

# Northumbria Research Link

Citation: Mann, Paul, Spencer, Robert, Hernes, Peter, Six, Johan, Aiken, George, Tank, Suzanne, McClelland, James, Butler, Kenna, Dyda, Rachael and Holmes, Robert (2016) Pan-arctic trends in terrestrial dissolved organic matter from optical measurements. *Frontiers in Earth Science*, 4. p. 25. ISSN 2296-6463

Published by: Frontiers

URL: <http://dx.doi.org/10.3389/feart.2016.00025>  
<<http://dx.doi.org/10.3389/feart.2016.00025>>

This version was downloaded from Northumbria Research Link:  
<http://nrl.northumbria.ac.uk/id/eprint/26135/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)

## Pan-arctic trends in terrestrial dissolved organic matter from optical measurements

Paul J. Mann<sup>1\*</sup>, Robert G. Spencer<sup>2</sup>, Peter J. Hernes<sup>3</sup>, Johan Six<sup>4</sup>, George R. Aiken<sup>5</sup>, Suzanne E. Tank<sup>6</sup>, James W. McClelland<sup>7</sup>, Kenna D. Butler<sup>5</sup>, Rachael Y. Dyda<sup>3</sup>, Robert M. Holmes<sup>8</sup>

<sup>1</sup>Department of Geography, Northumbria University, United Kingdom, <sup>2</sup>Department of Earth, Ocean and Atmospheric Science, Florida State University, USA, <sup>3</sup>Department of Land, Air, and Water Resources, University of California, USA, <sup>4</sup>Department of Environmental Systems Science, ETH-Zurich, Switzerland, <sup>5</sup>United States Geological Survey, USA, <sup>6</sup>Department of Biological Sciences, University of Alberta, Canada, <sup>7</sup>Marine Science Institute, University of Texas, USA, <sup>8</sup>Woods Hole Research Center, USA

*Submitted to Journal:*  
Frontiers in Earth Science

*Specialty Section:*  
Marine Biogeochemistry

*ISSN:*  
2296-6463

*Article type:*  
Original Research Article

*Received on:*  
05 Oct 2015

*Accepted on:*  
23 Feb 2016

*Provisional PDF published on:*  
23 Feb 2016

*Frontiers website link:*  
[www.frontiersin.org](http://www.frontiersin.org)

*Citation:*  
Mann PJ, Spencer RG, Hernes PJ, Six J, Aiken GR, Tank SE, McClelland JW, Butler KD, Dyda RY and Holmes RM(2016) Pan-arctic trends in terrestrial dissolved organic matter from optical measurements. *Front. Earth Sci.* 4:25. doi:10.3389/feart.2016.00025

*Copyright statement:*  
© 2016 Mann, Spencer, Hernes, Six, Aiken, Tank, McClelland, Butler, Dyda and Holmes. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

Provisional

# **Pan-arctic trends in terrestrial dissolved organic matter from optical measurements**

**Paul J. Mann<sup>1\*</sup>, Robert G. M. Spencer<sup>2</sup>, Peter J. Hernes<sup>3</sup>, Johan Six<sup>4</sup>, George R. Aiken<sup>5</sup>, Suzanne E. Tank<sup>6</sup>, James W. McClelland<sup>7</sup>, Kenna D. Butler<sup>5</sup>, Rachael Y. Dyda<sup>3</sup>, Robert M. Holmes<sup>8</sup>.**

**\*Correspondence:** Dr Paul J Mann, <sup>1</sup>Department of Geography, Northumbria University, Newcastle-upon-Tyne, UK.  
paul.mann@northumbria.ac.uk

**Running title:** Arctic tDOM export and composition

**Keywords:** Carbon Cycle, Arctic, Lignin, Colored Dissolved Organic Matter (CDOM), Parallel Factor Analysis (PARAFAC), DOC, Climate Change, Hydrology.

<sup>1</sup>Department of Geography, Northumbria University, Newcastle-upon-Tyne, UK.

<sup>2</sup>Department of Earth, Ocean and Atmospheric Science, Florida State University, Tallahassee, FL, 32306, USA.

<sup>3</sup>Department of Land, Air, and Water Resources, University of California, Davis, California, USA.

<sup>4</sup>Department of Environmental Systems Science, ETH-Zurich, 8092 Zurich, Switzerland.

<sup>5</sup>United States Geological Survey, Boulder, Colorado, USA.

<sup>6</sup>Department of Biological Sciences, University of Alberta, Edmonton, Canada.

<sup>7</sup>University of Texas, Marine Science Institute, 750 Channel View Drive, Port Aransas, TX 78373, USA.

<sup>8</sup>Woods Hole Research Center, Falmouth, MA, 02540, USA.

## 1 **Abstract**

2 Climate change is causing extensive warming across arctic regions resulting in  
3 permafrost degradation, alterations to regional hydrology, and shifting amounts and  
4 composition of dissolved organic matter (DOM) transported by streams and rivers.  
5 Here, we characterize the DOM composition and optical properties of the six largest  
6 arctic rivers draining into the Arctic Ocean to examine the ability of optical  
7 measurements to provide meaningful insights into terrigenous carbon export patterns  
8 and biogeochemical cycling. The chemical composition of aquatic DOM varied with  
9 season, spring months were typified by highest lignin phenol and dissolved organic  
10 carbon (DOC) concentrations with greater hydrophobic acid content, and lower  
11 proportions of hydrophilic compounds, relative to summer and winter months.  
12 Chromophoric DOM (CDOM) spectral slope ( $S_{275-295}$ ) tracked seasonal shifts in DOM  
13 composition across river basins. Fluorescence and parallel factor analysis identified  
14 seven components across the six Arctic rivers. The ratios of ‘terrestrial humic-like’  
15 versus ‘marine humic-like’ fluorescent components co-varied with lignin monomer  
16 ratios over summer and winter months, suggesting fluorescence may provide  
17 information on the age and degradation state of riverine DOM. CDOM absorbance  
18 ( $a_{350}$ ) proved a sensitive proxy for lignin phenol concentrations across all six river  
19 basins and over the hydrograph, enabling for the first time the development of a single  
20 pan-arctic relationship between  $a_{350}$  and terrigenous DOC ( $R^2 = 0.93$ ). Combining this  
21 lignin proxy with high-resolution monitoring of  $a_{350}$ , pan-arctic estimates of annual  
22 lignin flux were calculated to range from 156 to 185 Gg, resulting in shorter and more  
23 constrained estimates of terrigenous DOM residence times in the Arctic Ocean  
24 (spanning 7 months to 2½ years). Furthermore, multiple linear regression models  
25 incorporating both absorbance and fluorescence variables proved capable of

26 explaining much of the variability in lignin composition across rivers and seasons.  
27 Our findings suggest that synoptic, high-resolution optical measurements can provide  
28 improved understanding of northern high-latitude organic matter cycling and flux, and  
29 prove an important technique for capturing future climate-driven changes.

Provisional

## 30 **1. Introduction**

31 Northern high-latitude regions contain substantial quantities of organic carbon in  
32 perennially and seasonally frozen soils, comprising more than half the entire global  
33 carbon soil stock (Tarnocai et al., 2009). Large arctic rivers play an increasingly  
34 recognized role in regional carbon cycling by transporting a proportion of this  
35 terrigenous material from land to the ocean, whilst also acting as sites for active  
36 carbon metabolism and transformation (Holmes et al., 2011; Mann et al., 2015;  
37 Spencer et al., 2015; Striegl et al., 2005). Arctic riverine export is substantial enough  
38 (~ 10 % of the global freshwater discharge) that it imparts estuarine-like water quality  
39 characteristics throughout the Arctic Ocean, influencing coastal salinity structure on a  
40 localized basis (Aagaard and Carmack, 1989; McClelland et al., 2011; Serreze et al.,  
41 2006). Furthermore, significant quantities of dissolved organic matter (DOM)  
42 accompany this freshwater flux causing higher than average dissolved organic carbon  
43 (DOC) concentrations in the Arctic Ocean relative to other ocean basins (Hernes and  
44 Benner, 2006; Mathis et al., 2005; Opsahl et al., 1999). Six major arctic rivers account  
45 for the majority of freshwater flux, each draining vast watersheds on the Eurasian  
46 (Kolyma, Ob', Lena, Yenisey) or North American (Mackenzie, Yukon) continents,  
47 combined delivering ~ 64 % of the total freshwater supplied to the Arctic Ocean  
48 (Holmes et al., 2011).

49 Arctic rivers are characterized by their strong seasonality and large intra-  
50 annual variability in runoff, driven by extreme fluctuations in snow cover and air  
51 temperatures. Discharge rapidly peaks with the onset of snow melt and ice-breakup,  
52 resulting in dramatic spring freshet events and rapid transport of terrigenous DOM  
53 offshore (Amon et al., 2012; Mann et al., 2012; Stedmon et al., 2011a). By contrast,  
54 winter months are distinguished by low discharge and DOC concentrations, with

55 DOM exhibiting lower average aromaticity and molecular weight (O'Donnell et al.,  
56 2012; Spencer et al., 2008). Future changes in the fluxes and composition of  
57 terrigenous DOM released to and exported from arctic rivers are likely. River  
58 discharge across much of the pan-arctic watershed is increasing, particularly during  
59 winter months (Déry et al., 2009; McClelland et al., 2006; Peterson, 2002; Rawlins et  
60 al., 2010; Smith et al., 2007). Deepening of the seasonally thawed active layer will  
61 also result in leaching of deeper soil and permafrost horizons altering the amount and  
62 type of DOM liberated to inland waters (Romanovsky et al., 2010). Changes in the  
63 quality of DOM affect the reactivity and fate of terrigenous DOM, influencing carbon  
64 turnover rates and regional carbon budgets (Holmes et al., 2008; Mann et al., 2012;  
65 2014; Wickland et al., 2012). Tracing future alterations in the composition as well as  
66 concentration of riverine DOM is therefore crucial for understanding the effects of  
67 climate change.

68 Lignin phenols are unique biomarkers of vascular plant material and therefore  
69 act as sensitive indicators for the terrigenous component of aquatic DOM. As well as  
70 providing pertinent information on DOM source, lignin phenols also have the capacity  
71 to capture degradative processing and source information (Hernes et al., 2007; Opsahl  
72 and Benner, 1995; Spencer et al., 2010a). DOM source and composition have also  
73 been assessed via separation of the DOM pool using XAD fractionation techniques.  
74 DOC fractionation has been used to differentiate between high molecular weight,  
75 aromatic dominated carbon DOM fractions, primarily sourced from allochthonous  
76 materials, and those dominated by microbially-derived or photodegraded DOM (e.g.  
77 Aiken et al., 1992; Spencer et al., 2012). Despite providing critical information, both  
78 lignin phenol and XAD fractionation techniques are costly and extremely time  
79 consuming to conduct, limiting their applicability for high-resolution monitoring. The



80 remote nature of arctic watersheds and the rapid shifts in hydrology make effective  
81 sampling and observation of these regions incredibly challenging. Despite far greater  
82 understanding of constituent fluxes and biogeochemical cycles across Arctic river  
83 systems, much garnered from international sampling campaigns (e.g. PARTNERS,  
84 Arctic-GRO; [www.arcticgreatrivers.org](http://www.arcticgreatrivers.org)), insufficient temporal and spatial resolution  
85 in measurements still limits our ability to capture changes in terrigenous DOM supply  
86 and examine how it may alter under future scenarios. For example, the Arctic Great  
87 Rivers Observatory (Arctic-GRO) captures the major seasonal patterns in river  
88 chemistry and freshwater discharge across the six major arctic rivers, ensuring  
89 identical sampling and analytical protocols, yet is limited with respect to the number  
90 of samples that can be feasibly collected. The use of optical measurements, which can  
91 be rapidly collected and measured, remotely derived or determined *in-situ*, is one  
92 pathway that can help to address these problems.

93 A number of studies have investigated the ability of optical measurements to  
94 capture changes in DOM composition occurring across rivers or over the hydrograph,  
95 or to relate optical and lignin-based proxies to improve estimates of terrigenous DOM  
96 residence times in the Arctic Ocean (Spencer et al., 2009; Stedmon et al., 2011a;  
97 Walker et al., 2013). Recently, chromophoric DOM (CDOM) absorbance  
98 measurements from 30 unique US watersheds were shown to correlate to DOM  
99 composition, as derived via XAD fractionation, highlighting the potential of optical  
100 measurements to improve our understanding of DOM dynamics in fluvial systems  
101 (Spencer et al., 2012). Additionally, CDOM absorbance-lignin relationships have  
102 been developed for the Yukon River and then scaled to the pan-arctic, assuming  
103 similar relative loads of lignin in freshwater fluxes across all arctic rivers (Spencer et  
104 al., 2009). Using this approach, Spencer et al., (2009) found that terrigenous DOM

105 export to the Arctic Ocean was higher than previously thought, and thus concluded  
106 that a greater proportion must either be modified during transit through estuaries, or  
107 removal processes in the Arctic Ocean are greater than previously thought. CDOM  
108 fluorescence measurements have also been shown to be potentially useful proxies for  
109 lignin phenol concentration and composition in freshwaters (Hernes et al., 2009;  
110 Walker et al., 2013). Successful relationships have been reported between CDOM  
111 fluorescence, collected as excitation-emission matrices (EEMs) and decomposed  
112 using parallel factor analysis (PARAFAC), and lignin measurements across individual  
113 arctic rivers, yet pan-arctic relationships remain elusive (Walker et al., 2013). In  
114 particular, no studies have attempted to develop relationships between DOM optical  
115 properties from across all six arctic rivers and DOC fractionation measurements  
116 (XAD), or with vascular plant biomarkers (lignin phenols) as rapid proxies for  
117 terrestrial DOC export and composition across the Arctic. Additionally, no studies  
118 have examined the utility of combining absorbance and fluorescence techniques to  
119 develop arctic proxies for terrigenous DOM export.

120 Here, we characterize the DOM optical properties and composition (XAD and  
121 lignin phenol) of the six largest arctic rivers to examine the ability of optical  
122 measurements to provide meaningful insights into terrigenous carbon export patterns  
123 and biogeochemical cycling across broad spatial scales in the Arctic. Specifically, we  
124 attempt to identify common optical indices that trace DOC and lignin phenol  
125 concentration and compositional information across all six arctic rivers. Further, we  
126 examine the utility of using a combination of absorbance and fluorescence  
127 measurements to predict trends in DOC and lignin phenol biomarkers. Finally, we  
128 develop, for the first time, a pan-arctic optical proxy for estimating terrestrial OC flux

129 from arctic rivers to the Arctic Ocean and apply our findings to high-resolution  
130 optical measurements to improve terrigenous DOC export estimates.

131

132

## 133 **2. Material and Methods**

### 134 **2.1 Study Areas and Sample Collection**

135 Samples from each of the six largest Arctic rivers (Figure 1) were collected as part of  
136 the Arctic Great Rivers Observatory (Arctic-GRO; [www.arcticgreatrivers.org](http://www.arcticgreatrivers.org)). Each  
137 of the six rivers was sampled five times per year in 2009 and 2010 (except for 2009  
138 on the Yukon with six samples) using a standardized collection method as detailed  
139 elsewhere (Holmes et al., 2011; McClelland et al., 2008; Raymond et al., 2007).

