Biomarker-driven stratification of disease-risk in non-metastatic medulloblastoma: Results from the multi-center HIT-SIOP-PNET4 clinical trial

Steven C. Clifford¹*, Birgitta Lannering², Ed C. Schwalbe¹,³, Debbie Hicks¹, Kieran O’Toole¹, Sarah Leigh Nicholson¹, Tobias Goschzik⁴, Anja zur Mühlen⁴, Dominique Figarella-Branger⁵, François Doz⁶, Stefan Rutkowski⁷, Göran Gustafsson⁸, Torsten Pietsch⁴*, on behalf of the SIOP-Europe PNET group

¹Northern Institute for Cancer Research, Newcastle University, Newcastle upon Tyne, United Kingdom
²Department of Pediatrics, University of Gothenburg and The Queen Silvia Children’s Hospital, Gothenburg, Sweden
³Department of Applied Sciences, Northumbria University, Newcastle upon Tyne, United Kingdom
⁴Department of Neuropathology, University of Bonn, Bonn, Germany
⁵Department of Pathology and Neuropathology, Assistance Publique Hôpitaux de Marseille, Aix Marseille University, Marseille, France
⁶Institut Curie and University Paris Descartes, Paris, France
⁷University Medical Center Hamburg-Eppendorf, Hamburg, Germany
⁸Karolinska Institute, Stockholm, Sweden
*These authors have contributed equally to this work

Correspondence to:
Steven C. Clifford, e-mail: steve.clifford@ncl.ac.uk

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ABSTRACT

Purpose: To improve stratification of risk-adapted treatment for non-metastatic (M0), standard-risk medulloblastoma patients by prospective evaluation of biomarkers of reported biological or prognostic significance, alongside clinico-pathological variables, within the multi-center HIT-SIOP-PNET4 trial.

Methods: Formalin-fixed paraffin-embedded tumor tissues were collected from 338 M0 patients (>4.0 years at diagnosis) for pathology review and assessment of the WNT subgroup (MB_WNT) and genomic copy-number defects (chromosome 17, MYC/MYCN, 9q22 (PTCH1) and DNA ploidy). Clinical characteristics were reviewed centrally.

Results: The favorable prognosis of MB_WNT was confirmed, however better outcomes were observed for non-MB_WNT tumors in this clinical risk-defined cohort compared to previous disease-wide clinical trials. Chromosome 17p/q defects were heterogeneous when assessed at the cellular copy-number level, and predicted poor prognosis when they occurred against a diploid (ch17(im)/diploid(cen)), but not polyploid, genetic background. These factors, together with post-surgical tumor residuum (R+) and radiotherapy delay, were supported as independent prognostic markers in multivariate testing. Notably, MYC and MYCN amplification were not associated with adverse outcome. In cross-validated survival models derived for the clinical standard-risk (M0/R0) disease group, (ch17(im)/diploid(cen); 14% of patients) predicted high disease-risk, while the outcomes of patients without (ch17(im)/diploid(cen)) did not differ significantly from MB_WNT, allowing re-classification of 86% as favorable-risk.
INTRODUCTION

Medulloblastoma is the most common malignant brain tumor in children. Risk-adapted therapeutic protocols in non-infant patients encompass maximal surgical resection, cranio-spinal radiotherapy and chemotherapy. Treatment groups and intensity are defined by the presence (‘high-risk’ disease) or absence (‘standard-risk’) of clinical features associated with a poor prognosis; metastatic disease at diagnosis and/or significant post-operative tumor residues, and this stratification currently forms the basis of patient selection into clinical trials [1].

Recent advances in the biological sub-classification of medulloblastoma are leading to the conception of clinical trials aimed at more precise therapeutic stratification and improved outcomes [1]. Historical studies have identified biomarkers consistently associated with favorable (β-catenin nuclear immunopositivity as a marker of the WNT medulloblastoma molecular subgroup (MB\textsubscript{WNT})) and poor (large-cell/anaplastic (LCA) pathology, MYC gene family amplification) prognosis [2–9], and their retrospective evaluation in the SIOP-UKCCSG-PNET3 clinical trial has validated their use alongside clinical factors for the improved definition of disease risk-stratification groups in disease-wide studies of non-infant medulloblastoma [10]. These stratification schemes will now form the basis of treatment selection in contemporary international clinical trials [1]. Additionally, further biomarkers with potential prognostic value, most notably the discovery of the four consensus medulloblastoma molecular subgroups (MB\textsubscript{WNT}, Sonic hedgehog (MB\textsubscript{SHH}), Group 3 (MB\textsubscript{Group3}) and Group 4 (MB\textsubscript{Group4})), are emerging from recent research studies on retrospective cohorts of medulloblastoma patients [1, 11–14].

