256.
13. Gibney MJ, Walsh M, Brennan L, Roche HM, German B, van Ommen B: **Metabolomics in human nutrition: opportunities and challenges.** *Am J Clin Nutr* 2005, **82**:497–503.
14. Lodge JK: **Symposium 2: modern approaches to nutritional research challenges: targeted and non-targeted approaches for metabolite profiling in nutritional research.** *Proc Nutr Soc* 2010, **69**:95–102.
15. Fave G, Beckmann ME, Draper JH, Mathers JC: **Measurement of dietary exposure: a challenging problem which may be overcome thanks to metabolomics?** *Genes Nutr* 2009, **4**:135–141.
16. Primrose S, Draper J, Elsom R, Kirkpatrick V, Mathers JC, Seal C, Beckmann M, Halldar S, Beattie JH, Lodge JK, et al: **Metabolomics and human nutrition.** *Br J Nutr* 2011, **105**:1277–1283.
17. Stella C, Beckwith-Hall B, Cloarec O, Holmes E, Lindon JC, Powell J, van der Ouderaa F, Bingham S, Cross AJ, Nicholson JK: **Susceptibility of human metabolic phenotypes to dietary modulation.** *J Proteome Res* 2006, **5**:2780–2788.
18. Zhao X, Peter A, Fritsche J, Elcnerova M, Fritsche A, Haring HU, Schleicher ED, Xu G, Lehmann R: **Changes of the plasma metabolome during an oral glucose tolerance test: is there more than glucose to look at?** *Am J Physiol Endocrinol Metab* 2009, **296**:E384–393.
19. Cho JY, Kang DW, Ma X, Ahn SH, Krausz KW, Luecke H, Idle JR, Gonzalez FJ: **Metabolomics reveals a novel vitamin E metabolite and attenuated vitamin E metabolism upon PXR activation.** *J Lipid Res* 2009, **50**:924–937.
20. Johnson CH, Slanar O, Krausz KW, Kang DW, Patterson AD, Kim JH, Luecke H, Gonzalez FJ, Idle JR: **Novel metabolites and roles for alpha-tocopherol in humans and mice discovered by mass spectrometry-based metabolomics.** *Am J Clin Nutr* 2012, **96**:818–830.
21. Griffin JL, Muller D, Woograsingh R, Jowatt V, Hindmarsh A, Nicholson JK, Martin JE: **Vitamin E deficiency and metabolic deficits in neuronal ceroid lipofuscinosis described by bioinformatics.** *Physiol Genomics* 2002, **11**:195–203.
22. Moazzami AA, Andersson R, Kamal-Eldin A: **Changes in the metabolic profile of rat liver after alpha-tocopherol deficiency as revealed by metabolomics analysis.** *NMR Biomed* 2011, **24**:499–505.
23. Cheng J, Joyce A, Yates K, Aouizerat B, Sanyal AJ: **Metabolomic profiling to identify predictors of response to vitamin E for Non-Alcoholic Steatohepatitis (NASH).** *PLoS One* 2012, **7**:e44106.
24. Podda M, Weber C, Traber MG, Packer L: **Simultaneous determination of tissue tocopherols, tocotrienols, ubiquinol and ubiquinone.** *J Lipid Res* 1996, **37**:893–901.
25. Proteggente AR, Rota C, Majewicz J, Rimbach G, Minihane AM, Kraemer K, Lodge JK: **Cigarette smokers differ in their handling of natural (RRR) and synthetic (all rac) alpha-tocopherol: a biokinetic study in apoE4 male subjects.** *Free Radic Biol Med* 2006, **40**:2080–2091.
26. Traber MG, Rader D, Acuff RV, Ramakrishnan R, Brewer HB, Kayden HJ: **Vitamin E dose-response studies in humans with use of deuterated RRR-alpha-tocopherol.** *Am J Clin Nutr* 1998, **68**:847–853.