140 Depth and width integrated samples were collected from near the mouth of each river  
141 (above tidal influence) across the hydrograph, incorporating baseflow, spring melt,  
142 and summer conditions. Near-daily surface sampling (0.5 m) was also conducted over  
143 the spring freshet hydrographs on each of the six Arctic rivers during both years to  
144 provide high-resolution optical measurements for this period ( $n = 241$ ).

145 Samples collected for DOC concentration, optical properties, and lignin  
146 analyses were filtered within a few hours of collection into pre-cleaned high-density  
147 polyethylene bottles through pre-rinsed 0.45  $\mu\text{m}$  capsule filters (Geotech or Pall  
148 Aquaprep 600) and measured on unfractionated waters. Samples for DOC  
149 fractionation were filtered as above and acidified to pH 2.

150

### 151 **2.2 Dissolved Organic Carbon and XAD Fractionation**

152 Dissolved organic carbon (DOC) measurements were performed on a Shimadzu  
153 (TOC-V) organic carbon analyzer as the mean of 3 – 5 replicate injections where the

154 coefficient of variance was < 2% (Mann et al., 2012). River water DOC samples were  
155 chromatographically separated into operationally defined hydrophobic organic acid  
156 (HPOA), hydrophobic neutral (HPON), low molecular weight hydrophilic (HPI), and  
157 transphilic organic acid (TPIA) fractions using XAD-8 and XAD-4 resins and  
158 established methodologies (Aiken et al., 1992). The amount of organic matter within  
159 each fraction is expressed as a percentage of the total DOC concentration and the  
160 sample mass of each fraction. The HPOA fraction typically contains more aromatic  
161 humic and fulvic acids and the HPI fraction less aromatic and more aliphatic forms of  
162 carbon, providing information on DOC composition.

163

### 164 **2.3 Lignin Phenol Biomarkers**

165 Lignin phenols were measured via the CuO oxidation method described by Hedges  
166 and Ertel, (1982), with modifications as outlined by Spencer et al., (2010b). In brief,  
167 filtered whole waters were acidified to pH 2 with 12N HCl, rotary evaporated to ~ 3  
168 mL, and the concentrate transferred to Monel reaction vessels (Prime Focus, Inc.) and  
169 dried under vacuum centrifugation. All samples were alkaline oxidized at 155 °C in a  
170 stoichiometric excess of CuO, followed by acidification (pH = 1 with 12 N H<sub>2</sub>SO<sub>4</sub>)  
171 and extracted three times with ethyl acetate, passed through Na<sub>2</sub>SO<sub>4</sub> drying columns,  
172 and taken to dryness under a gentle stream of ultrapure nitrogen. After redissolution in  
173 pyridine, lignin phenols were silylated (BSTFA) and quantified on a GC-MS (Agilent  
174 6890 gas chromatograph equipped with an Agilent 5973 mass selective detector and a  
175 DB5-MS capillary column; 30 m, 0.25 mm inner diameter, Agilent) using cinnamic  
176 acid as an internal standard and a five-point calibration scheme. Eight lignin phenols  
177 were quantified for all samples, including three vanillyl phenols (vanillin,  
178 acetovanillone, vanillic acid), three syringyl phenols (syringaldehyde, acetosyringone,

179 syringic acid), and two cinnamyl phenols (p-coumaric acid, ferulic acid). One blank  
180 was run for every ten-sample oxidations and all samples were blank corrected. Blank  
181 concentrations of lignin phenols were low (30 – 40 ng) and consequently never  
182 exceeded 5 % of the total lignin phenols in a sample. Lignin phenol concentrations are  
183 reported as the sum of the cinnamyl, syringyl and vanillyl phenols ( $\Sigma_8$ ). Additionally,  
184 the carbon normalized sum of the lignin phenols ( $\Lambda_8$ ) were calculated.

185

#### 186 **2.4 DOM Absorbance and Fluorescence**

187 UV-visible absorbance was measured in a 1 cm quartz cuvette across 200 – 800 nm at  
188 room temperature (20°C) with a dual beam Shimadzu UV-1800 spectrophotometer.

189 Measurements were recorded in triplicate at 1 nm wavelength intervals and referenced  
190 against Milli-Q water blanks. Absorbance values were converted to Napierian  
191 absorption coefficients by multiplying raw absorbance values by 2.303 and dividing  
192 by the pathlength (m) (Hu et al., 2002). The slope ( $S$ ) of the absorbance spectra was  
193 calculated from wavelength ranges spanning 275-295, 290-350, and 350-400 nm and  
194 the slope ratio ( $S_R$ ) determined as  $S_{275-295} / S_{350-400}$  (Helms et al., 2008). Slope  
195 coefficients can provide information pertaining to CDOM composition and source,  
196 with steeper values and increasing  $S_R$  indicative of lower molecular weight and  
197 decreasing DOM aromaticity (Blough and Del Vecchio, 2002; Blough and Green,  
198 1995; Helms et al., 2008). Specific UV absorbance ( $SUVA_{254}$ ) was calculated by  
199 dividing the decadal UV absorbance at 254 nm by the DOC concentration (Weishaar  
200 et al., 2003). The specific UV absorbance ( $SUVA_{254}$ ) has been shown to be positively  
201 correlated to percent aromaticity within DOM (Weishaar et al., 2003).

202 Fluorescence was analyzed using a Horiba Fluoromax-4 spectrofluorometer  
203 (Jobin-Yvon). Excitation-emission matrices (EEMs) were collected at 20°C in ratio

204 (S/R) mode over excitation and emission wavelengths of 250 - 450 and 320 - 550 nm,  
205 in 5 and 2 nm increments respectively. Measurements were performed with 0.1s  
206 integration times and 5 nm slit widths on the excitation and emission  
207 monochromators. Instrument specific correction files were applied before further  
208 analyses. Fluorescence EEMs were blank corrected from at least daily Milli-Q blanks  
209 collected identically to samples. Daily water Raman scans were collected at Ex=350  
210 nm (e.g. Lawaetz and Stedmon, 2009). Raman and Rayleigh-Tyndall scatter were  
211 removed and interpolated using the *smootheem* function and absorbance  
212 measurements were used to correct EEMs for inner filter effects according to the  
213 method of Lakowicz, (2013) within the drEEM toolbox (Murphy et al., 2013). The  
214 fluorescence index (FI) was also calculated as the ratio of emission at 470 nm to 520  
215 nm, at an excitation wavelength of 370 nm (Cory et al., 2010; McKnight et al., 2001).

216

## 217 **2.5 PARAFAC and statistical analyses**

218 Exploratory analysis of fluorescence EEM data was conducted using parallel factor  
219 analysis (PARAFAC) to decompose the number, shape, and amounts of underlying  
220 spectral components among samples. PARAFAC was conducted using the drEEM  
221 (version 2.0) and *N*-way (version 3.20) toolbox (Murphy et al., 2013) within the  
222 MATLAB R2013a environment.

223 To aid decomposition and provide greater variance within the dataset,  
224 additional EEMs (total  $n = 645$ ) collected across a wide range of stream and river  
225 environments from the Kolyma River Basin were included and analyzed alongside the  
226 Arctic-GRO fluorescence dataset. PARAFAC modeling was performed after  
227 normalizing each EEM to its total signal (to unit norm) by dividing by the sum of the  
228 squared values of all variables in the sample, and imposing non-negativity constraints,

229 thus negating problems caused by large concentration gradients apparent in seasonal  
230 samples (Murphy et al., 2013). The number of components within the model was  
231 validated using all of the techniques recommended in Murphy et al., (2013), including  
232 examination of systematic variation in the dataset, visualization of spectral loadings,  
233 and split half analysis. The final model was successfully validated using four splits of  
234 the data and three validation tests across six different dataset halves (S<sub>4</sub>C<sub>6</sub>T<sub>3</sub>)  
235 (Harshman and Lundy, 1994; Murphy et al., 2013). Fluorescence loadings were  
236 calculated after normalizing the dataset ensuring unscaled model scores were  
237 recovered.

238 Principle component analysis (PCA; nonrotated solutions) was employed to  
239 explore relationships between optical properties, DOC and lignin phenol  
240 concentration, and composition using the PLS\_Toolbox (Eigenvector, Inc., Seattle,  
241 WA v. 8.0) within MATLAB (R2013a). Autoscaling was used on variables measured  
242 prior to PCA. Multiple linear regression models were developed using a forward  
243 stepwise approach minimizing the Akaike Information Criterion, and were conducted  
244 in SPSS v22 (IBM).

245

## 246 **2.6 Constituent Flux Calculations**

247 Constituent fluxes were estimated using the USGS LoadEstimator software  
248 (LOADEST) within the LoadRunner software interface (Booth et al. 2007; Runkel et  
249 al., 2004). LOADEST calculates daily constituent flux estimates by generating  
250 relationships between measured discharge and element concentrations and was run as  
251 in Holmes et al., (2012). Daily discharge data were obtained from US Geological  
252 Survey (Yukon), Water Survey of Canada (Mackenzie) and Roshydromet (Kolyma,  
253 Lena, Ob' and Yenisey), and are freely available from

254 <http://dx.doi.org/10.5066/F7P55KJN>, <http://arcticgreatrivers.org/data.html>, and the  
255 Water Survey of Canada [http://wateroffice.ec.gc.ca/mainmenu/  
256 historical\\_data\\_index\\_e.html](http://wateroffice.ec.gc.ca/mainmenu/historical_data_index_e.html). Corrections were applied to allow for the distance  
257 between the discharge measurement station and sample location as has been  
258 previously described (Holmes et al., 2012). Any gaps in the discharge data were filled  
259 by interpolation, however there were no gaps during peak flow on any river.

260

### 261 **3. Results**

262

#### 263 **3.1 Spatial and temporal patterns in chemical fractions of DOC**

264 Total DOC concentrations ranged over the study period from 2.6 to 17.5 mg L<sup>-1</sup>.  
265 Highest average DOC concentrations were measured in the Lena river (15.7 ± 0.9 mg  
266 L<sup>-1</sup>, ± SE) during spring, and lowest during winter in the Yukon river (2.9 mg L<sup>-1</sup>, *n* =  
267 1; Table 1). DOC concentrations from all six rivers were correlated with runoff (m<sup>3</sup>  
268 km<sup>-2</sup> d<sup>-1</sup>; R<sup>2</sup> = 0.44, not shown). Riverine DOC was mainly composed of high  
269 contributions from the HPOA fraction, averaging 53 ± 1% across the six Arctic rivers  
270 (Table 1). Eurasian rivers contained higher average proportions of the HPOA fraction  
271 over the year (54 to 56 ± 2%) relative to the North American Yukon (50 ± 2%) and  
272 Mackenzie rivers (45 ± 2%, *t*-test *p* < 0.001). HPI and TPIA fractions contributed a  
273 smaller proportion to bulk DOC, averaging 19 ± 0% and 17 ± 0% respectively. The  
274 average HPI and TPIA fractions were less variable than HPOA, ranging from 17 to  
275 23% and 16 to 19% respectively across all sites and seasons (Table 1). HPON  
276 fractions generally contributed < 10 % to bulk DOC across rivers and were thus  
277 omitted from further study.



278 The composition of DOC was further characterized by calculating specific  
279 UV-visible absorbance ( $SUVA_{254}$ ) of DOC and its major chemical fractions (Table 1).  
280 Mean  $SUVA_{254}$  values of total DOC varied considerably among rivers with lowest  
281 values measured in the Mackenzie River ( $2.5 \text{ L mgC}^{-1} \text{ m}^{-1}$ ) and highest in the Yenisey  
282 River ( $3.4 \text{ L mgC}^{-1} \text{ m}^{-1}$ ). Mean  $SUVA_{254}$  values of the HPOA fraction also varied  
283 among rivers with lowest values observed in the Kolyma River ( $3.6 \text{ L mgC}^{-1} \text{ m}^{-1}$ ) and  
284 highest in the Yenisey ( $4.3 \text{ L mgC}^{-1} \text{ m}^{-1}$ ), and were consistently higher than bulk DOC  
285 highlighting the greater number of highly aromatic compounds represented by this  
286 fraction. The  $SUVA_{254}$  values of the HPI ( $1.2$  to  $2.3 \text{ L mgC}^{-1} \text{ m}^{-1}$ ) and TPIA ( $2.0$  to  
287  $3.1 \text{ L mgC}^{-1} \text{ m}^{-1}$ ) fractions were less variable across rivers, and lower than bulk DOC,  
288 indicating the presence of a lower relative number of aromatic moieties (Table 1).

289 The contribution of HPOA to the total DOC pool was generally highest during  
290 spring months with maximum contributions varying considerably between rivers (47  
291 to 60 %; Table 1). Percent contributions of the HPOA fraction were typically lowest  
292 during winter flow periods. No clear seasonal differences in the fraction of HPOA  
293 present were observed in the Ob' River ( $55 \pm 3$  to  $57 \pm 1$  %). Mean  $SUVA_{254}$  values of  
294 total DOC were consistently highest across all rivers during spring months,  
295 intermediate during summer months ( $2.3$  to  $3.1 \text{ L mgC}^{-1} \text{ m}^{-1}$ ), and lowest in winter  
296 across all rivers ( $1.5$  to  $3.0 \text{ L mgC}^{-1} \text{ m}^{-1}$ ; Table 1).  $SUVA_{254}$  values of the HPOA  
297 fraction followed similar seasonal trends as total DOC. TPIA and HPI  $SUVA_{254}$   
298 values displayed less clear seasonal patterns (Table 1).

299

### 300 **3.2 Spatial and temporal patterns in lignin phenols**

301 Mean lignin phenol concentrations ( $\Sigma_8$ ) varied significantly among the six rivers, with  
302 lowest concentrations observed in the Mackenzie River ( $9.5 \mu\text{g L}^{-1}$ ) and highest in the

303 Lena River ( $70.0 \mu\text{g L}^{-1}$ ; Table 2). Highest carbon normalized lignin yields ( $\Lambda_8$ ) were  
304 observed in the Yenisey River, mostly due to lower mean DOC concentrations  
305 relative to the Lena River (Table 2). Lowest mean  $\Lambda_8$  values were measured in the  
306 Mackenzie River ( $0.19 (\text{mg}(100 \text{ mg OC}))^{-1}$ ). Lignin values measured in this study  
307 were consistent with prior measurements in the Yukon and Russian arctic rivers  
308 (Lobbés et al., 2000; Spencer et al., 2009) but notably lower than lignin measurements  
309 from the earlier PARTNERS project (Amon et al., 2012).

310 The Lena and Yenisey rivers displayed lowest mean cinnamyl (C) to vanillyl  
311 (V) phenol ratios (C/V), indicative of greater contributions of woody versus non-  
312 woody sources to bulk DOM of these rivers (Hedges and Mann, 1979). Highest C/V  
313 ratios were measured in the Mackenzie and Ob' Rivers (Table 2). Spatial variability in  
314 syringyl (S) to vanillyl ratios (S/V) mainly mirrored those of C/V, except for higher  
315 S/V values in the Yukon River as compared to the Mackenzie River (Table 2). Higher  
316 S/V ratios are indicative of greater proportions of angiosperm versus gymnosperm  
317 sources to DOM (Hedges and Mann, 1979).