The validation of novel biomarkers and risk-stratification schemes in clinically-controlled cohorts is thus essential to their clinical application. Moreover, the specific therapeutic regimens used may potentially impact the prognostic significance of specific biomarkers, and validation of their relevance within the defined treatment groups used in current clinical trials (i.e. the clinical standard- or high-risk disease groups) is necessary. This will require large-scale and coordinated international studies.

Here, we report the first European prospective study of medulloblastoma biomarkers, undertaken as part of the multi-center HIT-SIOP-PNET4 trial (2001–2006), which enrolled 338 children from 120 centres, with clinically-defined non-metastatic, standard-risk medulloblastoma [15]. Sufficient FFPE tumor material was collected for prospective assay (2004–2010) of a selected panel of biomarkers of previously reported biological or prognostic significance (i.e. in ≥2 published series). The study aimed to (i) improve the early identification of the ~20% patients with standard-risk medulloblastoma which cannot be cured by current treatment concepts, and (ii) identify patients with a favorable treatment concept who may qualify for a controlled reduction of adjuvant treatment schemes.

RESULTS

HIT-SIOP-PNET4: Clinical and treatment-related factors

338 patients, aged 4 to 21 at diagnosis, were enrolled and their clinical characteristics have been reported previously [15]. In summary, male patients predominated (211 male, 127 female) and the five-year EFS (all patients, including R+ disease) was 79 ± 2%. Features significantly associated with reduced EFS in univariate analysis in the clinical study were: (i) R+ disease (31/317 (9.8%); p = 0.020), and (ii) a delay to the start of radiotherapy (≥49 days after surgery (30/335 (9.0%); p = 0.050) or as a continuous variable (p = 0.025)). Patient gender and age at diagnosis were not associated with EFS [15]. Histopathological review was completed for 336/338 patients, and identified 273 CMB (81%), 47 DMB (14%) and 16 LCA (5%) tumors. There was no survival difference between CMB and DMB patients. The 16 patients with LCA subtype tumors enrolled on the study prior to amendment showed a higher frequency of relapses, but these did not reach significance [15]. The exclusion of further LCA patients, together with M+ patients, from the HIT-SIOP-PNET4 cohort, as well as sharpening of the definitions of DMB and LCA in the revised WHO classification of tumors of the CNS in 2007 [16] may account for any variation in the distribution of histopathological variants compared to previously-reported non-infant disease-wide trials (i.e. 71 CMB (61%), 22 DMB (19%) and 23 LCA (20%) in SJMB96 [3]; 174 CMB (84%), 14 DMB (7%) and 19 LCA (9%) in SIOP-UKCCSG-PNET3 [10]). Data are summarized in Table 1.

MB\textsubscript{WNT} subgroup

22.8% (58/254) of assessable tumors were MB\textsubscript{WNT} positive by β-catenin IHC [10] (Figure 1A). 15.9% of tumors (31/195) harbored CTNNB1 activating mutations...
(Supplementary Figure 1); all except one were observed in tumors with strong nuclear protein accumulation ($n = 30$; $p < 0.001$; Figure 1B). A further 12 tumors displayed $\beta$-catenin positivity (28.6% (12/42)) in the absence of CTNNB1 exon 3 mutation.

**MBWNT subgroup tumors were clinically, pathologically and molecularly distinct [11]: All except one displayed CMB or LCA histology ($p = 0.005$), and the group displayed gender parity in contrast to the male predominance in non-MBWNT patients ($p = 0.002$).

All copy number aberrations (CNAs) tested were infrequent or absent in MBWNT and polyploidy was less frequent ($p = 0.002$; Figure 1B). MBWNT patients showed a broader age distribution than non-MBWNT, with 11/58 (19.0%) MBWNT patients ≥16.0 years old at diagnosis, suggesting a secondary peak in adolescents/young adults (Figure 1B, 1C).

