Erosion-initiated stromatolite and thrombolite formation in a present-day coastal sabkha setting

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ABSTRACT

Laminated microbial mats and microbialites have been documented from a variety of coastal marine environments. This study aims to provide the first detailed descriptions of intertidal pools, along with their hosted thrombolite and stromatolite structures, from Abu Dhabi, and to propose a model for their formation and evolution. We propose that the development of pools within the upper intertidal zone was initiated by localised erosion of the laminated microbial mats during high energy events. The removal of the protective mats permitted erosion of the underlying unconsolidated sediment to produce erosional scours that continued to develop to produce the pools observed today. The margins of the newly-created submerged environment were colonised by a cyanobacteria dominated microbial community. The precipitation of aragonite cement, associated with the cyanobacteria, stabilised the pool walls and cemented the microbial communities to form stromatolitic and thrombolitic fabrics. Syndepositional cementation was further enhanced by the precipitation of marine cements as a result of evaporation-driven Ca$^{2+}$ and Mg$^{2+}$ supersaturation. Erosion behind and below the cemented pool wall eventually resulted in rim-collapse and the formation of the observed pool margin parallel thrombolite bands. Successive generations of lithification and erosion increased the area of the pool with the earliest thrombolites eroding and becoming increasingly isolated. In summary, the resultant microbialites developed through the complex interplay of erosion, abiotic early lithification and microbially-mediated processes and represent a continuum between unlithified laminated microbial mats and domal microbialites. These features are most likely produced during a sea level scenario of stillstand or transgression, and, as such, may be useful as a diagnostic tool to elucidate the onset of transgression. The newly proposed model for stromatolite formation has significant implications for the recognition and interpretation of similar structures observed in the fossil record.
KEYWORDS

sabkha, erosion, stromatolite, thrombolite, microbialite, lithification
INTRODUCTION

Stromatolites are laminated benthic microbial deposits (Riding, 1999) formed through lithification processes mediated or controlled by microbial communities of archaea, bacteria and/or diatoms (Dupraz et al., 2011). Stromatolites appear throughout much of the stratigraphic record, occurring from the Early Archean (Allwood et al., 2006) to the Recent with different organisms taking on the role of bioconstructors at different times. Extant stromatolites have been documented from a variety of coastal marine settings, the best well-known being the structures observed at Hamlin Pool, Shark Bay, Western Australia (Golubic, 1985; Jahnert and Collins, 2013; Logan, 1961; Reid et al., 2003) and from the Bahamas (Andres and Reid, 2006; Bowlin et al., 2012; Monty, 1972; Stolz et al., 2009). Microbial mats, thrombolites and stromatolites have also been described from numerous other coastal settings, including Lagoa Vermelha, Brazil (Vasconcelos et al., 2006), Cayo Coco lagoon, Cuba (Bouton et al., 2016), the Caicos Platform (Trembath-Reichert et al., 2016) and the coastal sabkhas of the Persian Gulf (Brauchli et al., 2016; Court et al., 2017; Evans, 1966; Kendall and Skipwith, 1968; Lokier et al., 2017).

Despite the long stratigraphic history and significant geographical range of these biologically-generated sedimentary structures, models for their mode of formation have been surprisingly limited to studies of Recent examples from only a limited number of depositional settings. Such a paucity of studies is even more surprising when one considers that in many ‘stressed’ environments the microbial communities associated with stromatolite and thrombolite formation were the dominant and, often, only biota interacting with sediments at the time of deposition. Succinctly, while stromatolites and thrombolites have a significant potential as a tool for paleoenvironmental interpretation, such utility requires that their mode of formation is considered from a range of depositional settings and potential distinguishing characteristics are identified.

STUDY AREA

The study area is located at the southern shore of the Gulf, approximately 40 km to the southwest of Abu Dhabi City (Fig. 1A, B). The Gulf is a shallow microtidal sea with an average water depth of 35 m and a tidal range of 1.5 m in open-marine areas and less than 1 m in lagoons. At the Abu Dhabi coastline, temperatures range between 7 °C at night during
the winter and exceed 50 °C during summer months (Lokier, 2012). Average annual rainfall
is 72 mm, predominantly occurring as brief, heavy, downpours during winter months (Raafat,
2007). Arid conditions result in evaporation (2.75 m per year) far exceeding precipitation
(Bottomley, 1996).