318 Acid to aldehyde ratios (Ad/Al) have been suggested to provide evidence of  
319 the relative degree of DOM degradation, with higher ratios indicating greater  
320 degradation of plant tissues (Hedges et al., 1988; Hernes and Benner, 2003; Opsahl  
321 and Benner, 1995). Mean ratios of vanillic acid to vanillin (Ad/Al)<sub>v</sub> ranged from 1.07  
322 in the Mackenzie River to 1.48 in the Yukon River (Table 2). Ratios of syringic acid  
323 to syringaldehyde (Ad/Al)<sub>s</sub> varied from 0.84 in the Yenisey to 1.06 in the Mackenzie  
324 River.

325  $\Sigma_8$  values among rivers were strongly linearly related to runoff ( $R^2 = 0.69$ ),  
326 suggesting terrigenous DOM export dynamics were largely controlled by hydrology.  
327 Accordingly, highest  $\Sigma_8$  concentrations were recorded across all rivers during the

328 spring freshet and lowest concentrations during base flow winter conditions. Highest  
329 individual  $\Sigma_8$  values were measured in the Lena River ( $120 \mu\text{g L}^{-1}$ ) and lowest in the  
330 Kolyma and Yukon Rivers ( $3.8 \mu\text{g L}^{-1}$ ). During the freshet,  $\Lambda_8$  yields were between  
331 2.6 and 4.8 times higher than winter  $\Lambda_8$  values across sites; the Yenisey River  
332 displayed the least variability and the Yukon the greatest (Table 2).

333 C/V and S/V ratios generally declined with increasing runoff across all rivers  
334 (Table 2). Acid aldehyde ratios (Al/Ad) were highly variable among rivers, generally  
335 increasing with greater runoff, yet in some cases (e.g. Lena River) demonstrated  
336 opposing patterns. Highest Ad/Al ratios during the spring freshet may represent the  
337 export of greater quantities of largely 'fresh' microbially unprocessed DOM relative  
338 to later in the year (Hernes et al., 2007; Spencer et al., 2008; Amon et al., 2012).

339

### 340 **3.3 Chromophoric and fluorescence DOM of arctic rivers**

341 The absorbance coefficient of CDOM at 350 nm ( $a_{350}$ ) ranged from 2.3 to  $42.6 \text{ m}^{-1}$   
342 among rivers and seasons, and similar to DOC and  $\Sigma_8$  concentrations, generally  
343 increased with greater freshwater runoff ( $R^2 = 0.57$ ,  $p < 0.001$ ,  $n = 60$ ; Supplemental  
344 Table 1). Spectral slope values ( $S_{275-295}$ ,  $S_{290-350}$ ,  $S_{350-400}$ ) steepened with decreasing  
345 runoff, in good agreement with previous studies (Spencer et al., 2009; Stedmon et al.,  
346 2011a), indicating the export of lower molecular weight material, or DOM with  
347 decreasing aromaticity as discharge rates decline (Blough and Del Vecchio, 2002;  
348 Blough and Green, 1995). The slope ratio ( $S_R$ ) showed an opposing pattern to spectral  
349 slopes, declining at higher runoff rates thus confirming an increase in DOM molecular  
350 weight during the spring freshet and reduction during winter base flow months  
351 (Helms et al., 2008; Spencer et al., 2010a; 2012).

352 Fluorescence index (FI) values were similar across rivers and averaged  $1.34 \pm$   
353  $0.01$  across all sites and sampling dates reflecting DOM from a mixture of terrigenous  
354 and microbial sources (McKnight et al., 2001; Supplemental Table 1). Highest FI  
355 values were measured across all rivers during winter months (mean  $1.41 \pm 0.03$ ),  
356 reflecting a potentially higher contribution of microbial derived or lower aromaticity  
357 DOM during this period. Lowest FI values among rivers (mean  $1.31 \pm 0.01$ ),  
358 indicative of greatest terrigenous and aromatic DOM supply, were observed during  
359 the high discharge spring freshet.

360

### 361 **3.4 Parallel factor analysis (PARAFAC)**

362 The PARAFAC analysis of DOM excitation-emission scans collected from all six  
363 arctic rivers over the hydrograph identified seven unique components of DOM  
364 fluorescence (Fig. 2; Supplemental Fig. 1). The spectra of each component identified  
365 were compared with the open fluorescence database (openfluor.org) containing  
366 spectra from previous studies (currently 53 studies) detailing PARAFAC models. All  
367 seven components closely matched (tucker congruence coefficient,  $TCC > 0.95$ ;  
368 Tucker, 1951) the excitation and emission spectra of previously identified  
369 components from 37 independent studies (Table 3).

370 Four of the seven AG components were very closely related ( $TCC \geq 0.97$ ) to  
371 those reported in the five-component Horsens catchment model (Murphy et al., 2014).  
372 For each of the seven components identified here, at least three independent studies  
373 had previously identified statistically similar spectra, except for component AG3 for  
374 which there was only a single match. Table 3 provides information and a description  
375 of the seven components identified.

376 AG3 was closely related (TCC = 0.97) to C1 in Murphy et al., (2014) where it  
377 was identified as displaying an emission spectrum identical to syringaldehyde, a  
378 product of lignin breakdown. The AG model shared two components with a five-  
379 component PARAFAC model explaining fluorescence DOM collected from five of the  
380 major Arctic rivers sampled here over 2004-2005 (Walker et al., 2013). AG6 was  
381 similar (TCC >0.95) to C1 from this model (Walker et al., 2013), while AG7 was  
382 identical to C5, which can be described as tryptophan-like and has been commonly  
383 associated with biological production in surface waters (Determann et al., 1994).

384 Two AG model components were also highly related to components  
385 previously reported in sea ice. AG1 was comparable to the terrestrially-derived  
386 component C2 found within Baltic sea ice (Stedmon et al., 2007). AG1 was also  
387 identical to C3 in coastal Canadian Arctic waters, which proved to be highly  
388 positively correlated with  $\Sigma_8$  (Walker et al., 2009). AG5 was identical to C6 in a study  
389 of Antarctic sea ice brines (Stedmon et al., 2011b), and is similar to the commonly  
390 described 'M' peak across a wide range of environments (Coble, 1996; 2007; Fellman  
391 et al., 2010).

392 Component AG4 contributed the greatest (23.4%  $\pm$  0.6%) and AG7 the lowest  
393 percentage (5.1%  $\pm$  0.3%) towards total DOM fluorescence across all rivers and  
394 seasons. In contrast to previous studies, no consistent pan-arctic seasonal or spatial  
395 patterns were apparent in the fluorescence loadings or percent contribution of any of  
396 the seven components (Walker et al., 2009). Individual patterns in fluorescence were,  
397 however, observed across rivers and seasons. Component AG1 contributed a  
398 substantially higher proportion of total fluorescence during the summer months in the  
399 Kolyma (20.5%  $\pm$  0.1%) and Lena Rivers (20.6%  $\pm$  0.6%) relative to each of the other  
400 rivers (16.8 to 17.7%), yet comprised similar amounts during the rest of the year.

401 Similarly, the proportion of AG3 was significantly higher in the Kolyma ( $23.2\% \pm$   
402  $0.8\%$ ) and Lena ( $19.1\% \pm 1.4\%$ ) Rivers relative to the others ( $11.3\%$  to  $17.6\%$ ) during  
403 the summer months alone. Opposing patterns were observed in AG6, with  
404 significantly lower proportions in the Kolyma ( $4.4\% \pm 0.1\%$ ) and Lena Rivers ( $5.3\%$   
405  $\pm 0.6\%$ ) relative to the others, in particular the Yenisey ( $9.7\% \pm 1.2\%$ ) and Ob' Rivers  
406 ( $8.4\% \pm 1.8\%$ ). The Mackenzie River contained high proportions of AG7 during the  
407 summer months ( $6.7\% \pm 1.1\%$ ) relative to all other rivers ( $3.9$  to  $5.2\%$ ).

408

409

## 410 **4. Discussion**

411

### 412 **4.1 Optical measurements, DOC concentration and DOM composition**

413 CDOM absorption ( $a_{350}$ ) correlated strongly with DOC concentration across all rivers  
414 during the standard Arctic-GRO sampling over 2009 and 2010 ( $R^2 = 0.89$ ;  $p < 0.001$ ;  $n$   
415  $= 60$ , not shown). This strong positive linear relationship persisted when DOC  
416 concentration and  $a_{350}$  values from the additional high-resolution measurements  
417 collected over the freshet period were included ( $R^2 = 0.81$ ;  $p < 0.001$ ;  $n = 301$ ; Figure  
418 3a). Despite this robust pan-arctic relationship however, when rivers were analyzed  
419 independently significant differences in the slopes and intercepts for the DOC to  $a_{350}$   
420 relationships were observed (Table 4). This indicates that the relative amount of non-  
421 chromophoric DOM varies across Arctic rivers, and suggests that the proportion of  
422 DOC per unit CDOM within individual river basins should in the future be separately  
423 determined (Table 4). Interestingly, we also find that the variability in river-specific  
424 slope and intercepts were well-explained by total annual river discharge, with  
425 increasing discharge resulting in higher DOC: $a_{350}$  intercepts ( $R^2 = 0.58$ ;  $p < 0.05$ , not

426 shown) and shallower slopes ( $R^2 = 0.72$ ,  $p < 0.05$ , not shown). The relationship  
427 between annual discharge and DOC: $a_{350}$  intercepts improved significantly ( $R^2 = 0.98$  ;  
428  $p < 0.01$ ) with the exclusion of the Mackenzie River. Thus, greater dilution of DOM  
429 and export of non-chromophoric organics occurs with increasing total discharge. The  
430 different relationship observed in the Mackenzie may be due to its relatively low  
431 DOM yield, high abundance of suspended sediments, as well as high proportion of  
432 lakes relative to other watersheds (Stedmon et al. 2011a).

433 HPOA fraction was closely related to  $S_{275-295}$  across rivers, with the relative  
434 proportion of HPOA decreasing with steepening slope ( $R^2 = 0.65$ ;  $p < 0.001$ ,  $n = 58$ ),  
435 as previously reported across five of these rivers in 2004-2005 (Walker et al., 2013).  
436 This suggests that average DOM molecular weight and aromaticity decreases as the  
437 proportion of HPOA declines, in good agreement with a number of previous studies  
438 (Neff et al., 2006; O'Donnell et al., 2012; Spencer et al., 2012; Striegl et al., 2007).  
439 This was further supported by a positive linear relationship between average HPOA  
440 and  $SUVA_{254}$  across rivers ( $R^2 = 0.56$ ,  $p < 0.01$ , Supplemental Fig. 2). The DOM  
441 composition of winter flow has been shown to contain lower proportions of the  
442 HPOA fraction as compared to HPI, with lower  $SUVA_{254}$  values relative to summer  
443 and spring months in the Yukon River (O'Donnell et al., 2012). The aromaticity of the  
444 HPOA fraction (HPOA- $SUVA_{254}$ ), was negatively correlated with FI ( $R^2 = 0.32$ ,  $p <$   
445  $0.001$ ,  $n = 47$ , not shown), confirming the role of terrigenous supply on delivering  
446 increased proportions of aromatic organics to the exported DOM pool.

447 No pan-arctic relationships were observed between any of the fluorescence  
448 component loadings and DOC concentration or composition (Supplemental Tables 2  
449 & 3). However, weak yet significant relationships were observed between the relative  
450 proportions of components AG3 and AG4 to total fluorescence and DOC

451 concentrations across all six rivers ( $R^2 = 0.15$  and  $0.16$  respectively,  $p < 0.01$ ). AG3  
452 proportions generally decreased with increasing DOC concentrations, where AG4  
453 proportions increased with greater DOC concentrations. These relationships were  
454 largely driven by particularly strong relationships across the Kolyma and Lena Rivers  
455 (AG3  $R^2 = 0.70$ ; AG4  $R^2 = 0.70$ ,  $p < 0.001$ ). The relatively weak pan-arctic  
456 relationships we observe here contrasts with the findings of Walker et al., (2013),  
457 whom report strong correlations between DOC concentration and fluorescence  
458 loadings. These conflicting findings may have been due to the additional  
459 normalization step we applied to scale each EEM to its total signal, thus ensuring the  
460 model focused entirely on compositional rather than concentration gradients.  
461 Alternately, the addition of a significant number of EEMs from upstream sources may  
462 have resulted in the validation of different components during PARAFAC  
463 decomposition. The latter seems unlikely however, as two of the seven components  
464 were spectrally identical ( $TCC > 0.95$ ) to the PARAFAC model used by Walker et al.,  
465 (2013), including component AG6 which was spectrally indistinguishable from a  
466 component (C1) identified as most closely tracing DOC and  $\Sigma_8$  concentrations. Other  
467 potential causes include differences in the treatment of inner filter effects. We applied  
468 a commonly employed post-hoc method by Lackowicz (2013) to correct our EEMs  
469 for inner filter effects using parallel CDOM absorbance measurements, whereas  
470 Walker et al. (2013) diluted samples prior to measurement. Our method more closely  
471 reflects direct measurement of field samples and is similar to information that could  
472 be derived from *in-situ* instruments. Our findings suggest that loadings derived from  
473 fluorescence EEMs decomposed using PARAFAC may not always be useful when  
474 tracing DOC concentration.

475



## 476 4.2 Optical measurements and lignin concentration and composition

477 CDOM ( $a_{350}$ ) measurements were highly correlated to  $\Sigma_8$  across all six rivers basins  
478 ( $R^2 = 0.93$ ;  $p < 0.001$ ;  $n = 31$ ; Figure 3b). This represents the first pan-arctic  
479 relationship to be reported between  $a_{350}$  and  $\Sigma_8$  across all six major rivers. The  
480 observed linear relationship ( $\Sigma_8 = -8.06 \pm 2.71 + (2.80 \pm 0.14a_{350})$ ) was similar to, yet  
481 displayed a slightly higher slope, than reported in Spencer et al., (2008) for the Yukon  
482 River Basin only ( $\Sigma_8 = -6.67 \pm 2.88 + (2.21 \pm 0.11a_{350})$ ). Previous studies have  
483 reported a much steeper linear relationship between  $a_{350}$  and  $\Sigma_8$ , with the Mackenzie  
484 and Ob' Rivers grouping separately from the Kolyma, Lena and Yenisey (Walker et  
485 al., 2013). The steeper slope of the previously reported relationship is caused by the  
486 substantially higher (often greater than double) lignin concentrations ( $\Sigma_8$ ) reported in  
487 Amon et al., (2012) and used in Walker et al., (2013) relative to those presented here.  
488 The differences in  $\Sigma_8$  concentrations may be due to methodological differences, as  
489 suggested by Walker et al., (2013), and raises concern over future potential in  
490 comparing datasets. For example, comparison of data from Spencer et al., (2008) in  
491 Walker et al., (2013) suggested low relative lignin concentrations in DOM from the  
492 Yukon, whereas we identify a similar  $\Sigma_8$  to  $a_{350}$  relationship across all six major arctic  
493 rivers.

494 Carbon normalized lignin ( $\Lambda_8$ ) yields decreased exponentially with steepening  $S_{275-295}$   
495 values across all rivers and seasons ( $R^2 = 0.80$ ;  $p < 0.01$ ,  $n = 31$ ; Table 2 and  
496 Supplementary Table 1). Steepening  $S_{275-295}$  values were associated with decreasing  
497 runoff rates (section 3.3), thus  $\Lambda_8$  yields typically increased as DOM average  
498 molecular weight and aromaticity increased, and during spring and summer months in  
499 response to greater allochthonous DOM supply.