Favorable outcomes for patients with MBWNT tumors were confirmed in univariate analysis, (Table 1, Figure 2 ($p = 0.019$, Cox proportional hazards test; $p = 0.003$, log-rank test)). CTNNB1 mutation did not reach significance.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Categories</th>
<th>$n$</th>
<th>Five-year pEFS ±SE</th>
<th>Univariate Hazard Ratio (± CI)</th>
<th>$p$-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>Male (M)</td>
<td>211</td>
<td>0.79 ± 0.03</td>
<td>1.0</td>
<td>0.533</td>
</tr>
<tr>
<td></td>
<td>Female (F)</td>
<td>127</td>
<td>0.80 ± 0.04</td>
<td>0.85 (0.52–1.41)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Ratio (M:F)</td>
<td>1.66:1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age*</td>
<td>Median (years) Min.-Max.</td>
<td>9.0</td>
<td>3–20</td>
<td>–</td>
<td>0.99 (0.93–1.05)</td>
</tr>
<tr>
<td>Pathology**</td>
<td>All others</td>
<td>320</td>
<td>0.80 ± 0.02</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>LCA</td>
<td>16</td>
<td>0.64 ± 0.14</td>
<td>1.76 (0.71–4.37)</td>
<td>0.262</td>
</tr>
<tr>
<td>Residual tumor</td>
<td>≤1.5 cm²</td>
<td>286</td>
<td>0.82 ± 0.02</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>&gt;1.5 cm²</td>
<td>31</td>
<td>0.64 ± 0.09</td>
<td>2.34 (1.22–4.50)</td>
<td>0.020</td>
</tr>
<tr>
<td>Time from diagnosis to radiotherapy</td>
<td>&lt;49 days</td>
<td>305</td>
<td>0.81 ± 0.02</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>≥49 days</td>
<td>30</td>
<td>0.67 ± 0.09</td>
<td>1.93 (0.99–3.79)</td>
<td>0.050</td>
</tr>
<tr>
<td>Time from diagnosis to radiotherapy*</td>
<td>Median (days) Min.-Max.</td>
<td>35</td>
<td>15–92</td>
<td>–</td>
<td>1.03 (1.00–1.05)</td>
</tr>
<tr>
<td>β-catenin nuclear accumulation</td>
<td>No</td>
<td>196</td>
<td>0.75 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>58</td>
<td>0.91 ± 0.04</td>
<td>0.40 (0.17–0.94)</td>
<td>0.019</td>
</tr>
<tr>
<td>CTNNB1 mutation</td>
<td>No</td>
<td>164</td>
<td>0.75 ± 0.04</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>31</td>
<td>0.89 ± 0.06</td>
<td>0.37 (0.12–1.21)</td>
<td>0.058</td>
</tr>
<tr>
<td>MYC/MYCN amplification (PCR)</td>
<td>No</td>
<td>160</td>
<td>0.79 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>23</td>
<td>0.72 ± 0.09</td>
<td>1.26 (0.53–3.00)</td>
<td>0.606</td>
</tr>
<tr>
<td>MYC amplification (iFISH)</td>
<td>No</td>
<td>157</td>
<td>0.81 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>4</td>
<td>1.0</td>
<td>0.22 (0.02–29.16)</td>
<td>0.542</td>
</tr>
<tr>
<td>MYCN amplification (iFISH)</td>
<td>No</td>
<td>147</td>
<td>0.82 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>13</td>
<td>0.77 ± 0.12</td>
<td>1.41 (0.42–4.67)</td>
<td>0.588</td>
</tr>
<tr>
<td>17p loss and/or 17q gain (diploid(cen)) (iFISH)</td>
<td>No</td>
<td>127</td>
<td>0.85 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>24</td>
<td>0.57 ± 0.10</td>
<td>3.12 (1.44–6.76)</td>
<td>0.007</td>
</tr>
<tr>
<td>Polyploid</td>
<td>No</td>
<td>72</td>
<td>0.78 ± 0.05</td>
<td>1.0</td>
<td></td>
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<tr>
<td></td>
<td>Yes</td>
<td>85</td>
<td>0.83 ± 0.04</td>
<td>0.81 (0.39–1.68)</td>
<td>0.572</td>
</tr>
<tr>
<td>PTCH1 (9q22) loss</td>
<td>No</td>
<td>138</td>
<td>0.80 ± 0.03</td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>13</td>
<td>0.85 ± 0.10</td>
<td>0.82 (0.19–3.46)</td>
<td>0.781</td>
</tr>
</tbody>
</table>

*Assessed as a continuous variable.
**Patients with LCA tumors recruited prior to study amendment in November 2003. pEFS, event-free survival probability; SE, standard error; CI, 95% confidence interval. Significant prognostic associations are marked in bold type.