27. Kemsley EK, Le Gall G, Dainty JR, Watson AD, Harvey LJ, Tapp HS, Colquhoun IJ: **Multivariate techniques and their application in nutrition: a metabolomics case study.** *Br J Nutr* 2007, **98**:1–14.
28. Han X, Gross RW: **Shotgun lipidomics: electrospray ionization mass spectrometric analysis and quantitation of cellular lipidomes directly from crude extracts of biological samples.** *Mass Spectrom Rev* 2005, **24**:367–412.
29. Yang J, Zhao X, Liu X, Wang C, Gao P, Wang J, Li L, Gu J, Yang S, Xu G: **High performance liquid chromatography-mass spectrometry for metabonomics: potential biomarkers for acute deterioration of liver function in chronic hepatitis B.** *J Proteome Res* 2006, **5**:554–561.
30. Xia YQ, Jemal M: **Phospholipids in liquid chromatography/mass spectrometry bioanalysis: comparison of three tandem mass spectrometric techniques for monitoring plasma phospholipids, the effect of mobile phase composition on phospholipids elution and the association of phospholipids with matrix effects.** *Rapid Commun Mass Spectrom* 2009, **23**:2125–2138.
31. Leonard SW, Paterson E, Atkinson JK, Ramakrishnan R, Cross CE, Traber MG: **Studies in humans using deuterium-labeled alpha- and gamma-tocopherols demonstrate faster plasma gamma-tocopherol disappearance and greater gamma-metabolite production.** *Free Radic Biol Med* 2005, **38**:857–866.
32. Kougas P, Chai H, Lin PH, Lumsden AB, Yao Q, Chen C: **Lysophosphatidylcholine and secretory phospholipase A2 in vascular disease: mediators of endothelial dysfunction and atherosclerosis.** *Med Sci Monit* 2006, **12**:RA5–16.
33. Ojala PJ, Hirvonen TE, Hermansson M, Somerharju P, Parkkinen J: **Acyl chain-dependent effect of lysophosphatidylcholine on human neutrophils.** *J Leukoc Biol* 2007, **82**:1501–1509.
34. Murohara T, Ikeda H, Katoh A, Takajo Y, Otsuka Y, Haramaki N, Imaizumi T: **Vitamin E inhibits lysophosphatidylcholine-induced endothelial dysfunction and platelet activation.** *Antioxid Redox Signal* 2002, **4**:791–798.
35. Kohno M, Yokokawa K, Yasunari K, Minami M, Kano H, Hanehira T, Yoshikawa J: **Induction by lysophosphatidylcholine, a major phospholipid component of atherogenic lipoproteins, of human coronary artery smooth muscle cell migration.** *Circulation* 1998, **98**:353–359.
36. Feldman C, Anderson R, Theron AJ, Steel HC, van Rensburg CE, Cole PJ, Wilson R: **Vitamin E attenuates the injurious effects of bioactive phospholipids on human ciliated epithelium in vitro.** *Eur Respir J* 2001, **18**:122–129.
37. Wu D, Liu L, Meydani M, Meydani SN: **Vitamin E increases production of vasodilator prostanoids in human aortic endothelial cells through opposing effects on cyclooxygenase-2 and phospholipase A2.** *J Nutr* 2005, **135**:1847–1853.
38. Yea K, Kim J, Yoon JH, Kwon T, Kim JH, Lee BD, Lee HJ, Lee SJ, Kim JI, Lee TG, et al: **Lysophosphatidylcholine activates adipocyte glucose uptake and lowers blood glucose levels in murine models of diabetes.** *J Biol Chem* 2009, **284**:33833–33840.
39. Soga T, Ohishi T, Matsui T, Saito T, Matsumoto M, Takasaki J, Matsumoto S, Kamohara M, Hiyama H, Yoshida S, et al: **Lysophosphatidylcholine enhances glucose-dependent insulin secretion via an orphan G-protein-coupled receptor.** *Biochem Biophys Res Commun* 2005, **326**:744–751.