500 Lignin phenols have been shown to comprise a major component of the HPOA  
501 fraction (Spencer et al., 2008; 2010a; Templier et al., 2005). This was confirmed by a  
502 significant positive correlation between proportion HPOA and  $\Lambda_8$  yields ( $R^2 = 0.71$ ,  $p$   
503  $< 0.001$ ,  $n = 29$ ; not shown). Lignin phenol biomarkers thus appear capable of  
504 providing information on the biogeochemical cycling of the entire hydrophobic DOM  
505 pool, which comprises up to two-thirds of aquatic DOM.

506 Lignin phenol C/V ratios increased with steepening  $S_{275-295}$  values ( $R^2 = 0.54$ ;  
507  $p < 0.001$ ,  $n = 31$ ) and declining total SUVA<sub>254</sub> values ( $R^2 = 0.48$ ;  $p < 0.001$ ,  $n = 31$ )  
508 (Tables 1 and 2 and Supplementary Table 1). The overall decline in C/V ratios with  
509 increasing freshwater runoff appears to represent increased contributions of lignin  
510 from litter and surface soil layers alongside greater proportions of aromatic and higher  
511 molecular weight DOM export (Hedges and Mann, 1979). These findings however  
512 appear counterintuitive relative to what we currently understand about hydrologic  
513 flow paths and sources of DOM to aquatic systems. Non-woody litter tissues  
514 associated with surficial, predominantly overland flow paths are expected to impart  
515 higher C/V ratios and lower degradative alteration than observed in DOM exported  
516 during base flow conditions derived from deeper flow paths. Physical processes, such  
517 as leaching and sorption, can also influence lignin phenol ratios (Hernes et al., 2007;  
518 2008) and may therefore be responsible for the observed trends. S/V and acid to  
519 aldehyde ratios did not correlate closely with spectral slope, SUVA<sub>254</sub>, or  $S_R$  values.  
520 The overall trends in lignin phenol composition we report are similar to those  
521 previously shown across Arctic rivers (Amon et al., 2012; Spencer et al., 2008; 2009),  
522 and demonstrate a shift from predominantly modern surface-derived and lignin-rich  
523 DOM during the spring freshet to older, less lignin-rich DOM under base-flow winter  
524 conditions.

525 No pan-arctic relationships were observed between fluorescent PARAFAC  
526 component loadings and lignin phenol concentration or composition measures  
527 (Supplemental Tables 2 & 3). A weak yet significant relationship was however found  
528 between the relative proportions of components AG3 and AG4 to  $\Sigma_8$  concentration  
529 ( $R^2 = 0.18$ ;  $p < 0.02$ ) but again was significantly stronger across the Kolyma and Lena  
530 Rivers in particular ( $R^2 = 0.79$  ;  $p < 0.001$ ). FI values positively correlated with  
531 increasing C/V ratios ( $R^2 = 0.48$ ,  $p < 0.001$ ,  $n = 30$ ) confirming losses in the  
532 proportion of woody tissues with increased autochthonous or less aromatic DOM  
533 supply.

534

### 535 **4.3 Linking optical properties to Arctic river DOM composition**

536 Underlying patterns and relationships between optical DOM parameters, DOC, and  
537 lignin were further explored using principle component analysis (PCA), which can  
538 identify the structure of data that best explains the variance within the dataset. The  
539 optical properties of DOM varied with season across all rivers, as demonstrated by  
540 PCA plots containing PARAFAC fluorescence components (percent contribution) and  
541 spectral slope information. The addition of FI and  $S_R$  values added little additional  
542 information to the PCA analyses. Furthermore,  $SUVA_{254}$  followed identical patterns  
543 to each of the spectral slope parameters and its inclusion led to similar PCA plots.  
544 These indices were therefore omitted from the final PCA model for clarity. Three  
545 principle components (PCs; eigenvalue  $> 1$ ) were identified that together explained  
546 80% of the total variance in the optical data (PCopt 1-3). PC1opt was related to  
547 increasing fluorescence contributions from AG3, AG1, and AG5, but decreasing  
548 contributions from AG6, AG4, and AG2 (Fig. 4). Components AG1, 3, and 5  
549 represent DOM fluorescence signatures that have all previously been reported to be

550 susceptible to microbial processing, or to be a byproduct of vascular material  
551 degradation (Table 3 and references herein). These fluorescence signatures may  
552 therefore represent indicators of ‘degraded’ or processed humic-like components. In  
553 contrast, components AG 2, 4, and 6 appear to represent more unreactive and stable  
554 components, previously being described as refractory in nature and shown not to co-  
555 vary with bacterial production (Table 3). PC1opt may therefore reflect potential  
556 reactivity or be an indicator of prior DOM processing. PC2opt appeared to be related  
557 to the shifting molecular weight of DOM, as indicated by strong relationships with  
558 changes in all spectral slopes (and SUVA<sub>254</sub>, not shown), whereas PC3opt was  
559 positively related to increased protein-like or phenolic DOM (AG7) and decreasing  
560 contributions from humic-like DOM (AG2 and AG1). PCA models run with only  
561 PARAFAC components contained two principle components, each indistinguishable  
562 from PC1opt and PC3opt, demonstrating that information on DOM potential  
563 reactivity and the relative contribution of protein-like versus humic-like could be  
564 obtained from fluorescence measurements alone.

565 Seasonal changes in DOM composition across all six rivers were most clearly  
566 separated along the PC2opt axis, with spring waters containing higher molecular  
567 weight material with shallower spectral slopes than summer and winter month waters.  
568 Positive scores on PC3opt during spring and winter months, relative to summer,  
569 suggest greater contributions of protein-like or phenolic material (as inferred by the  
570 proportion of component AG7), potentially representing reductions in allochthonous  
571 supply or increased export of fresh organics from surface layers, respectively. No  
572 clear separation among the six different rivers was apparent with optical properties  
573 alone across any of the PC axes.

574 To examine if the observed trends in DOM optical properties were related to  
575 geochemical changes in organic matter we conducted a separate PCA incorporating  
576 all lignin phenol and DOC fractionation variables. We subsequently compared the  
577 identified PCs with those extracted from optical measurements alone. Two PCs  
578 (PCmol 1-2) were identified, in combination explaining 67 % of the total variance in  
579 geochemical composition (Fig. 5). PC1mol positively related to increasing HPOA,  $\Sigma_8$ ,  
580 and  $\Lambda_8$  contributions and negatively with C/V ratio and proportions of the  
581 hydrophobic neutral and hydrophilic fractions. The axis therefore primarily separates  
582 seasonal variability observed in DOC, with spring months delivering greater  
583 proportions of HPOA with high concentrations of  $\Sigma_8$  and  $\Lambda_8$  values. PC2mol most  
584 strongly correlated to (Ad/Al) ratios suggesting it represented changing proportions of  
585 DOM degradation state. S/V ratio was also positively related with PC2mol, indicating  
586 that shifts in the relative proportions of sources waters may also be represented by this  
587 axis or similar processes (e.g. leaching and sorption) may be driving the observed S/V  
588 and (Ad/Al) ratios.

589 Comparing separate PCs from both optical and geochemical PCAs across all  
590 sites and sampling dates, only a single significant positive correlation was observed  
591 between PC2opt extracted from optical characteristics and PC1mol from DOC and  
592 lignin composition ( $r^2 = 0.65$ ,  $p < 0.001$ ,  $n = 30$ ; Fig. 5). Thus, seasonal variability in  
593 DOC composition was explained by relatively simple CDOM slope (and  $SUVA_{254}$ )  
594 metrics. Separating over seasons, PC1opt correlated significantly with PC2mol during  
595 the summer ( $r^2 = 0.85$ ,  $p < 0.05$ ,  $n = 8$ ) and winter months ( $r^2 = 0.65$ ,  $p = 0.06$ ,  $n = 6$ ),  
596 but not over spring periods across all six rivers. This further suggests that shifts in the  
597 relative proportions of PARAFAC components reflect shifts in the relative  
598 degradation signature of DOC inferred from acid:aldehyde ratios during certain

599 periods of the year. Information pertaining to the source (S/V ratios) of DOM may  
600 therefore also be contained in the relative ratio of more or less reactive or degraded  
601 PARAFAC components. Interestingly, (Ad/Al)<sub>v</sub> ratios have also been shown to  
602 correlate with the average <sup>14</sup>C age of DOC within these Arctic rivers (Amon et al.,  
603 2012). Fluorescence measurements may therefore provide information pertaining to  
604 both the age and degradation history of DOM across Arctic systems. Broad patterns in  
605 the temporal variability of DOM composition over pan-arctic scales therefore appear  
606 best captured using simple CDOM spectral slope and SUVA<sub>254</sub> measurements.  
607 Information on DOM processing, source, and age may instead be contained within  
608 CDOM fluorescence spectra and the relative contributions of PARAFAC components.

609

#### 610 **4.4 Modeling terrestrial biomarkers with optical measurements**

611 We ran a series of multiple linear regression models with the aim of predicting  $\Lambda_8$ ,  
612 C/V, S/V, (Ad/Al)<sub>s</sub>, and (Ad/Al)<sub>v</sub> across all sampling dates and rivers. Incorporating  
613 absorbance ( $a_{350}$ , slope ratios, SUVA<sub>254</sub>,) and fluorescence optical measurements (FI,  
614 % PARAFAC component contributions) as potential parameters,  $\Lambda_8$  ( $R^2 = 0.76$ ; Fig. 6  
615 a), C/V ( $R^2 = 0.70$ ; Fig. 6 b) and S/V values ( $R^2 = 0.68$ , Fig. 6 c) could be  
616 successfully predicted by model fits (all  $p < 0.001$ ;  $n=31$ ). Modeled values for  
617 (Ad/Al)<sub>v</sub> were also strongly correlated with observed values ( $p < 0.001$ ), but  
618 predictive capability was low ( $R^2 = 0.49$ , not shown). Model fits failed to accurately  
619 predict the variability in the (Ad/Al)<sub>s</sub> ratios ( $p > 0.05$ ).

620 The optical parameters providing the greatest predictive power varied between  
621 each lignin parameter. Models predicting  $\Lambda_8$  only incorporated  $S_{275-295}$  and  $a_{350}$  values  
622 ( $\Lambda_8 = 1.136 \pm 0.250 + (55.742 \pm 14.152 * S_{275-295}) + (0.006 \pm 0.003 * a_{350})$ ). Modeling  
623 C/V values also used  $S_{275-295}$  and  $a_{350}$  values (alone explaining 58% of the variance),

624 but improved with inclusion of %AG7 and  $S_{290-350}$  values ( $C/V = -0.153 \pm 0.127-$   
625  $(57.265 \pm 17.921 * S_{275-295}) + (0.040 \pm 0.015 * \%AG7) + (45.225 \pm 18.604 * S_{290-350}) -$   
626  $(0.002 * a_{350})$ ). Models explaining S/V values incorporated  $S_{290-350}$ ,  $a_{350}$ , %AG7, and FI  
627 ( $S/V = -2.704 \pm 0.827 + (64.131 \pm 14.843 * S_{290-350}) + (2.896 \pm 0.695 * FI) - (0.009$   
628  $\pm 0.002 * a_{350}) + (0.074 \pm 0.027 * \%AG7)$ ). S/V could not be explained with absorbance  
629 measurements alone. The ability to predict lignin composition as well as  
630 concentration using fluorescence measurements has previously been reported using  
631 partial least squares models of samples collected over a two year period on the  
632 Sacramento River/ San Joaquin River Delta, California (Hernes et al., 2009). The  
633 authors demonstrated that the most significant predictive capability for lignin was  
634 within the commonly referred to protein-like fluorescence region (similar to our  
635 component AG7). Fluorescence of propylphenol monomers, that structurally comprise  
636 lignin, can generate fluorescence signatures in a similar region to amino acids and in  
637 the region known as ‘protein-like’, thus our results may indicate information obtained  
638 from changes in phenolics rather than amino acid or proteins (Hernes et al., 2009).  
639 Therefore, it seems that rapid, inexpensive optical measurements may be capable of  
640 acting as a proxy for dissolved lignin compositional parameters as well as  
641 concentration across pan-arctic scales and catchments. The combination of  
642 absorbance and fluorescence metrics can also add predictive power when attempting  
643 to predict shifts in the composition of terrigenous DOC.

644

#### 645 **4.4 Improving terrigenous OC export estimates**

646 The absorbance coefficient at 350nm ( $a_{350}$ ) has previously been shown to be a  
647 sensitive and inexpensive proxy for lignin phenol concentration across a range of  
648 freshwater environments within Arctic river basins (Spencer et al., 2008; 2009;

649 Stedmon et al., 2011a). Furthermore, increased sampling frequency of Arctic rivers  
650 has led to significantly higher and better constrained DOC export estimates,  
651 particularly after the inclusion of samples from across the spring freshet period (e.g.  
652 Köhler et al., 2003; Striegl et al., 2005; Holmes et al., 2012). Here, we investigate if  
653 the combination of a lignin proxy with high-resolution monitoring of  $a_{350}$  over Arctic  
654 river hydrographs may be used to develop improved estimates of pan-arctic  
655 terrigenous DOC export, hereby refining land-to-ocean carbon flux estimates.

656 CDOM-derived lignin phenol concentrations (lignin<sub>350</sub>) were calculated using  
657 the linear regression of  $\Sigma_8$  and  $a_{350}$  (Fig. 3b). Lignin<sub>350</sub> values were derived from  $a_{350}$   
658 measurements taken from waters collected over the main Arctic-GRO sampling  
659 campaign and additional high-resolution samples taken over the freshet hydrographs.  
660 Inclusion of near-daily absorbance measurements collected over the peak discharge  
661 period alongside measurements spanning the entire year was crucial in adequately  
662 constraining fluxes during the spring freshet, when the majority of annual lignin  
663 export is expected (Amon et al., 2012; Spencer et al., 2008). Lignin<sub>350</sub> concentrations  
664 calculated for samples with concurrent  $\Sigma_8$  measurements were highly correlated across  
665 all Arctic rivers ( $r^2 = 0.92$ ,  $p < 0.01$ ,  $n = 31$ , Standard error of estimate,  $SE_E = 8.8\%$ )  
666 demonstrating the robust nature of this approach.

667 Daily model loads (mass d<sup>-1</sup>) of  $\Sigma_8$  in each river were calculated using a  
668 hydrologic load estimation model (LOADEST) integrating the lignin<sub>350</sub>  
669 concentrations and twelve years of daily discharge data ranging from 1999 – 2010  
670 (see Methods). Estimated lignin loads varied from 4.0 Gg y<sup>-1</sup> in the Mackenzie to 43.1  
671 Gg y<sup>-1</sup> in the Lena River (Table 5). The Lena, Yenisey, and Ob' Rivers export > 85 %  
672 of the total annual lignin discharge from the six largest Arctic rivers, a proportion that  
673 is very similar to that found by Amon et al., (2012). Flux estimates using lignin



674 phenol concentrations measured using identical methods and approaches compared  
675 well. Our mean annual Yukon River lignin flux derived for 2001-2009 ( $5.4 \pm 1.7$  Gg  
676  $y^{-1}$ ; Table 5) is similar to previous estimates of  $5.3 \pm 1.3$  Gg  $y^{-1}$  independently derived  
677 from measurements from 2004-2005 (Spencer et al., 2009), confirming the modeling  
678 approach is reproducible and robust. Our estimated lignin loads from all six major  
679 arctic rivers ( $98.4$  Gg  $yr^{-1}$ ) were, however, almost half of the  $192.0$  Gg  $yr^{-1}$  reported by  
680 Amon et al., (2012) for the same rivers from 2003 to 2007. These differences were  
681 primarily due to the significantly higher lignin concentrations ( $\Sigma_8$ ) reported by Amon  
682 et al., (2012) versus those of Spencer et al., (2009) and reported here, demonstrating  
683 the necessity for more standardization and intercomparison across lignin phenol  
684 measurements to ensure comparable datasets across studies.