Previously reported data shown for information [15].
(Table 1, Figure 2A, 2B). 6/58 (10.3%) MBWNT tumors relapsed; all had β-catenin nuclear accumulation in >50% of cells and 3/4 assessed tumors harbored a CTNNB1 mutation (Supplementary Figure 2A). Of note, the EFS rate in MBWNT patients aged ≥16.0 years at diagnosis appeared lower than in MBWNT patients <16.0 years (p = 0.058; Figure 1D). Direct comparison of patients aged below 16.0 years from the HIT-SIOP-PNET4 and SIOP-UKCCSG-PNET3 trials showed equivalent five-year survival rates (±SE) for MBWNT patients (0.951 (±0.034) vs. 0.877 (±0.058); p = 0.434), but significantly better outcomes (0.754 (±0.034) vs. 0.671 (±0.036); p = 0.033) in non-MBWNT patients, and reduced frequencies of clinical high-risk features (M+ and/or R+ disease), in the HIT-SIOP-PNET4 cohort (Supplementary Figure 3).

**Copy number aberrations**

MYC (2.5% (4/161) tumors tested) and MYCN (8.1% (13/160)) amplifications detected by iFISH were mutually exclusive and were detected in non-MBWNT disease (Figure 1). Neither was associated with previously reported high-risk clinical (residual tumor) or pathological (LCA tumors included in this study) [1] disease features, or an adverse prognosis (Table 1, Figure 2D, 2E). iFISH and qPCR estimations of MYC/MYCN copy number were concordant in 90% of tumors analyzed by both methods (n = 131). Polyploidy (54% (85/157)) and PTCH1 (9q22) losses (10.6% (13/125)) were not associated with prognosis (Table 1, Figure 2C, 2G).

Chromosome 17 imbalances (q-arm gains and/or p-arm losses) were frequent (45.7% (69/151) tumors assessed). Imbalances were molecularly heterogeneous (Figure 3), and occurred against diploid (2 signals) and polyploid (>2 signals) centromeric reference backgrounds (Figure 3A, 3C). Strikingly, this heterogeneity was clinically significant. Tumors with chromosome 17 imbalances/diploid background (ch17(im)/diploid(cen); 16% (24/151)) were significantly associated with a poor outcome (p < 0.007; Table 1, Figure 2F), while tumors with imbalances/polyploid background (ch17(im)/polyploid(cen); 30% (45/151)) were not, and behaved equivalently to balanced tumors (Figure 3B). Ch17(im)/polyploid(cen) (n = 24) most commonly involved p-arm loss
(to a single copy) in conjunction with q-arm gain (17/24), consistent with isochromosome (17q), but isolated p-arm losses (4/24) and q-arm gains (3/24) also contributed. This tumor group peaked in children 6–10 years at diagnosis and all but one tumor was found in non-MB_WNT disease, but the group was not associated with other clinico-pathological factors (Figure 3D). Notably, the prognostic significance of ch17(im)/diploid(cen) was gender-specific and 8/9 relapses in this group occurred in male patients (Supplementary Figure 4).

Clinical and biological prognostic factors: Multivariate analysis

Multivariate analyses were performed separately on the clinically-defined non-metastatic (i.e. M0) and standard-risk (i.e. M0/R0) patient cohorts within the HIT-SIOP-PNET4 study; these analyses were based on all features showing significance in univariate analysis ($p < 0.05$; Table 1, Figure 2). To account for all features not having been assessed in all patients, we performed this multivariate analysis on different patient groups, based on the data available (Table 2). In the non-metastatic patient cohort, R+ disease was independently significant in all analyses, while MB_WNT and time to radiotherapy were either significant or marginally significant, consistent with findings from the univariate analysis. In the standard-risk cohort (i.e. following removal of R+ patients), ch17(im)/diploid(cen) was the only feature independently and significantly predictive of a poor outcome (Table 2).