The southeast region of the Gulf comprises a low-angle, north-sloping, ramp geometry that
was rapidly flooded as post-glacial eustatic sea level rise transgressed an aeolian dune system
resulting in a transition from aeolian-siliciclastic to marine-carbonate dominated deposition
(Evans et al., 1969; Lambeck, 1996; Lokier and Steuber, 2008). A short-lived still stand,
between 7,100 - 6,890 cal yrs BP (Lokier et al., 2015), facilitated the development of coastal
sabkhas prior to renewed transgression with the onset of the Middle Holocene Atlantic stage.
The subsequent relative sea level fall and stillstand resulted in the development of a strongly
progradational geometry throughout the Middle to Late Holocene (Lokier and Steuber, 2008;
Lokier et al., 2015).

From seaward to landward, the surface facies of the modern sabkha (Fig. 1C) are
classified by peloidal carbonate sands in the lower and middle intertidal zone (lagoon), a
coast-parallel microbial mat belt in the upper intertidal zone, and evaporite precipitates in the
supratidal zone (Evans et al., 1969; Kendall and Skipwith, 1969). The spatial occurrence of
these facies belts is controlled by the local angle of slope which governs the duration of tidal
flooding (Court et al., 2017). The microbial mat belt varies in width between 150 to 800 m
and is classified (Court et al., 2017) based on surface morphologies resulting from the
duration of inundation.

**METHODOLOGY**

Google Earth satellite imagery was used to identify areas of the intertidal zone where
channels and pools were present. A field reconnaissance campaign, in January 2016,
identified an area containing an ephemeral ponds network, tidal channels and the intertidal
pool that is the focus of this study. The morphology of the pool was mapped using a Garmin
GPSmap 78 device; the data was subsequently imported and processed in Quantum GIS 2.18.
Microbial, including stromatolite and, thrombolites features, and other sedimentary and
biological characteristics of the pool were documented in detail. Monitoring and sampling
visits were regularly made between January 2016 and May 2017 with any changes to the
morphology of the pool or the associated features being documented in detail. Three
specimens of microbialite from the pool margin and one specimen of thrombolite from the
pool centre were recovered and subsequently stored in seawater in plastic containers.

A weather station (WS-GP1, Delta-T Devices), 3 km landwards from the study site (Fig. 1C)
recorded air temperature, air humidity, wind direction, wind speed, and precipitation at a
height of 1.2 m between April 2016 and July 2017. Battery failure resulted in data loss
between February and March 2017. Barometric pressure and temperature data were recorded
at 30 minute intervals between February 2016 and June 2017 using a Baro-Diver barometric
logger (Van Essen Instruments) sited approximately 2 km to the east of the weather station
(Fig. 1C). Data analysis and visualisation were conducted in the statistical computing
language R, in the programming language Python version 2.7.10 and in PAST version 3.15.
The physico-chemical characteristics of the water in the studied pool were measured using an Ultrameter II 6PFC device (Myron L Company) to record temperature, conductivity, total dissolved solids (TDS), oxidation-reduction potential (ORP), resistivity and pH. Very high dissolved solid load resulted in resistivity measurements that exceeded instrument parameters and, thus, were not used for interpretations. Salinity was measured using a Brix-type refractometer with automatic temperature compensation. The water level in the studied pond was monitored between February to October 2016 using a CTD-diver water level logger (Van Essen Instruments) installed 5 cm above the pool base. Water level was recorded as pressure in mbar, with a barometric conversion applied by subtracting barometric pressure from the water level pressure, under the assumption that 1 mbar = 1 cm of H₂O.