685 Freshwater fluxes were scaled to the unsampled proportion of the Arctic using  
686 two published estimates of total Arctic Ocean watershed area. The smallest estimate  
687 (PA1; black line Fig. 1) spans an area of  $16.8 \times 10^6$   $km^2$ , where the largest (PA2; red  
688 line; Fig. 1) encompassing Hudson Bay drainage covers an area of  $20.5 \times 10^6$   $km^2$   
689 (Hernes et al., 2014). Pan-arctic lignin fluxes were estimated to span between  $155.5$   
690 Gg  $y^{-1}$  (PA1) to  $185.3$  Gg  $y^{-1}$  (PA2; Table 5) across these two geographic regions.

691 Dissolved lignin concentrations have previously been applied as a tracer of  
692 terrigenous DOM to the Arctic Ocean (Benner et al., 2005; Fichot et al., 2013; Opsahl  
693 et al., 1999) and used to estimate turnover rates of terrigenous DOC in the ocean  
694 (Hernes and Benner, 2006; Opsahl et al., 1999). Applying our pan-arctic flux (derived  
695 using our lignin<sub>350</sub> proxy) and assuming Arctic Ocean lignin concentrations ranging  
696 between  $84$  to  $320$   $ng\ L^{-1}$  (Opsahl et al., 1999), we calculate the residence time of  
697 terrigenous DOC in polar surface waters to be in the order of 7 months to 2.5 years.  
698 This compares well, yet slightly shorter than residence time estimates of  $<1$  to 4 years

699 calculated with comparable freshwater fluxes but scaled from the Yukon River alone  
700 (Spencer et al., 2009). Assuming the export of lignin phenol concentrations twice as  
701 high, similar to those reported by Amon et al., (2012), would result in even faster  
702 residence time estimates of < 4 months to 1 year. Overall, the short timeframes  
703 identified by these studies indicate either rapid losses of terrigenous DOC, via  
704 microbial, photochemical, or flocculation processes, or faster physical transport from  
705 Arctic Ocean waters to the North Atlantic than previously thought.

706

## 707 **5. Conclusions**

708 Employing optical techniques can increase the temporal and spatial coverage of DOM  
709 measurements across arctic river systems, shedding light on future changes in the  
710 composition and concentration of exported DOM, and help to more accurately  
711 estimate the amount and timing of terrigenous DOC flux. Here, river-specific  
712 relationships between  $a_{350}$  and DOC concentrations are presented and attributed to the  
713 export of varying proportions of non-chromophoric DOM from arctic catchments. We  
714 show that simple absorbance proxies ( $a_{350}$ ,  $S_{275-295}$ ), which can be measured with *in-*  
715 *situ* techniques, are capable of tracing dissolved lignin concentrations ( $\Sigma_8$ ) and  
716 seasonal changes in geochemical DOM composition (e.g.  $\Lambda_8$  and percent HPOA)  
717 occurring across the six major arctic rivers. Furthermore, we demonstrate that lignin  
718 phenol biomarkers appear capable of providing information on the biogeochemical  
719 cycling of the hydrophobic DOC fraction, thus knowledge on a major proportion of  
720 the aquatic DOM pool. More complex fluorescence DOM measurements followed by  
721 PARAFAC decomposition provided few direct pan-arctic proxies of DOM  
722 concentration or composition. However, the proportion of fluorescence signatures  
723 previously attributed to microbial processing or suggested to be by-products of

724 vascular material degradation co-varied with lignin monomer ratios over much of the  
725 year, suggesting these optical measurements may be capable of offering insights into  
726 changing DOM degradation state and source. Combining fluorescence and absorbance  
727 indices further strengthened our ability to predict DOM composition; in particular,  
728 inclusion of fluorescence index and protein-like contributions with absorbance  
729 coefficient and spectral slope measurements enabled predictive models of lignin  
730 ratios, suggesting potential to distinguish DOM source characteristics. Finally, we  
731 combine our pan-arctic relationship between  $a_{350}$  and  $\Sigma_8$  with high resolution  
732 monitoring of  $a_{350}$  to develop more accurately constrained residence times for  
733 terrigenous DOC in the Arctic Ocean of between 7 months to 2 ½ years. Optical  
734 measurements can provide key insights into the flux and biogeochemical cycling of  
735 terrigenous DOC in the Arctic, which will prove critical for understanding how  
736 carbon budgets and fluxes alter under future climate change scenarios.

### 737 738 **Acknowledgments**

739 This work was supported by grants from the National Science Foundation for the  
740 Global Rivers Project (0851101), the PARTNERS Project (0229302), the Arctic Great  
741 Rivers Observatory I & II (0732522 and 1107774), Detecting the Signature of  
742 Permafrost Thaw in Arctic Rivers (1203885 and 1500169), and the US Geological  
743 Survey's National Research Program. This work was aided by an NSF equipment  
744 grant to upgrade the analytical facilities at the North-East Science Station (0938254).  
745 We thank Alexander Shiklomanov for providing additional river discharge data, and  
746 the field staff of the USGS Alaska Science Center for their sample collection efforts.  
747 Special thanks to Ekaterina Bulygina and Lydia Russell-Roy for help running  
748 laboratory analyses. Also thanks to Greg Fiske for production of Figure 1. Any use of  
749 trade, firm, or product names is for descriptive purposes only and does not imply

750 endorsement by the U.S. Government.

751

## 752 **Figure legends**

753

754 **Figure 1.** Map showing the six Arctic river catchments sampled. Black dots indicate  
755 sampling locations. The thick black (PA1) and red lines (PA2) represent two  
756 estimates of the total pan-arctic watershed area ( $16.8 \times 10^6 \text{ km}^2$  and  $20.5 \times 10^6 \text{ km}^2$ ,  
757 respectively).

758

759 **Figure 2.** Seven independent fluorescent components (AG1-7) identified using  
760 PARAFAC analysis of 645 excitation-emission matrices. Excitation and emission  
761 peak positions are reported alongside descriptions in Table 3.

762

763 **Figure 3. Panel A:** Dissolved organic carbon (DOC) concentration versus the CDOM  
764 absorbance coefficient at  $a_{350}$  nm from across the hydrograph of the six Arctic rivers.  
765 **Panel B:** Lignin phenol concentration ( $\Sigma_8$ ) versus the CDOM absorbance coefficient  
766 at  $a_{350}$  nm from across the hydrographs of the six Arctic rivers. Black line represents  
767 the linear regression of the data. Thin grey lines represent the extent of the prediction  
768 of fit (95 % confidence limit).  $SS_E$  indicates the error of the sum of squares providing  
769 an indication of variation around the fit.

770

771 **Figure 4. Panel A:** Principle component scores for optical properties across PC1opt  
772 and PC2opt for all optical samples labelled by season. **Panel B:** PC scores of each  
773 optical parameter across PC1opt and PC2opt. **Panel C:** Principle component scores  
774 across PC2opt and PC3opt for all optical samples labelled by season. **Panel D:** PC

775 scores of each optical parameter across PC2opt and PC3opt. FI and SUVA<sub>254</sub> values  
776 were not included in the final optical PCA, see text for details.

777

778 **Figure 5. Panel A:** Principle component scores for geochemical composition (lignin  
779 phenol and DOC fractionation variables) across PC1mol and PC2mol for all  
780 compositional measurements labelled by season. **Panel B:** PC scores of each  
781 compositional parameter across PC1mol and PC2mol. **Panel C:** Principle component  
782 scores across PC1mol and PC2mol for all compositional measurements labelled by  
783 river. **Panel D:** Relationship between PC2opt derived from optical data alone and  
784 PC1mol explaining compositional measurements. Black line represents the linear  
785 regression ( $r^2 = 0.65$ ,  $p < 0.001$ ).

786

787 **Figure 6.** Model predicted values against measured measurements of: **Panel A:**  
788 Carbon normalized sum of the lignin phenols ( $\Lambda_8$ ; mg (100 mg OC)<sup>-1</sup>), **Panel B:**  
789 Cinnamyl to vanillyl phenol ratios (C/V), and **Panel C:** Syringyl to vanillyl phenol  
790 ratios (S/V).

Table 1. Total dissolved organic carbon concentrations (DOC) and major chemical fractions of DOC and fraction-specific ultraviolet absorbance (SUVA<sub>254</sub>) across the six major arctic rivers (mean ± standard error) during Spring (May and June), Summer (July through to October) and Winter (November through to April). Hydrophobic acids (HPOA), transphilic acids (TPI) and hydrophilic organic matter (HPI) presented as percentage of total DOC concentrations and the sample mass of each fraction (HPON comprises the remaining <10 % of the DOC pool).

Site	Season	Total DOC (mgC <sup>-1</sup> )	Total SUVA <sub>254</sub> (L mgC <sup>-1</sup> m <sup>-1</sup> )	HPOA (%)	HPOA SUVA <sub>254</sub> (L mgC <sup>-1</sup> m <sup>-1</sup> )	TPIA (%)	TPIA SUVA <sub>254</sub> (L mgC <sup>-1</sup> m <sup>-1</sup> )	HPI (%)	HPI SUVA <sub>254</sub> (L mgC <sup>-1</sup> m <sup>-1</sup> )
Kolyma	Spring	10.8 ± 1.7	2.9 ± 0.1	54 ± 2	3.9 ± 0.1	16 ± 1	2.4 ± 0.0	21 ± 1	1.5 ± 0.2
	Summer	3.7 ± 0.2	2.5 ± 0.0	56 ± 7	3.4 ± 0.2	18 ± 0	2.2 ± 0.0	20 ± 2	1.7 ± 0.2
	Winter	4.3 ± 1.7	2.0 ± 0.1	50 ± 7	2.9 ± 0.1	18 ± 0	2.0	20	2.1
Lena	Spring	15.7 ± 0.9	3.7 ± 0.0	57 ± 1	4.3 ± 0.0	17 ± 0	2.9 ± 0.0	17 ± 0	1.7 ± 0.0
	Summer	7.4 ± 0.6	2.8 ± 0.4	52 ± 3	4.0 ± 0.4	16 ± 1	2.5 ± 0.1	18 ± 0	1.5 ± 0.1
	Winter	9.7 ± 2.0	2.6 ± 0.3	53 ± 1	3.7 ± 0.1	17 ± 1	2.6 ± 0.1	16 ± 0	1.7 ± 0.2
Mackenzie	Spring	4.7 ± 0.3	2.5 ± 0.6	47 ± 1	3.6 ± 0.2	19 ± 0	2.4 ± 0.1	20 ± 1	1.4 ± 0.1
	Summer	5.4 ± 0.5	2.3 ± 0.1	46 ± 2	3.8 ± 0.2	20 ± 1	2.4 ± 0.1	20 ± 1	1.3 ± 0.0
	Winter	5.1	1.5	40	3.2	16	2.2	24	1.2
Ob'	Spring	9.0 ± 0.4	3.4 ± 0.0	57 ± 1	4.1 ± 0.0	15 ± 1	2.8 ± 0.1	18 ± 1	1.8 ± 0.1
	Summer	11.4 ± 0.9	3.1 ± 0.4	56 ± 1	4.2 ± 0.1	16 ± 1	2.8 ± 0.2	17 ± 1	-
	Winter	9.2 ± 1.9	3.0 ± 0.1	55 ± 3	4.0 ± 0.0	17 ± 1	2.8 ± 0.2	16 ± 1	-
Yenisey	Spring	10.1 ± 0.2	3.9 ± 0.0	60 ± 2	4.4 ± 0.1	16 ± 1	3.1 ± 0.1	16 ± 0	1.7 ± 0.0
	Summer	7.4 ± 1.8	2.8 ± 0.2	49 ± 3	4.2 ± 0.1	17 ± 1	2.6 ± 0.0	22 ± 2	1.5
	Winter	4.7 ± 0.7	2.6 ± 0.2	49 ± 6	3.9 ± 0.2	18 ± 1	2.5 ± 0.1	23 ± 3	2.3
Yukon	Spring	9.8 ± 2.3	3.2 ± 0.1	53 ± 1	4.2 ± 0.1	15 ± 0	2.8 ± 0.0	19 ± 1	2.2 ± 0.2
	Summer	7.1 ± 1.5	2.5 ± 0.2	50 ± 2	4.0 ± 0.4	19 ± 1	2.6 ± 0.1	21 ± 2	1.9 ± 0.2
	Winter	2.9	2	43	3.3	18	2.1	22	2.2

Table 2. Sampling location and date of discharge (Q), dissolved organic carbon (DOC) and lignin phenol concentration ( $\Sigma_8$ ), carbon normalized sum of lignin yields ( $\Lambda_8$ ), lignin ratios (C/V and S/V) and acid aldehyde ratios (Ad/Al) measurements; ratios of vanillic acid to vanillin (Ad/Al)<sub>v</sub> and ratios of syringic acid to syringaldehyde (Ad/Al)<sub>s</sub>. Site-normalized discharge (Q) is also presented.