Risk stratification models for standard-risk medulloblastoma

The standard-risk patient group, defined by M0/R0 disease, forms the basis of current clinical trials [1]. We therefore next used our data to develop risk-stratification models for this patient group. First, we generated Cox models from all variables (listed in Table 1) using 90% of patients in our cohort, and selected the most significant...
model to predict survival for the remaining 10% of patients, using a 10-fold cohort re-selection strategy for cross-validation. Ch17(im)/diploid(cen) was selected as the sole prognostic feature in every fold, thus forming a model for the prediction of poor outcome within the standard-risk disease group (61 ± 13% vs. 89 ± 3% survival at five-years, \( p = 0.009 \)) (Figure 4). In each fold, the model was not improved by the addition of any other covariate. We next compared model performance in our standard-risk cohort, against the current clinico-biological stratification scheme for the SIOP-PNET5 trial [1], which was defined previously based on disease-wide non-infant cohorts [1, 10, 14] (Figure 4a). Survival prediction at 5-years using the new cross-validated model improved performance and increased the area-under-curve (AUC) from 0.609 to 0.630 in ROC curve analysis (Figure 4d), and allowed 86% (102/118) of patients to be classified into a favorable-risk group, compared to 20% (24/118) in the established model. In view of the established favorable prognosis of MBWNT tumors [1, 3, 10], confirmed in our cohort, we assessed the impact of MBWNT status within this new model (Figure 4c). Inclusion was not detrimental to model performance (Figure 4d), however five-year survival for the favorable-risk MBWNT (96 ± 4%) and non-MBWNT ch17(im)/diploid(cen) negative (87 ± 4%) patient groups were not significantly different (\( p = 0.189 \)). Finally, findings were equivalent when patients aged up to 16.0 years at diagnosis were considered in isolation (Supplementary Figure 5).

**DISCUSSION**

The prospective assessment within HIT-SIOP-PNET4 of disease-relevant biomarkers, with reported significance in ≥2 previous retrospective series, alongside clinical and pathological factors, has provided important new insights to biomarker-driven risk stratification in clinically-defined standard-risk medulloblastomas. Post-operative residual tumor, delayed radiotherapy and MBWNT were validated as independent prognostic factors. Importantly, in the standard-risk (M0/R0) disease group, which forms the basis of current clinical trials [1], the use of distinct biomarkers (chromosome 17 status determined at the cellular copy-number level) and novel survival models allows the improved stratification

**Figure 3: Chromosome 17 defects in HIT-SIOP-PNET4 cohort tumors.** Patterns (A, C) and prognostic significance (B; ‘p’, log-rank tests) of chromosome 17 defects detected by iFISH. C. iFISH analysis showing (i) 17p loss (single green signals) and (ii) 17q gain (three green signals) against a diploid centromeric background (two red signals). Nuclei are counterstained blue. D. Relationship of ch17(im)/diploid(cen) defects to clinico-pathological and molecular disease features assessed (‘p’, Fisher’s exact or \( \chi^2 \) tests; corrected and uncorrected values are shown). Abbreviations: ch, chromosome; im, imbalance (p-gain and/or q-loss); diploid(cen), diploid centromeric signal; polyploidy(cen), polyploidy centromeric signal.
Table 2: Multivariate analysis of independent risk factors in the HIT-SIOP-PNET4 cohort, shown for patients within the clinically-defined non-metastatic (M0, R+/R0) disease and standard-risk (M0, R0) patient groups

<table>
<thead>
<tr>
<th>Variable</th>
<th>Category</th>
<th>Non-metastatic (M0, R+/R0) disease</th>
<th>Standard-risk (M0, R0) disease</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Cohort 1 (n = 315)</td>
<td>Data available: Residual tumor status, Time to radiotherapy</td>
</tr>
<tr>
<td></td>
<td></td>
<td>n</td>
<td>HR (± CI)</td>
</tr>
<tr>
<td>Residual tumor</td>
<td>≤1.5 cm²</td>
<td>284 (90%)</td>
<td>1.0</td>
</tr>
<tr>
<td>Residual tumor</td>
<td>&gt;1.5 cm²</td>
<td>31 (10%)</td>
<td>2.74 (1.41–5.34)</td>
</tr>
<tr>
<td>Time to radiotherapy</td>
<td>Continuous</td>
<td>315</td>
<td>1.03 (1.01–1.06)</td>
</tr>
<tr>
<td>β-catenin nuclear accumulation</td>
<td>No</td>
<td>Data not available</td>
<td></td>
</tr>
<tr>
<td>β-catenin nuclear accumulation</td>
<td>Yes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>17p loss and/or 17q gain (diploid(cen))</td>
<td>No</td>
<td>Data not available</td>
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<tr>
<td>17p loss and/or 17q gain (diploid(cen))</td>
<td>Yes</td>
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</table>

Patients with central radiological and pathological review were analyzed (n = 315). Variables which displayed statistical significance in univariate analysis (Table 1) were tested in separate multivariate analyses based on patients with available data. Statistically significant independent risk-factors and associated hazard ratios are marked in bold type. HR, hazard ratio; CI, 95% confidence interval.
of disease-risk. Moreover, we have established first mechanisms for prospective pan-European biological studies as a basis for the assessment of biomarker-driven therapies, and future therapeutic advances.