The microbialite structures were investigated at the micro-scale using a Quanta 200 (FEI) scanning electron microscope (SEM), located at The Petroleum Institute of Khalifa University of Science & Technology. One sample was selected from which representative sections were broken off, cleaned with compressed air, mounted on metal stubs, and coated with a palladium/gold mixture. The energy-dispersive X-ray spectroscopy mode (EDX) of the SEM was used to semi-quantitatively characterise element compositions of specific features. Samples were also prepared for analysis under the low-vacuum conditions of the environmental SEM mode. These samples rapidly dehydrated and became charged, it was therefore decided to focus on careful sample preparation and application of the conventional SEM mode.

Petrographic thin sections were prepared from three microbialite specimens, including the specimen that was investigated in the SEM. The sample were cut using a diamond rock saw and subsequently left to dry in the laboratory for 24 hours. Each sample was then impregnated with blue resin to enhance the visibility of pore space under the optical microscope.

DNA was extracted from samples of polygonal microbial mat, using a FastDNA SPIN kit for soil (Q-BIOgene, Cambridge, UK), according to the manufacturer’s instructions. DNA sequencing was performed on a MiSeq Personal Sequencer (Illumina) at the NUomics Facility (Northumbria University, Newcastle, UK). Gene amplicon libraries (16S rRNA) were prepared, including PCR amplification, purification of amplicons and quantification procedures, according to the procedures of Kozich et al., 2013. Aliquots of purified amplicons were sequenced to produce 2 x 250 bp paired-end reads with v2 chemistry. Raw sequencing reads were analysed using QIIME2 (Bolyen et al., 2019), with Amplicon Sequence Variants (ASVs) determined using Divisive Amplicon Denoising Algorithm (DADA2, Callahan et al., 2016). Taxonomic classification of representative ASV sequences was performed using the Naïve Bayesian classifier (Bokulich et al., 2018). The relative abundance of ASVs was calculated by taking the sum of the reads for individual ASV abundances and dividing by the total number of reads for all ASVs within a sample and multiplying by 100.

RESULTS

Environmental data
Air temperatures recorded by the weather station ranged between 8.4 °C and 48.9 °C; air temperatures recorded by the barometric logger ranged between 8.9 °C and 53.7 °C (Fig. 2A). The highest temperatures were recorded during July and August, while the lowest temperatures were recorded during February. Total precipitation over the measurement period amounted to 7.6 mm, mainly resulting from torrential rainfall events in January and February 2017 (Fig. 2A). March 2017 experienced heavy rainfall, but, as previously mentioned, this data was not recorded by the weather station due to a battery failure. Therefore, the rainfall data represent a minimum value for the measurement interval. The primary wind direction throughout the recording period was from the north-west, with subordinate winds from the south (Fig. 2C). Wind speed varied between 0.3 - 15.4 m/s with a mean of 3.7 m/s (Fig. 2C, D).

Water temperatures in the pool ranged between 11.7 °C and 46.8 °C during the measurement period (Fig. 2B). The lowest water temperatures were recorded in February while the highest water temperatures were recorded in August. Salinity ranged between 75.0 ‰ to 93.0 ‰, accompanied by pH values between 7.3 to 8.1. Of the remaining physico-chemical parameters, conductivity ranged between 100.0 - 118.7 mS/cm, TDS ranged between 75.0 - 91.26 parts per thousand (ppt), and ORP values ranged from 79 mV to 100 mV. The tidal regime in the studied pool was semi-diurnal micro-tidal, with a water depth ranging from a minimum of 19 cm when the pool was emergent up to a maximum of 109 cm during an unusually high tide (Fig. 2B).

**Pool morphology and hydrological regime**

The studied intertidal pool is located at the seaward margin of the polygonal microbial mat zone (Fig. 1C). The pool is broadly U-shaped and hydrologically open at its seaward and landward sides (Fig. 3A, B). The seaward opening of the pool corresponds to the landward termination of a tidal channel that extends into the lower intertidal and subtidal zones. The landward opening to the pool is defined by a narrow, well-lithified, sill that provides a connection to an elevated landward ephemeral pond system (Fig. 4). Ebb tide water continues to flow into the pool over this sill even whilst the next flood tide is recharging the pool via the tidal channel. The pool exhibits a perimeter of approximately 80 m and an area of approximately 250 m². The height difference between the cemented floor of the pool and its upper rim ranges between 20 to 30 cm. The floor of the pool is always submerged, this is in stark contrast to the elevated surrounding areas of microbial mat that experience daily cycles of exposure and inundation.