River	Date	Q (m <sup>3</sup> s <sup>-1</sup> )	DOC (mgC L <sup>-1</sup> )	$\Sigma_8$ (μg L <sup>-1</sup> )	$\Lambda_8$ (mg(100 mg OC) <sup>-1</sup> )	C/V	S/V	(Ad/Al) <sub>v</sub>	(Ad/Al) <sub>s</sub>
Kolyma	05 Jun 2009	12,800	10.7	54.2	0.51	0.16	0.51	1.58	1.14
Kolyma	12 Jun 2009	11,100	9.1	45.1	0.50	0.18	0.58	1.42	1.00
Kolyma	21 Jun 2009	8,270	5.5	16.8	0.31	0.18	0.41	1.29	1.06
Kolyma	09 Sep 2009	8,390	3.9	17.0	0.44	0.19	0.64	1.51	1.08
Kolyma	08 Nov 2009	3,850	2.6	3.8	0.15	0.24	0.47	1.41	1.02
Lena	31 May 2009	68,328	17.5	120.0	0.69	0.04	0.17	1.28	0.98
Lena	05 Jun 2009	128,769	16.7	107.2	0.65	0.05	0.19	1.28	0.94
Lena	11 Jun 2009	83,800	12.9	85.2	0.66	0.07	0.25	1.30	0.96
Lena	22 Aug 2009	33,400	6.8	20.9	0.31	0.14	0.28	1.36	1.05
Lena	18 Nov 2009	4,102	7.7	16.5	0.22	0.12	0.30	1.34	1.09
Mackenzie	11 Jun 2009	24,300	4.3	16.2	0.38	0.17	0.34	1.01	0.81
Mackenzie	30 Jun 2009	20,900	5.3	9.3	0.18	0.28	0.50	1.24	1.38
Mackenzie	02 Jul 2009	21,100	6.6	8.3	0.13	0.19	0.43	0.98	0.89
Mackenzie	08 Sep 2009	13,800	5.0	8.0	0.16	0.18	0.37	1.06	1.00
Mackenzie	25 Mar 2010	4,380	5.1	5.5	0.11	0.32	0.51	1.09	1.13
Ob'	02 Jun 2009	36,300	8.1	55.5	0.69	0.23	0.61	1.39	0.93
Ob'	07 Jun 2009	36,100	8.5	40.4	0.48	0.19	0.58	1.39	0.89
Ob'	13 Jun 2009	35,400	7.7	55.7	0.73	0.19	0.59	1.38	0.88
Ob'	25 Aug 2009	13,500	10.5	22.4	0.22	0.20	0.46	0.93	0.90
Ob'	01 Dec 2009	5,152	7.3	17.2	0.24	0.31	0.68	0.97	0.75
Yenisey	17 Jun 2009	85,400	9.8	71.2	0.73	0.07	0.29	1.39	0.91
Yenisey	21 Jun 2009	67,200	9.5	74.7	0.79	0.08	0.29	1.40	0.86

Yenisey	27 Jun 2009	45,600	9.6	57.6	0.60	0.07	0.26	1.17	0.77
Yenisey	08 Aug 2009	15,200	5.6	18.7	0.33	0.15	0.37	0.99	0.80
Yenisey	30 Nov 2009	11,167	4.0	12.2	0.31	0.18	0.35	0.84	0.88
Yukon	14 May 2009	11,836	5.2	17.2	0.34	0.10	0.57	1.70	1.05
Yukon	20 May 2009	10,874	13.0	62.1	0.48	0.11	0.48	1.90	1.03
Yukon	26 May 2009	26,901	15.0	95.5	0.64	0.13	0.54	1.79	1.28
Yukon	07 Jul 2009	15,008	5.6	11.5	0.21	0.15	0.46	0.99	0.77
Yukon	18 Aug 2009	7,759	2.6	4.2	0.16	0.27	0.51	1.02	0.77
Yukon	12 Jan 2010	1,869	2.9	3.8	0.13	0.33	0.63	1.10	1.14

---



Table 3. Excitation and emission maxima ( $E_{x_{max}}$ /  $E_{m_{max}}$ ) of the seven components identified using parallel factor analysis of DOM fluorescence (Fig. 2). Description of previously identified components displaying similar optical properties (TCC > 0.95; see text for details). \*ID number refers to assigned study number in OpenFluor (<http://www.openfluor.org>).

Component	$E_{x_{max}}$ (nm)	$E_{m_{max}}$ (nm)	Comparable study ID* (component with TCC > 0.95)	Description
AG1	265	492	7 (C3), 8 (C2), 28 (C2), 31 (C4), 37 (C3), 44 (C2), 48 (C2), 54 (C2)	Humic-like fluorophore, terrigenous or autochthonous source, fulvic acid-like, present in all environments; Positively related to agriculture and bacterial production. Identified in many models and possibly formed as intermediate during photochemical degradation. Susceptible to microbial degradation.
AG2	270	448	29 (C5), 31 (C3), 69 (C4)	Similar to classical 'C peak'. Terrigenous component identified across a range of environments.
AG3	315	434	8 (C1)	Humic-like, emission spectrum identical to syringaldehyde (produced in breakdown of lignin) associated with waters containing high DOM loadings.
AG4	365	444	22 (C3), 34 (C1), 41 (C2), 47 (C2), 53 (C5), 55 (C3)	Similar to classical 'A peak', Terrigenous humic-like substances, refractory in nature.
AG5	320	392	8 (C4), 22 (C2), 26 (C6), 32 (C4), 47 (C3), 68 (C2)	Similar to classical 'M peak', marine and terrigenous humic material source, possibly derived from microbial reprocessing.
AG6	305	424	9 (C1), 28 (C3), 35 (C1), 44 (C3), 48 (C1), 54 (C3), 64 (C1)	Humic-like fluorophore, terrigenous; not correlated with land use or bacterial production
AG7	280	364	8 (C5), 9 (C7), 33 (C5), 34 (C7), 35 (C5), 39 (C5), 64 (C5)	Tryptophan-like associated with biological production in surface waters. Also a region known to be associated with phenolic fluorescence.

ID\*: <sup>7</sup>[Murphy et al., 2006], <sup>8</sup>[Murphy et al., 2014], <sup>9</sup>[Murphy et al., 2008], <sup>22</sup>[Kothawala et al., 2012], <sup>26</sup>[Stedmon et al., 2011b], <sup>28</sup>[Stedmon et al., 2007], <sup>29</sup>[Stedmon and Markager, 2005], <sup>31</sup>[Søndergaard et al., 2003], <sup>32</sup>[Jørgensen et al., 2011], <sup>33</sup>[Stedmon et al., 2003], <sup>34</sup>[Stedmon and Markager, 2005], <sup>35</sup>[Osburn and Stedmon, 2011], <sup>37</sup>[Walker et al., 2009], <sup>39</sup>[Yamashita et al., 2011], <sup>41</sup>[Yamashita et al., 2010a], <sup>44</sup>[Yamashita et al., 2010b], <sup>47</sup>[Kowalczyk et al., 2009], <sup>48</sup>[Graeber et al., 2012], <sup>53</sup>[Kothawala et al., 2013], <sup>54</sup>[Osburn et al., 2012], <sup>55</sup>[Osburn et al., 2011], <sup>64</sup>[Walker et al., 2013], <sup>68</sup>[Tanaka et al., 2014], <sup>69</sup>[Lapierre and del Giorgio, 2014].

Table 4. Linear regression fits for linear relationships between dissolved organic carbon concentration (DOC) and absorbance coefficient ( $a_{350}$ ) determined for each river.  $n$  represents number of measurements,  $R^2$  the coefficient of determination and  $SE_E$  the standard error of the mean. <sup>a</sup>Annual average discharge from each river from Holmes et al. (2012).

River	$n$	$R^2$	slope	intercept	$SE_E$ (%)	Discharge <sup>a</sup> ( $\text{km}^3 \text{y}^{-1}$ )
Kolyma	23	0.84	$0.478 \pm 0.045$	$0.845 \pm 1.022$	2.0	111
Lena	57	0.83	$0.333 \pm 0.020$	$3.429 \pm 0.666$	1.5	581
Mackenzie	60	0.86	$0.310 \pm 0.017$	$3.871 \pm 0.256$	0.9	298
Ob'	54	0.74	$0.295 \pm 0.024$	$2.240 \pm 0.531$	1.0	427
Yenisey	58	0.66	$0.231 \pm 0.022$	$3.793 \pm 0.528$	0.8	636
Yukon	49	0.95	$0.405 \pm 0.014$	$1.346 \pm 0.310$	1.0	208
ALL	301	0.81	$0.343 \pm 0.009$	$2.414 \pm 0.229$	1.6	2261

Provisional

Table 5. Total annual mean fluxes of CDOM-derived lignin (Lignin<sub>350</sub>) and DOC calculated using LOADEST. Arctic-GRO refers to the sum of all 6 rivers studied, and pan-arctic1 & 2 are the regions delineated in Figure 1. Mean annual discharge is calculated for the 1999 – 2010 period. <sup>#</sup>Averaged over 2001 – 2010.

River/ region	Watershed Area (10 <sup>6</sup> km <sup>2</sup> )	Discharge (km <sup>3</sup> y <sup>-1</sup> )	Lignin <sub>350</sub> (Gg y <sup>-1</sup> )		
			2009	2010	1999-2010
Kolyma	0.65	132	3.6	3.2	5.0 ± 1.2
Lena	2.4	591	50.5	36.2	43.1 ± 8.6
Mackenzie	1.75	319	5.2	3.5	4.0 ± 1.0
Ob'	2.95	421	15.5	16.2	18.6 ± 4.8
Yenisey	2.56	671	20.9	21.9	22.3 ± 2.3
Yukon	0.83	207	4.6	6.3	5.4 ± 1.7 <sup>#</sup>
Arctic-GRO	11.14	2342	100.3	87.3	98.4
pan-arctic1 (PA1)	16.8	3700	158.5	137.9	155.5
pan-arctic2 (PA2)	20.5	4410	188.9	164.4	185.3

Provisional

## References.

- Aagaard, K., and Carmack, E. C. (1989). The role of sea ice and other fresh water in the Arctic circulation. *Journal of Geophysical Research: Atmospheres (1984–2012)* 94, 14485–14498. doi:10.1029/JC094iC10p14485.
- Aiken, G. R., McKnight, D. M., Thorn, K. A., and Thurman, E. M. (1992). Isolation of hydrophilic organic acids from water using nonionic macroporous resins. *Organic Geochemistry* 18, 567–573. doi:10.1016/0146-6380(92)90119-I.
- Amon, R. M. W., Rinehart, A. J., Duan, S., Louchouart, P., Prokushkin, A., Guggenberger, G., et al. (2012). Dissolved organic matter sources in large Arctic rivers. *Geochimica et Cosmochimica Acta* 94, 217–237. doi:10.1016/j.gca.2012.07.015.
- Benner, R., Louchouart, P., and Amon, R. M. W. (2005). Terrigenous dissolved organic matter in the Arctic Ocean and its transport to surface and deep waters of the North Atlantic. *Global Biogeochem. Cycles* 19, GB2025 doi:10.1029/2004GB002398.
- Blough, N. V., and Del Vecchio, R. (2002). “Chromophoric DOM in the coastal environment,” in *Biogeochemistry of Marine Dissolved Organic Matter*, eds. D. A. Hansell and C. A. Carlson, 509–546.
- Blough, N. V., and Green, S. A. (1995). “Spectroscopic characterization and remote sensing of non-living organic matter,” in *The Role of Non-Living Organic Matter on the Earths Carbon Cycle*, eds. R. G. Zepp and C. Sonntag, 23–45.
- Booth, G. P., Raymond, P. A., and Oh, N. H. (2007) LoadRunner. Software and website. Yale University: New Haven. <http://research.yale.edu/environment/loadrunner/>.
- Coble, P. G. (1996). Characterization of marine and terrestrial DOM in seawater using excitation-emission matrix spectroscopy. *Marine Chemistry* 51, 325–346. doi:10.1016/0304-4203(95)00062-3.
- Coble, P. G. (2007). Marine optical biogeochemistry: the chemistry of ocean color. *Chem. Rev.* 107, 402–418. doi:10.1021/cr050350.
- Cory, R. M., Miller, M. P., and McKnight, D. M. (2010). Effect of instrument-specific response on the analysis of fulvic acid fluorescence spectra. *Limnology and Oceanography: Methods*. 67-78. doi:10.4319/lom.2010.8.67.
- Determann, S., Reuter, R., Wagner, P., and Willkomm, R. (1994). Fluorescent matter in the eastern Atlantic Ocean. Part 1: method of measurement and near-surface distribution. *Deep Sea Research Part I: Oceanographic Research Papers* 41, 659–675.
- Déry, S. J., Stahl, K., Moore, R. D., Whitfield, P. H., Menounos, B., and Burford, J. E. (2009). Detection of runoff timing changes in pluvial, nival, and glacial rivers

- of western Canada. *Water Resour. Res.* 45, W04426.  
doi:10.1029/2008WR006975.
- Fellman, J. B., Hood, E., and Spencer, R. G. M. (2010). Fluorescence spectroscopy opens new windows into dissolved organic matter dynamics in freshwater ecosystems: A review. *Limnol. Oceanogr.* 55, 2452–2462.  
doi:10.4319/lo.2010.55.6.2452.
- Fichot, C. G., Kaiser, K., Hooker, S. B., Amon, R. M. W., Babin, M., Bélanger, S., et al. (2013). Pan-Arctic distributions of continental runoff in the Arctic Ocean. *Sci. Rep.* 3, 1053. doi:10.1038/srep01053.
- Graeber, D., Gelbrecht, J., Pusch, M. T., Anlanger, C., and Schiller, von, D. (2012). Agriculture has changed the amount and composition of dissolved organic matter in Central European headwater streams. *Science of the Total Environment*, 438, 435–446. doi:10.1016/j.scitotenv.2012.08.087.
- Harshman, R. A., and Lundy, M. E. (1994). PARAFAC: Parallel factor analysis. *Computational Statistics & Data Analysis* 18, 39–72. doi:10.1016/0167-9473(94)90132-5.
- Hedges, J. I., and Ertel, J. R. (1982). Characterization of lignin by gas capillary chromatography of cupric oxide oxidation products. *Analytical Chemistry* 54, 174–178. doi:10.1021/ac00239a007.
- Hedges, J. I., and Mann, D. C. (1979). The characterization of plant tissues by their lignin oxidation products. *Geochimica et Cosmochimica Acta* 43, 1803–1807.
- Hedges, J. I., Clark, W. A., and Cowie, G. L. (1988). Organic matter sources to the water column and surficial sediments of a marine bay. *Limnol. Oceanogr.* 33, 1116–1136. doi:10.4319/lo.1988.33.5.1116.
- Helms, J. R., Stubbins, A., and Ritchie, J. D. (2008). Absorption spectral slopes and slope ratios as indicators of molecular weight, source, and photobleaching of chromophoric dissolved organic matter. *Limnology and Oceanography: Methods* 53, 955–969.
- Hernes, P. J., and Benner, R. (2003). Photochemical and microbial degradation of dissolved lignin phenols: Implications for the fate of terrigenous dissolved organic matter in marine environments. *J. Geophys. Res.* 108, 3291. doi:10.1029/2002JC001421.
- Hernes, P. J., and Benner, R. (2006). Terrigenous organic matter sources and reactivity in the North Atlantic Ocean and a comparison to the Arctic and Pacific oceans. *Marine Chemistry* 100, 66–79. doi:10.1016/j.marchem.2005.11.003.
- Hernes, P. J., Bergamaschi, B. A., Eckard, R. S., and Spencer, R. G. M. (2009). Fluorescence-based proxies for lignin in freshwater dissolved organic matter. *J. Geophys. Res.* 114, G00F03–10. doi:10.1029/2009JG000938.
- Hernes, P. J., Holmes, R. M., Raymond, P. A., Spencer, R. G. M., and Tank, S. E. (2014). “Fluxes, processing, and fate of riverine organic and inorganic carbon in