Assessment of MB_wnt subgroup patients in the HIT-SIOP-PNET4 cohort revealed significant new insights to their clinical behavior, and comparison to other trials-based studies, to support future trial design. Their favorable prognosis was validated, supporting their consideration for individualized risk-stratified therapies; notably, despite equivalent EFS rates in the MB_wnt groups, a significantly higher EFS was observed for the non-MB_wnt group in our non-metastatic HIT-SIOP-PNET4 cohort (75% five-year EFS) compared to previous studies of the SIOP-UKCCSG-PNET3 trials cohort which included high-risk patients [2, 3]. MB_wnt relapses were observed at higher frequency in patients with delayed radiotherapy or aged >16.0 years at diagnosis (Figure 1D; Supplementary Figure 2). Patients >16.0 years were not ascertained in our previous analyses of SIOP-UKCCSG-PNET3 which defined the favorable prognosis of the MB_wnt group [2, 10], and the present data are consistent with the bi-modal age distribution and worse prognosis reported for adults compared to children within MB_wnt in retrospective series [4, 17, 18]. Although the incidence of such cases is too low to draw firm conclusions, these observations thus indicate patient age >16.0 years and delayed radiotherapy negatively influence survival in MB_wnt, and indicate radiotherapy is important component of its multi-modal treatment. Similarly, only 1 event was observed in MB_wnt tumors also displaying previously reported high-risk disease features (MB_wnt/R+ (n = 7), MB_wnt/LCA (n = 5)) supporting their favorable prognosis following standard-risk therapy. CTNNB1 mutation
rates in MB\textsubscript{WNT} tumors, defined by β-catenin IHC, were equivalent to SIOP-UKCCSG-PNET3 (30/42 (71.4%) and 20/31 (64.5%), respectively [10]) and our data did not support reduced survival rates in mutation-negative MB\textsubscript{WNT} (Supplementary Figure 2A).

Study design, and the prospective collection of clinical material and biological data, was undertaken in 2000–2010. This limited our ability to assess in the PNET4 cohort the other non-MB\textsubscript{WNT} molecular subgroups, on which consensus emerged in 2012 [11]. Insufficient FFPE-derived tumor material remained following our planned prospective analysis (see Materials and Methods; typically <200 ng double-stranded (ds)DNA [measured by picogreen fluorometric quantitation] and ≤1 4 μm section remaining), to enable subgroup assessment using established methods (approximately 1 μg FFPE-extracted dsDNA required for DNA methylation array analysis [14, 19, 20]; >2 sections for IHC-based assignment to MB\textsubscript{SHH} subgroup [21]).

From our findings, high-risk biomarkers previously validated in disease-wide studies appear to have different prognostic relevance in the standard-risk clinical disease group, from which tumors with high-risk clinical features (e.g. M\textsuperscript{+} disease) have been excluded. In this clinical context, MYC/MYCN amplification and LCA were not associated with each other, with clinical high-risk factors (one MYCN/R\textsuperscript{+} and one MYCN/LCA tumor were observed) or with poor prognosis when observed in isolation as risk-factors in the clinically-defined non-metastatic/standard-risk group reported. These findings are consistent with observations based on SIOP-UKCCSG-PNET3 (which included high-risk patients), where MYC/MYCN amplification were associated with LCA pathology and were prognostic in high-risk (i.e. when observed in tumors from patients with other high-risk disease features) but not standard-risk disease [7, 10]. In most recent genomics studies, MYC-amplified tumors were most commonly observed in MB\textsubscript{group1}, while MYCN-amplified tumors were associated with MB\textsubscript{SHH} (where they were also associated with TP53 mutation and LCA) and MB\textsubscript{group3} but were only prognostic in MB\textsubscript{SHH} [12, 14, 22–24]. Our findings may thus reflect (i) the numbers of MYC/MYCN amplified tumors observed in this cohort, or (ii) biological heterogeneity and/or subgroup-dependency, including the trial amendment to cease recruitment of patients with LCA tumors (see methods), potentially limiting ascertainment of tumors with interactions between these factors. Evaluation/outcome monitoring of further patients defined by these features will now be required, prior to any refinements of their prognostic relevance used in the design of future clinical studies.