**Macro-scale sedimentary and biological features**

The general stratigraphy in the area of the pool (Fig. 3C) consists of Pleistocene to Early Holocene aeolian mixed siliciclastic-carbonate sands, overlain by a shallow-marine transgressive carbonate hardground of mid-Holocene age (Ge et al.; Lokier and Steuber, 2009; Paul and Lokier, 2017), followed by an overlying unconsolidated, to lightly-cemented, bioclastic rudstone. This sequence is terminated by laminated microbial mats with a
polygonal surface architecture. At the pool-floor, the hardground is locally exposed or is covered by an organic ooze that increases in thickness (up to 7 cm) landward (Figs 3 and 5). Accumulations of spongy, unlithified, mobile, gravel-sized grains are observed throughout the pool. These grains vary in colour and typically accumulate in small troughs and in the lee of thrombolite patches and bands. The grains vary in diameter between 5-10 mm and are irregularly shaped (Fig. 6A). The grains contain sub-mm scale inclusions of bioclastic grains including benthic foraminifera (Fig. 6B-D).

Clotted microbial (thrombolite) fabrics are distributed within the pool as isolated domes or discontinuous dm to m-long bands that are typically oriented parallel to the pool margin (Fig. 7A). The thrombolite domes typically measure between 5-20 cm in width, and exhibit relief above the hardground of between 10-25 cm. The structures are coloured brown to dark brown, and are characterised by a spongy consistency that reflects a dominantly cyanobacteria composition. Cyanobacteria were detected throughout polygonal mats in the area (Σ relative abundance; 8.5% ±2.1), with different morphologies including coccoid (Halothecia, Gleocapsa) and filamentous (Coleofasciculus, Lyngbya, Phormidium) forms. A mm-thick cover of bioclastic grains is locally present at the upper surface of the thrombolites (Fig. 7B). Marine brown and green algae grow at the outer margins of individual thrombolites, defining the minimum water depth within the pool. Internally, the domal structures comprise two distinct intervals (Fig. 3C). The upper interval is a thrombolite, exhibiting crude laminations corresponding to different colours of microbial communities interlayered with bioclasts (Fig. 8A). The lower layer is a ~10 cm thick laminated stromatolite that is attached to the underlying hardground (Fig. 8B). The margins of the pool exhibit the same vertical facies distribution as the thrombolite domes and bands but are locally undercut by up to 10 cm (Figs 3 and 9).

Moving away from the pool, into the adjacent microbial mats, the lithified cyanobacterial communities of the pool margin transition over decimetres into an unlithified finely-laminated and polygonal microbial mat dominated by filamentous cyanobacteria (Fig. 10). Elongate scours in excess of 1 m long are distributed throughout the polygonal microbial mat zone (Fig. 11). The scours are oriented perpendicular to the shoreline, are shallowest at their seaward end and gradually deepen landward. The scours host a variety of microbial communities that closely resemble the organic ooze observed in the pool.

Micro-facies and cement fabrics

The pool margin microbialites contain abundant lithoclasts, bioclasts, peloids and coated grains (Fig. 12A) surrounded by pervasive microbial extracellular polymeric substance (EPS) (Figs 13A, B). The skeletal assemblage observed within the microbialites is dominated by benthic foraminifera (primarily peneropolids) and ostracods with subordinate bivalves. Skeletal grains are typically whole with little evidence of abrasion or breakage (Fig. 12A). SEM observations reveal the presence of cyanobacteria tubes and filaments within the EPS (Fig. 13 B, E). Primary microbial laminae are difficult to discern in thin section, occurring as discontinuous degraded organic seams that could be easily confused with dissolution seams (Fig. 12E, F).