- the Arctic Ocean,” in *Biogeochemical Dynamics at Major River-Coastal Interfaces: Linkages with Global Change*, eds. T. S. Bianchi, M. A. Allison and W.-J. Cai, 530–553. doi:10.1017/CBO9781139136853.026.
- Hernes, P. J., Robinson, A. C., and Aufdenkampe, A. K. (2007). Fractionation of lignin during leaching and sorption and implications for organic matter “freshness.” *Geophysical Research Letters* 34, L17401–6. doi:10.1029/2007GL031017.
- Hernes, P. J., Spencer, R. G. M., Dyda, R. Y., Pellerin, B. A., Bachand, P. A. M., and Bergamaschi, B. A. (2008). The role of hydrologic regimes on dissolved organic carbon composition in an agricultural watershed. *Geochimica et Cosmochimica Acta* 72, 5266–5277. doi:10.1016/j.gca.2008.07.031.
- Holmes, R. M., McClelland, J. W., Peterson, B. J., Tank, S. E., Bulygina, E., Eglinton, T. I., et al. (2012). Seasonal and Annual Fluxes of Nutrients and Organic Matter from Large Rivers to the Arctic Ocean and Surrounding Seas. *Estuaries and Coasts* 35, 369–382. doi:10.1007/s12237-011-9386-6.
- Holmes, R. M., McClelland, J. W., Raymond, P. A., Frazer, B. B., Peterson, B. J., and Stieglitz, M. (2008). Lability of DOC transported by Alaskan rivers to the Arctic Ocean. *Geophysical Research Letters* 35, L03402–5. doi:10.1029/2007GL032837.
- Hu, C., Muller-Karger, F. E., and Zepp, R. G. (2002). Absorbance, absorption coefficient, and apparent quantum yield: A comment on common ambiguity in the use of these optical concepts. *Limnol. Oceanogr.* 47, 1261–1267. doi:10.4319/lo.2002.47.4.1261.
- Jørgensen, L., Stedmon, C. A., Kragh, T., Markager, S., Middelboe, M., and Søndergaard, M. (2011). Global trends in the fluorescence characteristics and distribution of marine dissolved organic matter. *Marine Chemistry* 126, 139–148. doi:10.1016/j.marchem.2011.05.002.
- Köhler, H., Meon, B., Gordeev, V. V., Spitzky, A., and Amon, R. M. W. (2003). “Dissolved organic matter (DOM) in the estuaries of Ob and Yenisei and the adjacent Kara Sea, Russia,” in *Siberian river run-off in the Kara Sea*, eds. R. Stein, K. Fahl, D. K. Fütterer, E. M. Galimov, and O. V. Stepanets.
- Kothawala, D. N., Stedmon, C. A., Müller, R. A., Weyhenmeyer, G. A., Köhler, S. J., and Tranvik, L. J. (2013). Controls of dissolved organic matter quality: Evidence from a large-scale boreal lake survey. *Global Change Biology* 20, 1101–1114. doi:10.1111/gcb.12488.
- Kothawala, D. N., Wachenfeldt, von, E., Koehler, B., and Tranvik, L. J. (2012). Selective loss and preservation of lake water dissolved organic matter fluorescence during long-term dark incubations. *Science of the Total Environment*, 433, 238–246. doi:10.1016/j.scitotenv.2012.06.029.
- Kowalczyk, P., Durako, M. J., Young, H., Kahn, A. E., Cooper, W. J., and Gonsior, M. (2009). Characterization of dissolved organic matter fluorescence in the South

- Atlantic Bight with use of PARAFAC model: Interannual variability. *Marine Chemistry* 113, 182–196. doi:10.1016/j.marchem.2009.01.015.
- Lakowicz, J. R. (2013). *Principles of Fluorescence Spectroscopy*. Springer Science & Business Media doi:10.1007/978-0-387-46312-4.
- Lapierre, J.-F., and del Giorgio, P. A. (2014). Partial coupling and differential regulation of biologically and photochemically labile dissolved organic carbon across boreal aquatic networks. *Biogeosciences* 11, 5969–5985. doi:10.5194/bg-11-5969-2014.
- Lawaetz, A. J., and Stedmon, C. A. (2009). Fluorescence intensity calibration using the Raman Scatter Peak of water. *Appl Spectrosc* 63, 936–940. doi:10.1366/000370209788964548.
- Lobbes, J. M., Fitznar, H. P., and Kattner, G. (2000). Biogeochemical characteristics of dissolved and particulate organic matter in Russian rivers entering the Arctic Ocean. *Geochimica et Cosmochimica Acta* 64, 2973–2983.
- Mann, P. J., Davydova, A., Zimov, N., Spencer, R. G. M., Davydov, S., Bulygina, E., et al. (2012). Controls on the composition and lability of dissolved organic matter in Siberia's Kolyma River basin. *J. Geophys. Res.* 117, G01028. doi:10.1029/2011jg001798.
- Mann, P. J., Eglinton, T. I., McIntyre, C. P., Zimov, N., Davydova, A., Vonk, J. E., et al. (2015). Utilization of ancient permafrost carbon in headwaters of Arctic fluvial networks. *Nature Communications* 6, 7856. doi:10.1038/ncomms8856.
- Mann, P. J., Spencer, R. G. M., Dinga, B. J., Poulsen, J. R., Hernes, P. J., Fiske, G., et al. (2014). The biogeochemistry of carbon across a gradient of streams and rivers within the Congo Basin. *J. Geophys. Res. Biogeosci.* 119, 687–702. doi:10.1002/(ISSN)2169-8961.
- Mathis, J. T., Hansell, D. A., and Bates, N. R. (2005). Strong hydrographic controls on spatial and seasonal variability of dissolved organic carbon in the Chukchi Sea. *Deep Sea Research Part II: Topical Studies in Oceanography* 52, 3245–3258. doi:10.1016/j.dsr2.2005.10.002.
- McClelland, J. W., Déry, S. J., Peterson, B. J., Holmes, R. M., and Wood, E. F. (2006). A pan-arctic evaluation of changes in river discharge during the latter half of the 20th century. *Geophysical Research Letters* 33, L06715–4. doi:10.1029/2006GL025753.
- McClelland, J. W., Holmes, R. M., Dunton, K. H., and Macdonald, R. W. (2011). The Arctic Ocean Estuary. *Estuaries and Coasts* 35, 353–368. doi:10.1007/s12237-010-9357-3.
- McClelland, J. W., Holmes, R. M., Peterson, B. J., Amon, R. M. W., Brabets, T., Cooper, L. W., et al. (2008). Development of a Pan-Arctic Database for River Chemistry. *Eos*. 89, 24, 217-218. doi:10.1029/2005JG000031.
- McKnight, D. M., Boyer, E. W., Westerhoff, P. K., Doran, P. T., Kulbe, T., and

- Andersen, D. T. (2001). Spectrofluorometric characterization of dissolved organic matter for indication of precursor organic material and aromaticity. *Limnol. Oceanogr.* 46, 38–48. doi:10.4319/lo.2001.46.1.0038.
- Murphy, K. R., Bro, R., and Stedmon, C. A. (2014). “Chemometric analysis of organic matter fluorescence,” in *Aquatic Organic Matter Fluorescence*, eds. P. Coble, J. Lead, A. Baker, D. Reynolds, and R. G. M. Spencer, 339–375. doi:10.13140/2.1.2595.8080.
- Murphy, K. R., Ruiz, G. M., Dunsmuir, W. T. M., and Waite, T. D. (2006). Optimized parameters for fluorescence-based verification of ballast water exchange by ships. *Environ. Sci. Technol.* 40, 2357–2362. doi:10.1021/es0519381.
- Murphy, K. R., Stedmon, C. A., Graeber, D., and Bro, R. (2013). Fluorescence spectroscopy and multi-way techniques. PARAFAC. *Anal. Methods* 5, 6557–6566. doi:10.1039/c3ay41160e.
- Murphy, K. R., Stedmon, C. A., Waite, T. D., and Ruiz, G. M. (2008). Distinguishing between terrestrial and autochthonous organic matter sources in marine environments using fluorescence spectroscopy. *Marine Chemistry* 108, 40–58. doi:10.1016/j.marchem.2007.10.003.
- Neff, J. C., Finlay, J. C., Zimov, S. A., Davydov, S. P., Carrasco, J. J., Schuur, E. A. G., et al. (2006). Seasonal changes in the age and structure of dissolved organic carbon in Siberian rivers and streams. *Geophysical Research Letters* 33, L23401. doi:10.1029/2006GL028222.
- O'Donnell, J. A., Aiken, G. R., Walvoord, M. A., and Butler, K. D. (2012). Dissolved organic matter composition of winter flow in the Yukon River basin: Implications of permafrost thaw and increased groundwater discharge. *Global Biogeochem. Cycles* 26, 4. doi:10.1029/2012GB004341.
- Opsahl, S., and Benner, R. (1995). Early diagenesis of vascular plant tissues: Lignin and cutin decomposition and biogeochemical implications. *Geochimica et Cosmochimica Acta* 59, 4889–4904. doi:10.1016/0016-7037(95)00348-7.
- Opsahl, S., Benner, R., and Amon, R. M. W. (1999). Major flux of terrigenous dissolved organic matter through the Arctic Ocean. *Limnol. Oceanogr.* 44, 2017–2023. doi:10.4319/lo.1999.44.8.2017.
- Osburn, C. L., Handsel, L. T., Mikan, M. P., Paerl, H. W., and Montgomery, M. T. (2012). Fluorescence Tracking of Dissolved and Particulate Organic Matter Quality in a River-Dominated Estuary. *Environ. Sci. Technol.* 46, 8628–8636. doi:10.1021/es3007723.
- Osburn, C. L., and Stedmon, C. A. (2011). Linking the chemical and optical properties of dissolved organic matter in the Baltic–North Sea transition zone to differentiate three allochthonous inputs. *Marine Chemistry* 126, 281–294. doi:10.1016/j.marchem.2011.06.007.
- Osburn, C. L., Wigdahl, C. R., Fritz, S. C., and Saros, J. E. (2011). Dissolved organic



- matter composition and photoreactivity in prairie lakes of the U.S. Great Plains. *Limnol. Oceanogr.* 56, 2371–2390. doi:10.4319/lo.2011.56.6.2371.
- Peterson, B. J. (2002). Increasing river discharge to the Arctic Ocean. *Science* 298, 2171–2173. doi:10.1126/science.1077445.
- Rawlins, M. A., Steele, M., Holland, M. M., Adam, J. C., Cherry, J. E., Francis, J. A., et al. (2010). Analysis of the Arctic system for freshwater cycle intensification: Observations and expectations. *J. Climate* 23, 5715–5737. doi:10.1175/2010JCLI3421.1.
- Raymond, P. A., McClelland, J. W., Holmes, R. M., Zhulidov, A. V., Mull, K., Peterson, B. J., et al. (2007). Flux and age of dissolved organic carbon exported to the Arctic Ocean: A carbon isotopic study of the five largest arctic rivers. *Global Biogeochem. Cycles* 21, 4. doi:10.1029/2007GB002934.
- Romanovsky, V. E., Drozdov, D. S., Oberman, N. G., Malkova, G. V., Kholodov, A. L., Marchenko, S. S., et al. (2010). Thermal state of permafrost in Russia. *Permafrost Periglac. Process.* 21, 136–155. doi:10.1002/ppp.683.
- Runkel, R. L., Crawford, C. G., and Cohn, T. A. (2004). LoadEstimator (LOADEST): A FORTRAN Program for estimating constituent loads in streams and rivers, In U.S. Geological Survey Techniques and Methods, Book 4, Chapter A5, 69
- Serreze, M. C., Barrett, A. P., Slater, A. G., Woodgate, R. A., Aagaard, K., Lammers, R. B., et al. (2006). The large-scale freshwater cycle of the Arctic. *J. Geophys. Res.* 111, C11010–19. doi:10.1029/2005JC003424.
- Smith, L. C., Pavelsky, T. M., MacDonald, G. M., Shiklomanov, A. I., and Lammers, R. B. (2007). Rising minimum daily flows in northern Eurasian rivers: A growing influence of groundwater in the high-latitude hydrologic cycle. *J. Geophys. Res.* 112, G04S47. doi:10.1029/2006JG000327.
- Søndergaard, M., Stedmon, C. A., and Borch, N. H. (2003). Fate of terrigenous dissolved organic matter (DOM) in estuaries: Aggregation and bioavailability. *Ophelia* 57, 161–176. doi:10.1080/00785236.2003.10409512.
- Spencer, R. G. M., Aiken, G. R., Butler, K. D., Dornblaser, M. M., Striegl, R. G., and Hernes, P. J. (2009). Utilizing chromophoric dissolved organic matter measurements to derive export and reactivity of dissolved organic carbon exported to the Arctic Ocean: A case study of the Yukon River, Alaska. *Geophysical Research Letters* 36, L06401–6. doi:10.1029/2008GL036831.
- Spencer, R. G. M., Aiken, G. R., Dyda, R. Y., Butler, K. D., Bergamaschi, B. A., and Hernes, P. J. (2010b). Comparison of XAD with other dissolved lignin isolation techniques and a compilation of analytical improvements for the analysis of lignin in aquatic settings. *Organic Geochemistry* 41, 445–453. doi:10.1016/j.orggeochem.2010.02.004.
- Spencer, R. G. M., Aiken, G. R., Wickland, K. P., Striegl, R. G., and Hernes, P. J. (2008). Seasonal and spatial variability in dissolved organic matter quantity and

- composition from the Yukon River basin, Alaska. *Global Biogeochem. Cycles* 22, 4. doi:10.1029/2008GB003231.
- Spencer, R. G. M., Butler, K. D., and Aiken, G. R. (2012). Dissolved organic carbon and chromophoric dissolved organic matter properties of rivers in the USA. *J. Geophys. Res.* 117, G03001–14. doi:10.1029/2011JG001928.
- Spencer, R. G. M., Hernes, P. J., Ruf, R., Baker, A., Dyda, R. Y., Stubbins, A., et al. (2010a). Temporal controls on dissolved organic matter and lignin biogeochemistry in a pristine tropical river, Democratic Republic of Congo. *J. Geophys. Res.* 115, G03013. doi:10.1029/2009JG001180.
- Spencer, R. G. M., Mann, P. J., Dittmar, T., Eglinton, T. I., McIntyre, C., Holmes, R. M., et al. (2015). Detecting the signature of permafrost thaw in Arctic rivers. *Geophysical Research Letters* 42, 2830–2835. doi:10.1002/2015GL063498.
- Stedmon, C. A., Amon, R., Rinehart, A. J., and Walker, S. A. (2011a). The supply and characteristics of colored dissolved organic matter (CDOM) in the Arctic Ocean: Pan Arctic trends and differences. *Marine Chemistry* 124, 108–118. doi:10.1016/j.marchem.2010.12.007.
- Stedmon, C. A., and Markager, S. (2005). Resolving the variability in dissolved organic matter fluorescence in a temperate estuary and its catchment using PARAFAC analysis. *Limnol. Oceanogr.* 50, 686–697. doi:10.4319/lo.2005.50.2.0686.
- Stedmon, C. A., Markager, S., and Bro, R. (2003). Tracing dissolved organic matter in aquatic environments using a new approach to fluorescence spectroscopy. *Marine Chemistry* 82, 239–254. doi:10.1016/S0304-4203(03)00072-0.
- Stedmon, C. A., Thomas, D. N., Granskog, M., Kaartokallio, H., Papadimitriou, S., and Kuosa, H. (2007). Characteristics of dissolved organic matter in Baltic coastal sea ice: Allochthonous or autochthonous origins? *Environ. Sci. Technol.* 41, 7273–7279. doi:10.1021/es071210f.
- Stedmon, C. A., Thomas, D. N., Papadimitriou, S., Granskog, M. A., and Dieckmann, G. S. (2011b). Using fluorescence to characterize dissolved organic matter in Antarctic sea ice brines. *J. Geophys. Res.* 116, G03027–9. doi:10.1029/2011JG001716.
- Striegl, R. G., Aiken, G. R., Dornblaser, M. M., Raymond, P. A., and Wickland, K. P. (2005). A decrease in discharge-normalized DOC export by the Yukon River during summer through autumn. *Geophysical Research Letters* 32, L21413. doi:10.1029/2005GL024413.
- Striegl, R. G., Dornblaser, M. M., Aiken, G. R., Wickland, K. P., and Raymond, P. A. (2007). Carbon export and cycling by the Yukon, Tanana, and Porcupine rivers, Alaska, 2001–2005. *Water Resour. Res.* 43, 2. doi:10.1029/2006WR005201.
- Tanaka, K., Kuma, K., Hamasaki, K., and Yamashita, Y. (2014). Accumulation of humic-like fluorescent dissolved organic matter in the Japan Sea. *Sci. Rep.* 4, 1–7.

doi:10.1038/srep05292.