40. Wang-Sattler R, Yu Z, Herder C, Messias AC, Floegel A, He Y, Heim K, Campillos M, Holzapfel C, Thorand B, et al: **Novel biomarkers for pre-diabetes identified by metabolomics.** *Mol Syst Biol* 2012, **8**:615.
41. Zingg JM: **Modulation of signal transduction by vitamin E.** *Mol Aspects Med* 2007, **28**:481–506.
42. Chan AC, Wagner M, Kennedy C, Chen E, Lanuville O, Mezl VA, Tran K, Choy PC: **Vitamin E up-regulates arachidonic acid release and phospholipase A2 in megakaryocytes.** *Mol Cell Biochem* 1998, **189**:153–159.
43. Kwag OG, Kim SO, Choi JH, Rhee IK, Choi MS, Rhee SJ: **Vitamin E improves microsomal phospholipase A2 activity and the arachidonic acid cascade in kidney of diabetic rats.** *J Nutr* 2001, **131**:1297–1301.
44. Chandra V, Jasti J, Kaur P, Betzel C, Srinivasan A, Singh TP: **First structural evidence of a specific inhibition of phospholipase A2 by alpha-tocopherol (vitamin E) and its implications in inflammation: crystal structure of the complex formed between phospholipase A2 and alpha-tocopherol at 1.8 Å resolution.** *J Mol Biol* 2002, **320**:215–222.
45. Rhee SJ, Jeong YC, Choi JH: **Effects of vitamin E on phospholipase A2 activity and oxidative damage to the liver in streptozotocin-induced diabetic rats.** *Ann Nutr Metab* 2005, **49**:392–396.
46. Zingg JM, Azzi A: **Non-antioxidant activities of vitamin E.** *Curr Med Chem* 2004, **11**:1113–1133.
47. Silbert PL, Leong LL, Sturm MJ, Strophair J, Taylor RR: **Short term vitamin E supplementation has no effect on platelet function, plasma phospholipase A2 and lyso-PAF in male volunteers.** *Clin Exp Pharmacol Physiol* 1990, **17**:645–651.
48. Wang X, Quinn PJ: **Vitamin E and its function in membranes.** *Prog Lipid Res* 1999, **38**:309–336.
49. Fuchs B, Schiller J, Wagner U, Hantzschel H, Arnold K: **The phosphatidylcholine/lysophosphatidylcholine ratio in human plasma is an indicator of the severity of rheumatoid arthritis: investigations by 31P NMR and MALDI-TOF MS.** *Clin Biochem* 2005, **38**:925–933.
50. Iwase M, Sonoki K, Sasaki N, Ohdo S, Higuchi S, Hattori H, Iida M: **Lysophosphatidylcholine contents in plasma LDL in patients with type 2 diabetes mellitus: relation with lipoprotein-associated phospholipase A2 and effects of simvastatin treatment.** *Atherosclerosis* 2008, **196**:931–936.
51. Pietilainen KH, Sysi-Aho M, Rissanen A, Seppanen-Laakso T, Yki-Jarvinen H, Kaprio J, Oresic M: **Acquired obesity is associated with changes in the serum lipidomic profile independent of genetic effects - a monozygotic twin study.** *PLoS One* 2007, **2**:e218.
52. Zhao Z, Xiao Y, Elson P, Tan H, Plummer SJ, Berk M, Aung PP, Lavery IC, Achkar JP, Li L, et al: **Plasma lysophosphatidylcholine levels: potential biomarkers for colorectal cancer.** *J Clin Oncol* 2007, **25**:2696–2701.
53. Drobnik W, Liebisch G, Audebert FX, Frohlich D, Gluck T, Vogel P, Rothe G, Schmitz G: **Plasma ceramide and lysophosphatidylcholine inversely correlate with mortality in sepsis patients.** *J Lipid Res* 2003, **44**:754–761.

doi:10.1186/1743-7075-9-110

Cite this article as: Wong and Lodge: A metabolomic investigation of the effects of vitamin E supplementation in humans. *Nutrition & Metabolism* 2012 **9**:110.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