Grains are coated by an isopachous fringing acicular (Sandberg et al., 1985) aragonite cement with fibrous needles oriented perpendicular to grain surfaces (Figs 12B-D). The aragonite
needles exhibit pointed crystal terminations, have a length of 16-185 μm with a width of 3-5 μm (axial ratio typically >22). Rare dolomite cements occur as isolated rhombs between 3-6 μm and more-massive clusters of slightly coarser (up to 8 μm) equant euhedral crystals (Fig. 13C, D). These dolomite crystals are enclosed in microbial EPS and acicular aragonite. Semi-quantitative EDX elemental analysis of the dolomite crystals yield an Mg content of between approximately 20-40% compared to significantly lower values (~14%) in the surrounding matrix (Fig. 13 C, D).

Primary interparticle porosity has been largely occluded by the pervasive precipitation of the aragonite cement phase that has cemented grains and significantly restricted pore throats (Fig. 12C, D). Secondary, yet syndepositional, mouldic porosity after bioclasts (Fig. 12B), has also been partially to completely occluded by the precipitation of the aragonite cement phase.

**INTERPRETATION AND DISCUSSION**

On the basis of field and laboratory observations, it is here proposed that the formation of the intertidal pools and the subsequent genesis of the hosted stromatolite and thrombolite microbial fabrics are genetically related.

**Formation of intertidal pools and associated microbialites**

As has been previously documented, the development of significant modern-day microbial mat communities requires stressed environmental conditions that prevent competition from macroalgae and other eukaryotic organisms (Bouton et al., 2016; Bowlin et al., 2012; Suosaari et al., 2016b). In the case of the Abu Dhabi sabkha, these conditions are met in the uppermost intertidal zone where diurnal flooding is sufficient to maintain microbial growth, yet exposure inhibits competition. Under such conditions, laminated, unlithified, microbial communities are able to thrive.

Pool formation is initiated where a high energy event, such as a storm surge or high tide, removes the protective binding microbial mat and exposes the underlying unconsolidated sabkha sediment (Fig. 11A). Under the prevailing relatively low-energy conditions of the Abu Dhabi sabkha, such events are relatively rare and of limited duration, thus normally allowing recolonisation of the sediment surface by a new generation of microbial mat and the effective ‘healing’ of the protective binding layer. However, prolonged episodes of high energy, or multiple high energy events over a short time interval, will erode the unprotected sediment to produce an erosional scour that may extend to the depth of the underlying lithified hardground (Figs 11B, 14B). Initially, the unprotected vertical walls of the pond are susceptible to continued erosion, thereby facilitating lateral growth of the pool.

With a return to quiescent conditions, the newly formed pool walls (Fig. 11B) provide a submerged substrate for the development of cyanobacteria dominated microbial communities that overgrow the exposed truncated surface of the laminated microbial mats (Figs 9, 10) and promote aragonite precipitation (Figs 12, 13). Lithification stabilises both the underlying laminated microbial mat substrate, producing stromatolite fabrics, and the clotted textures to
form a thrombolitic fabric (Fig. 14C). Increasing lithification stabilises the pool walls and inhibits erosion during subsequent high energy events.

Microtidal flooding cycles ensure regular marine inundation of both the pool and surrounding microbial mats by seawater that is oversaturated with respect to Ca$^{2+}$ and Mg$^{2+}$ (Paul and Lokier, 2017; Shinn, 1969; Wood and Sanford, 2002). Ebb tide surface flow is concentrated into tidal channels that rapidly drain much of the upper intertidal zone. High evaporation rates and shallow sub-surface bacterial sulphate reduction (Lokier and Steuber, 2008) further increase the concentrations of major ions in ponded surface and shallow subsurface waters. Localised dissolution of aragonite allochems (Fig. 12B) occurs where the pH of the pore-water is reduced by the oxidation of hydrogen sulphide (Jordan et al., 2015). These highly saline, Ca$^{2+}$ and Mg$^{2+}$ oversaturated pore-waters flow seawards in the shallow subsurface until they intercept the pool margin and discharge into the pool, further promoting the precipitation of aragonite cements (Fig. 14C).