Tarnocai, C., Canadell, J. G., Schuur, E. A. G., Kuhry, P., Mazhitova, G., and Zimov, S. (2009). Soil organic carbon pools in the northern circumpolar permafrost region. *Global Biogeochem. Cycles* 23, 2. doi:10.1029/2008GB003327.

Templier, J., Derenne, S., Croué, J.-P., and Largeau, C. (2005). Comparative study of two fractions of riverine dissolved organic matter using various analytical pyrolytic methods and a <sup>13</sup>C CP/MAS NMR approach. *Organic Geochemistry* 36, 1418–1442. doi:10.1016/j.orggeochem.2005.05.003.

Tucker, L. R. (1951). *A method for synthesis of factor analysis studies* (Personnel Research Section Report No. 984). Washington, DC: Department of the Army.

Walker, S. A., Amon, R. M. W., and Stedmon, C. A. (2013). Variations in high-latitude riverine fluorescent dissolved organic matter: A comparison of large Arctic rivers. *J. Geophys. Res. Biogeosci.*, 118, 4, 1689-1702. doi:10.1002/2013JG002320.

Walker, S. A., Amon, R. M. W., Stedmon, C. A., Duan, S., and Louchouart, P. (2009). The use of PARAFAC modeling to trace terrestrial dissolved organic matter and fingerprint water masses in coastal Canadian Arctic surface waters. *J. Geophys. Res.* 114, G00F06–12. doi:10.1029/2009JG000990.

Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R., and Mopper, K. (2003). Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. *Environ. Sci. Technol.* 37, 4702–4708. doi:10.1021/es030360x.

Wickland, K. P., Aiken, G. R., Butler, K., Dornblaser, M. M., Spencer, R. G. M., and Striegl, R. G. (2012). Biodegradability of dissolved organic carbon in the Yukon River and its tributaries: Seasonality and importance of inorganic nitrogen. *Global Biogeochem. Cycles* 26, 4. doi:10.1029/2012GB004342.

Yamashita, Y., Kloeppel, B. D., Knoepp, J., Zausen, G. L., and Jaffé, R. (2011). Effects of watershed history on dissolved organic matter characteristics in headwater streams. *Ecosystems* 14, 1110–1122. doi:10.1007/s10021-011-9469-z.

Yamashita, Y., Maie, N., Briceño, H., and Jaffé, R. (2010b). Optical characterization of dissolved organic matter in tropical rivers of the Guayana Shield, Venezuela. *J. Geophys. Res.* 115, G00F10–15. doi:10.1029/2009JG000987.

Yamashita, Y., Scinto, L. J., Maie, N., and Jaffé, R. (2010a). Dissolved organic matter characteristics across a subtropical wetland's landscape: Application of optical properties in the assessment of environmental dynamics. *Ecosystems* 13, 1006–1019. doi:10.1007/s10021-010-9370-1.

Figure 1.TIF



Figure 2.TIF

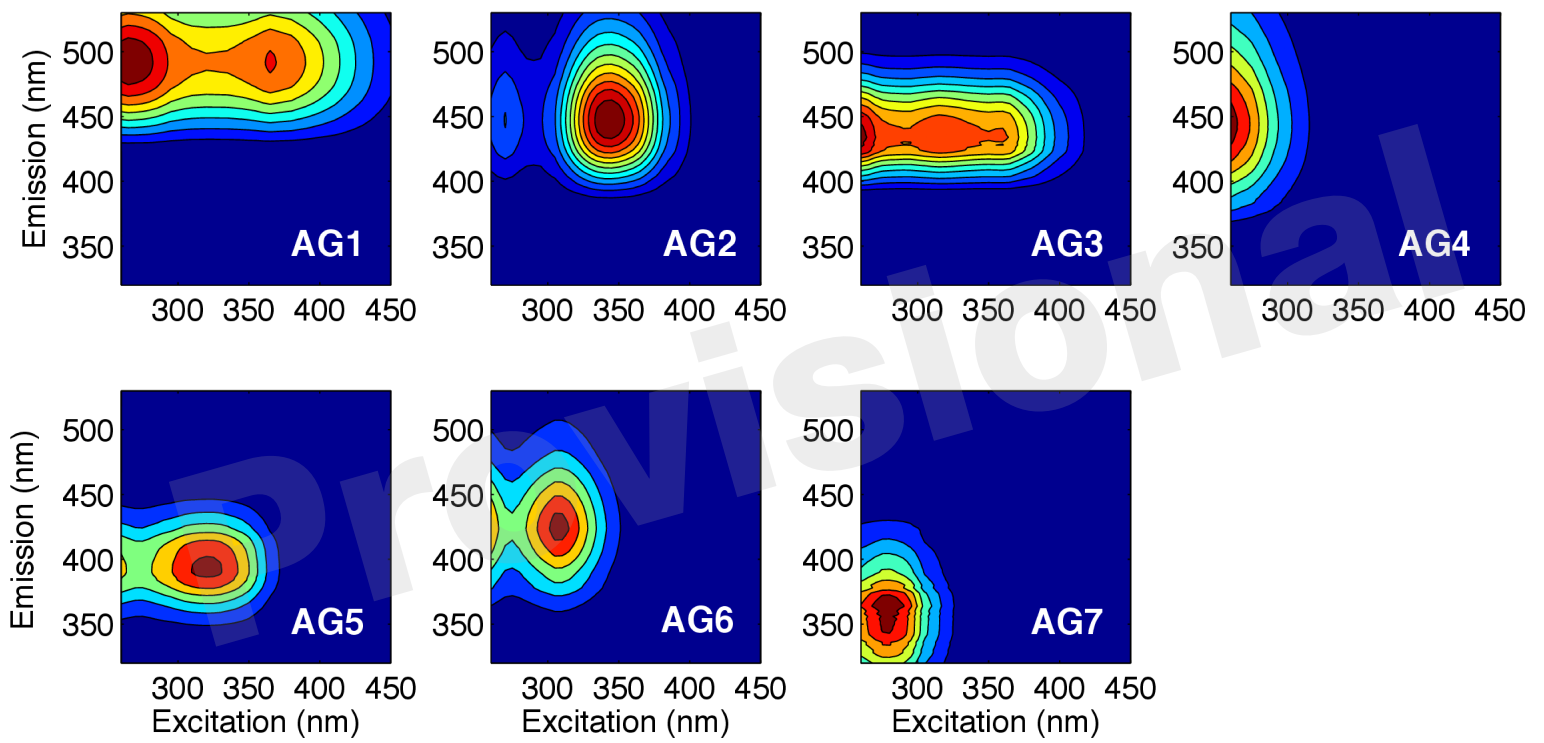


Figure 3.TIF

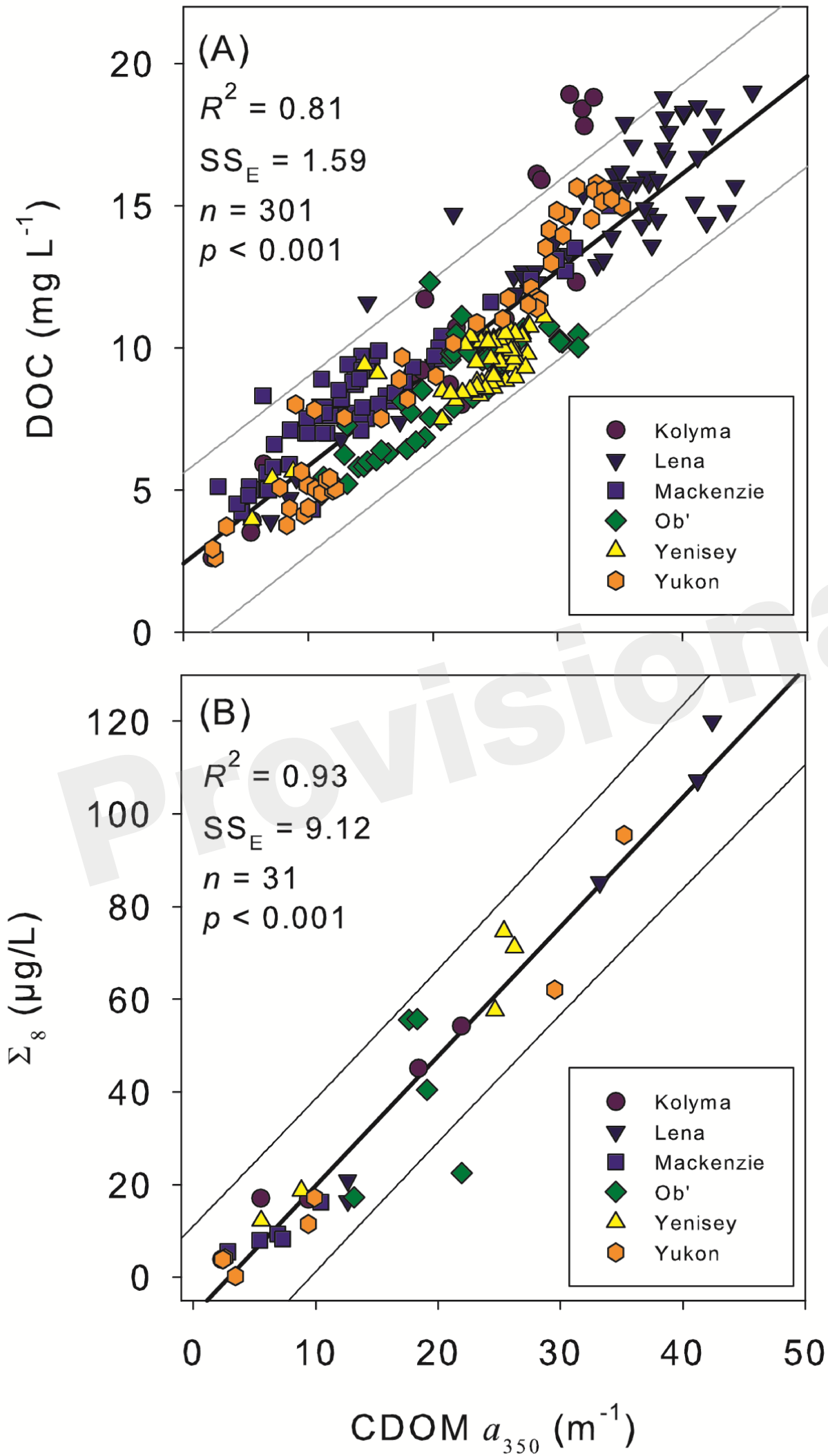


Figure 4.TIF

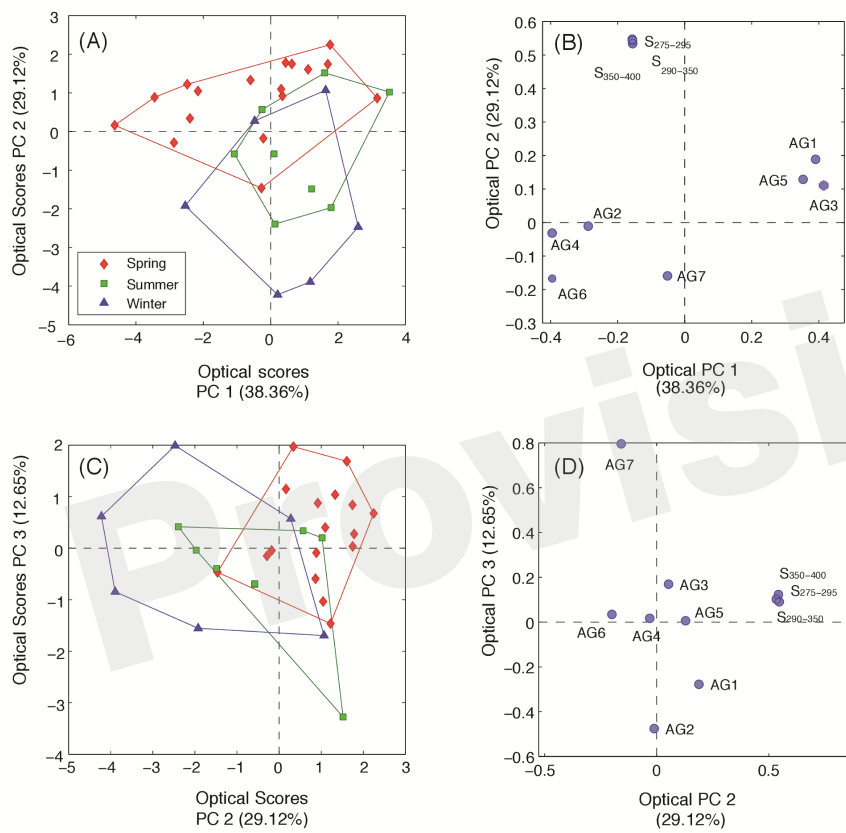


Figure 5.TIF

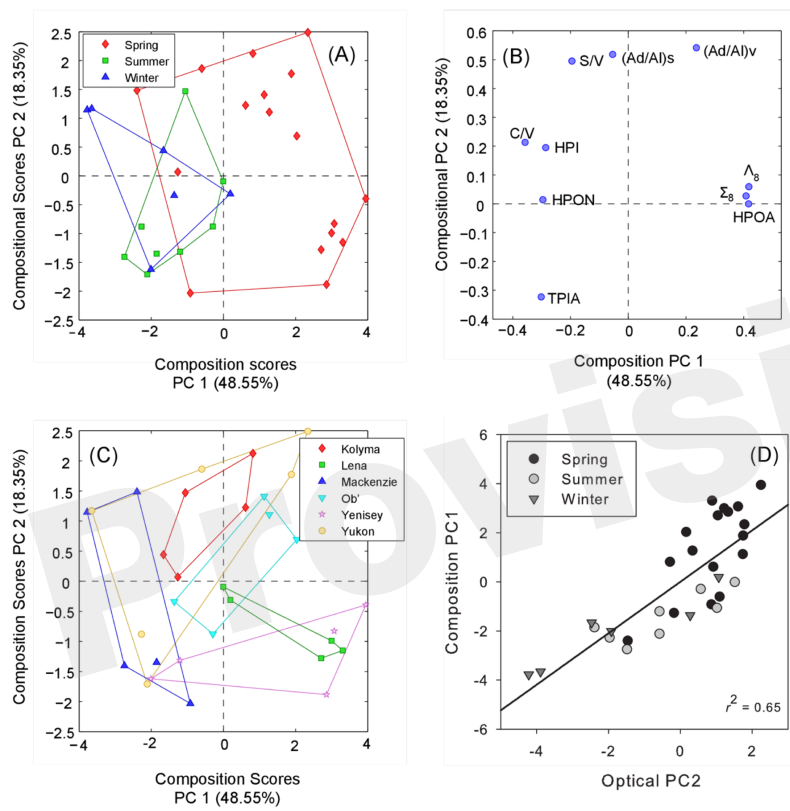




Figure 6.TIF