Our findings fully support the continued consideration of R\textsuperscript{+} patients as high-risk, and their exclusion from the standard-risk disease group [1]. Most notably, ch17(im)/diploid(cen) was the strongest independent biomarker risk-factor in M0/R0 standard-risk disease, characterizing a patient group with <60% five-year EFS; however, imbalances against a polyploid background were not significant. This previously undisclosed and clinically-significant heterogeneity observed between chromosome 17 imbalanced tumors is important, and these data suggest the biological impact of different patterns of chromosome 17 imbalance is equivalently heterogeneous. Variable prognostic associations have been reported for chromosome 17 imbalances in previous large studies [6, 10]; however, the complex patterns of imbalance revealed by iFISH analysis of individual tumor cells in this study were either not investigated or not detectable using the whole-biopsy copy number methodologies (e.g. array-CGH, SNP array) employed in many previous studies [9, 24–28]. Chromosome 17 imbalances are predominantly observed in MB\textsubscript{group3} and MB\textsubscript{group4} [24] and, in whole-biopsy SNP-array based investigations, Shih et al. [12] recently reported that the association between iso (17q) and a poor prognosis was restricted to MB\textsubscript{group3} patients. Concerted studies are now required to reconcile these findings and establish the relationship between tumor subgroup, cellular patterns of chromosome 17 imbalance, and prognosis.

Following designation of ch17 (im)/diploid (cen) tumors as high-risk in cross-validated survival models of standard-risk patients within the HIT-SIOP-PNET4 cohort, the EFS of remaining patients did not differ significantly from the MB\textsubscript{WNT} group, allowing the classification of >80% of patients into a favorable-risk category. This model outperforms established prognostication schemes in our standard-risk cohort.

Alongside methods developed for testing chromosome 17 imbalances at the cellular level in routinely-collected tumor material, these findings provide a straightforward scheme for risk-stratification in the clinically homogeneous group of children with standard-risk medulloblastoma, and a strong basis for their validation and further investigation in future clinical trials of this group. Future study concepts must ensure collection of sufficient FFPE alongside high-quality biological material (e.g. snap-frozen, histologically-controlled tumor tissue), from large patient numbers, to support further biomarker discovery and validation, including understanding their behavior in the context of the consensus medulloblastoma expression / DNA methylation subgroups.

MATERIALS AND METHODS

Patient cohort, pathological review, material collection & processing

338 patients with non-metastatic (M0 [29]) medulloblastoma, treated in 120 European centers and 11 countries, were enrolled on HIT-SIOP-PNET4 over a 6 year period (2001–2006) [15]. Patients were randomized
to receive post-operative treatment with either hyper-
fractionated (HFRT) or conventionally fractionated/
standard (STRT) radiotherapy and were followed up
for a median of 4.8 years; all patients received the same
chemotherapy. Clinical features of the cohort have been
reported [15] (summarized in Table 1). The two treatment
arms showed no significant difference in 5-year event-
free survival (EFS) [15], and were considered together for
biological analysis.

Post-operative radiological review was undertaken
for 317/338 (93.7%) patients, the remainder were
reviewed locally. Patients without significant post-surgical
tumor residuum (≤1.5 cm²; R0) were defined as standard-
risk [1]. A histopathological diagnosis of medulloblastoma
was confirmed by five neuropathologists (including DF-B
and TP), who performed central reference review of all
patients. Tumors were classified using WHO criteria
[30], and assigned to the classic (CMB), desmoplastic/
nodular (DMB) or large-cell/anaplastic (LCA) sub-entities
[16]. LCA were defined by a predominant component of
tumor cells with either characteristic large-cell or severe
anaplastic cytology, or both [8, 16, 31, 32]. DMB showed
a significant tumor component with reticulin fiber-free
islands (nODULEs) surrounded by reticulin fiber-rich
tumor areas. Reactive fiber induction (‘desmoplastic
reaction’, e.g. due to leptomeningeal growth) did not
qualify as DMB [16, 33]. A study amendment was made
in November 2003 to not enroll further LCA tumors, on
the basis of their reported poor prognosis [8, 15, 31, 32].