Over time, the pool walls become increasingly lithified through the continued precipitation of aragonite cements. During high tides, waves breaking at the pool ‘shore’ will erode the un lithified microbial mats immediately behind the wave-resistant cemented pool margin (Figs 3, 14C). Simultaneously, currents will erode the un lithified basal rudstone to undercut the pool wall (Fig. 9) and further destabilise the edge of the pool. This process will eventually result in the detachment of a section of the cemented microbial thrombolites and stromatolites which will founder onto the exposed hardground at the pool floor to create detached sub-aqueous margin-parallel thrombolite bands (Figs 3, 7A, 14D). The submerged thrombolite bands will continue to grow and precipitate aragonite cements within their clotted fabric (Fig. 13). High energy events, such as storm surges, will detach and remove sections of the thrombolite bands. Erosion will decrease over time as the precipitation of early marine aragonite (Lokier and Steuber, 2008; Paul and Lokier, 2017) cements the thrombolite to the underlying hardground and stabilises the structure (Fig. 14E).

The freshly-exposed pool margin is now subject to renewed microbial colonisation and associated aragonite precipitation and the process is repeated with the genesis of a new generation of margin-parallel thrombolite band (Fig. 14F). With repeated cycles, this process will result in significant lateral expansion of the pool, possibly resulting in coalescence with any adjacent pools. Over time, the earlier thrombolites will become increasingly isolated (Fig. 3) and may be subjected to undercutting to produce the distinctive dome-shaped morphology with clotted thrombolitic fabrics sitting atop laminated stromatolites (Figs 7B, 8, 14F). These thrombolite domes superficially resemble the ‘isolated patch’ macrofabrics described from the Cayo Coco Lagoon, Cuba (Bouton et al., 2016). However, the Cayo Coco structures appear to be entirely thrombolitic in nature, lacking either a laminated stromatolite component or a relationship with an underlying hardground.

The accumulations of un lithified, spongy, microbial gravel-grade grains at the pool floor (Figs 6, 7A) are comprised of cyanobacteria and EPS forming a clotted microbial fabric akin to that of the thrombolites. These soft grains are inferred to have been sourced from the thrombolite domes or pool margin thrombolite bands either through erosion or via exfoliation due to high rates of microbial production. Once disarticulated, the low-density grains are particularly susceptible to reworking and concentration in the lee of thrombolites and in bathymetric lows. Where a significant amount of carbonate material is bound within these grains, it is possible that they may be preserved in, and recognised from, the geologic record as large aggregate grains with a clotted matrix (Gerdes et al., 1994). In more-open areas of
the pond, agitation of the microbial grains, by wave and current action, will result in abrasion and disarticulation to generate the microbial ooze that covers much of the pool floor.

The rare small, EPS-hosted, dolomite crystals observed in the pool-margin stromatolites are consistent with a microbially-mediated origin as proposed previously (Bontognali et al., 2010). Under this model, the fluctuating hypersaline conditions of the sabkha environment result in the EPS generating carboxyl functional groups that overcome the kinetic barrier to the low temperature incorporation of Mg into carbonate minerals (Bontognali et al., 2014; Petrash et al., 2017). These clusters of dolomite crystals bare a remarkably similarity, in terms of size, distribution and association with biofilms, to those previously documented from the Lower Messinian of Sicily (Oliveri et al., 2010).

The well-lithified nature of the sill separating the pool from the landward ephemeral ponds is inferred to result from a combination of microbially-mediated aragonite precipitation enhanced by degassing-related precipitation as the solubility product of the outflowing water is lowered as it cascades over the sill margin. This process is similar to the precipitation of carbonates at travertine terraces and tufas (Fig. 15; Gandin and Capezzuoli, 2014; Özkul et al., 2014).

Microbialite formation in a sea level context

The formation of the pools and their associated microbial communities is inferred to preferentially occur under prevailing conditions of relative sea level stillstand or, more likely, under the increased hydrodynamic conditions associated with relative sea level rise. The present-day rate of eustatic sea level rise (3.2 mm yr$^{-1}$) (Church et al., 2013) has been previously documented as resulting in significant shoreline retreat (up to 29 m yr$^{-1}$) and associated hydrodynamic energy increase at the Abu Dhabi coastline (Lokier et al., 2018). Under such a transgressive (flooding) scenario it is expected that we shall observe an increase in the frequency and amount of erosion of the unlithified upper intertidal microbial mats and, consequently, the increased initiation and development of pools with their associated stromatolite and thrombolite fabrics. However, under conditions where the rate of relative sea level rise is ‘too rapid’, the associated increase in energy at the shoreline may result in dynamic coastline retreat at a rate that exceeds the ability of the microbial systems to re-establish and ‘track’ retrogradation.

From an environmental perspective, as the rate of present-day eustatic sea level rise continues to increase (Church et al., 2013), coastal microbial communities will be placed increasingly under threat and should thus be added to the growing list of coastal and shallow-marine ecosystems (Lovelock et al., 2015; Perry et al., 2018; Thorner et al., 2014) that are at risk due to anthropogenic-driven global warming and associated sea level rise.

Microbialite preservation and identification in the fossil record

Under a rising sea level scenario, as observed in the study area today, the preservation potential of the described structures is primarily dependent on the degree of early-lithification
and the subsequent rate of burial. As sea level rises, increased hydrodynamic energy easily erodes the unconsolidated microbial mats and thrombolites, transporting much of the material into the marine system where it is consumed. The likelihood of preservation of unconsolidated microbial features is further compromised as increased, and more regular, inundation reduces environmental stress in the intertidal and shallow subtidal zone. Such increased flooding opens this niche-environment to colonisation by macro-algae and other eukaryotes that will outcompete and displace the microbial communities. On entering the burial realm, ongoing degradation of the organic component of the unconsolidated microbial mats and thrombolites will further reduce the potential for preservation and identification within the stratigraphic record (Court et al., 2017; Kenig et al., 1990).

Conversely, cemented stromatolites and thrombolites have a significantly higher prospect of preserving primary depositional fabrics during inundation and subsequent burial and will, thus, have a higher preservation and recognition potential into the stratigraphic record. Where these early-lithified structures remain exposed at the sea floor, or are covered by only a thin veneer of sediment, they will be subject to early-marine cementation, thereby further enhancing the potential of preservation. If exposed at the sea floor, the lithified structures will act as a stable substrate for colonisation by sessile benthic communities, again enhancing the potential of preservation but providing a challenge for identification in the fossil record.

Where ponds and their associated structures have formed during a period of high sea level or falling stage stillstand, the chances of preservation into the fossil record are significantly reduced. During post-stillstand base level fall, unconsolidated microbial communities are subject to biodegradation and physical erosion. On entering the meteoric realm, lithified microbial structures will be subject to increased dissolution of aragonite cements and allochems. As erosion removes the surrounding unconsolidated sediments, the increasingly exposed cemented pond margins will lose their support, becoming unstable and subject to increased likelihood of erosion. This process may continue until all of the sediments are eroded to expose the underlying, strongly lithified, transgressive hardground, thereby potentially removing all evidence of the preceding transgressive cycle. Under these circumstances, transgressive sequences will only be preserved into the stratigraphic record where high frequency (4th-5th Order) transgressive cycles occur during a theme of overall, lower order, sea level rise. This scenario will result in repeated transgressive cycles punctuated by stillstands to produce parasequences as observed in the sabkha stratigraphy of the Jurassic Arab Formation (Grötsch et al., 2003).

Under the above-proposed scenarios, preservation of these structures into the geological record is only likely under a rising sea level scenario and where the microbial structures have been lithified. As such, the recognition of these systems from the stratigraphic record offers a potential diagnostic tool to elucidate the onset and, possibly, rate of marine transgression.