During sample processing for reference pathology,
excess tumor material was collected for biological studies
where available. Two slides (4 μm tissue sections) were
prepared for β-catenin immunohistochemistry (IHC). In
addition, two tubes with 4 × 20 μm sections were
collected, one to isolate nuclei for interphase fluorescence
in situ hybridization (iFISH) analysis, the other for
genomic DNA extraction (CTNNB1 mutation, MYC/
MYCN qPCR analysis (see below)). DNA was extracted
using the QIaAmp DNA Mini Tissue Kit (Qiagen GmbH,
Düsseldorf, Germany) according to the manufacturer’s
instructions.

Assessment of copy number aberrations

Tumor nuclei were isolated and CNAs of reported
biological or prognostic significance assessed by iFISH
using probes specific for MYCN (2p24) and MYC (8q24)
amplification previously associated with poor prognosis
and LCA pathology [5–7, 10, 21]), PTCH1 (9q22; loss
associated with the sonic hedgehog medulloblastoma
molecular subgroup (MBshh) and DMB [5, 6, 11, 17, 21]),
and the p- and q- arms of chromosome 17 (imbalance
associated with a poor prognosis [5, 6]), alongside
reference probes to the chromosome 2, 8, 9 and 17
centromeric regions, as previously described [5, 10]. Signals
in >200 non-overlapping nuclei were scored to give region of interest: centromere signal ratios for
individual cells. For chromosome gains and losses, the
modal score was considered representative of genetic
status. MYC or MYCN gene amplification was defined by
double-minute patterns or homogeneously staining
regions in ≥5% of nuclei [5, 7, 10]. Tumor ploidy was
determined as the modal status of the four centromeric
probes (>2 signals at ≥2 probes, polyploid) assessed.
Quantitative PCR (qPCR) was used to estimate MYC and
MYCN gene copy numbers, as described [34, 35].

MBwnt subgroup status

Assessment of β-catenin nuclear accumulation by
IHC has been widely studied as an MBwnt biomarker
[9, 14, 36–38], and was performed as described using
the monoclonal anti-β-catenin antibody 14 (Transduction
laboratories) [2, 10, 39]. Tumors with >10% positive
nuclei were scored positive (nuclear accumulation); the
same cut-off as used in the published SIOP-UKCCSG-
PNET3 cohort [10]. The few tumors showing nuclear
accumulation in single cells (typically <5%) were
classified negative. For CTNNB1 mutation analysis,
exon 3 was PCR-amplified from tumor DNA using the
primers 5’-GATTTGATGGAGTTGGACATGG-3’/5’-
TGTTCTTGAGTGAAGGACTGAG-3′, and sequenced
using standard methods [39].

Prospective biological studies: molecular
biomarker assessment

Overall, biological data were collected prospectively
from FFPE tumor material for 269/338 (79.5%) patients.
Data collection rates varied according to the individual
assays and the amount of tissue available. Notably, the
success rate of sampling for β-catenin IHC did not differ
between centers recruiting low (≤2; 77% success) and
high (>7; 80%) patient numbers. Cases with available
biological data from each assay were distributed randomly
across the major disease demographics were thus
considered representative of the whole cohort for further
analysis (Supplementary Table 1).

Statistical analysis

EFS was defined as time from diagnosis to
recurrence, progression or death during remission (of
any reason). Patients not experiencing an event were
censored at last follow-up. The database and biological
data collection for this analysis was closed July 1st 2010.
Kaplan–Meier curves, log-rank tests and unadjusted Cox
proportional hazards models were used to test univariate
EFS markers. Adjusted Cox proportional hazards
models were used to test univariate
Kaplan–Meier curves, log-rank tests and unadjusted Cox
proportional hazards models were used to test univariate
EFS markers. Adjusted Cox proportional hazards
models were used to test univariate
disease-risk markers. For univariate and multivariate survival
analyses which included ‘time to radiotherapy’ (as a
continuous variable), EFS times were landmarked to
the commencement of radiotherapy. Risk stratification
models for standard-risk medulloblastoma were developed in the cohort of M0/R0 patients with data available for all prognostic parameters (n = 118). This cohort was demographically representative of the entire standard-risk cohort within HIT-SIOP-PNET4 (n = 286; Supplementary Table 2). Associations between clinico-pathologic and/or molecular variables were examined using Fisher’s exact or X² tests, as appropriate. P-values were corrected for multiplicity using the Bonferroni method where indicated. Analysis was performed using SPSS (SPSS, Chicago, U.S.A.) and R [40] software. Data proportions presented in Tables and Figures are based on patients with available data and may not add to 100% due to rounding.

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CONFLICTS OF INTEREST

No author had any potential conflicts of interest to disclose.

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