The differential preservation potential of lithified and unconsolidated components together with a limited thickness and laterally isolated morphology of these systems offers many challenges to their observation and interpretation in the geological record. Identification of the system requires recognition of lateral and vertical facies relationships and gross morphologies that is best suited to observation in laterally extensive outcrops. Conclusive identification of these systems from core would be significantly more challenging. A vertical stacking pattern of transgressive hardgrounds succeeded by stromatolites and overlying thrombolites, possibly with encrusting marine benthos, would hint at the presence of these systems. However, these vertical relationships could also be observed under other formational mechanisms, such as...
CONCLUSIONS

Intertidal pools and associated microbial fabrics were observed and documented from the coastal sabkha of Abu Dhabi. We propose a new, erosion-based, model for the formation of these systems under a scenario of rising relative sea level. Increasing energy at the shoreline results in erosion of protective microbial mats to expose underlying sediments to erosion and initiate pool formation. These newly-formed submerged environments provided an ideal habitat for the pervasive growth of cyanobacteria that promoted the cementation of stromatolite and thrombolite textures thereby significantly increasing the potential for the preservation of these microbial structures. Successive erosional events enlarged the pools and created isolated microbialites with a range of textures.

Although the recognition of these features in the stratigraphic record may be challenging, their association with the onset of transgression makes them a potentially powerful diagnostic tool for constraining the initiation of relative sea level rise.

ACKNOWLEDGEMENTS AND DATA AVAILABILITY

All sample site locations are clearly defined in the manuscript text and figures. If additional sample or other data are required then these can be accessed by contacting the corresponding author. This study was funded by the Petroleum Institute Research Centre (PIRC/ADRIC) through project LTR15003 titled ‘Understanding ancient petroleum carbonate systems; carbonate precipitation in Abu Dhabi microbial mats as a modern analogue’. The authors declare no conflict of interest related to this study. Our sincere appreciation goes to Sion Kennaway for general laboratory and field support, to Warren Marilag for preparing the petrographic thin sections, and to Prasanth Thiagarajan for his assistance with the SEM and EDS analyses. Xin Bixiao and Yuan Peng are thanked for their help during the many long field-days in the sabkha of Abu Dhabi. We thank Tomaso Bontognali, Edoardo Perri and two anonymous reviewers for their very constructive comments. We also acknowledge Associate Editor Alexander Brasier and Chief Editor Peir Pufahl for their helpful comments and guidance through the editorial process.

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Table and figure captions


Figure 1. A) Location of the study area in relation to the Gulf. B) Location of study area in the context of the Abu Dhabi coastline. C) Satellite image of the coastal sabkha of Abu Dhabi, showing facies belts and anthropogenic changes. The study site is indicated by a white star. Imagery source: ESA Sentinel 2A Multi-spectral Imager (MSI), date: 18 March 2017.

Figure 2. A) Mean daily air temperatures in the coastal sabkha measured between February 2016 and June 2017 using a barometric logger (black line) and the GP1 weather station (red line). Daily maximum and daily minimum temperatures are shown in the background. Precipitation in January and February 2017 is indicated by the blue columns. B) Record of water temperatures and water depth in the studied intertidal pool, showing the daily tidal regime as well as long term annual trends in water depth. Note that the pool was never empty. C) Wind rose showing the dominant wind directions from the WNW - NW and S, and wind speeds. D) Frequency distribution plots of wind speeds for two intervals in January – February 2017 and in July – August 2016. Each vertical bar represents a 0.5 m/s wind speed interval.

Figure 3. A) Overview photograph of the studied pool. Note that the scale is not uniform from bottom to top due to the mode of image acquisition; person in background is 175 cm high. B) Interpretative overview of the features observed in the pool. Seaward direction is to the left. Note the detached bands at the pool margins, and the thrombolites towards the centre of the pool. C) Schematic cross-section through the pool showing the general stratigraphy, lithologies and features described in this study.

Figure 4. A) Close-up view of the lithified sill – see Fig. 3 for image location in the studied pool. The ephemeral pond system is located at the top of the image, while the pool is towards the bottom (not visible). The location of the water level logger is indicated by an arrow. Note the colonisation of the logger by cyanobacteria and algae. B) Schematic cross-section showing with the relationship between the ephemeral pond system, lithified sill and the pool.

Figure 5. A) Photograph of the organic ooze covering parts of the pool floor. Grazing trails of gastropods are visible with the respective gastropods at the end-points (arrows). B) Close-up view of the ooze and a gastropod (large arrow) and its grazing trail (small arrows).

Figure 6. A) Microbial ‘gravel’ collected from the pool floor. Different colours correspond to different organic pigments. B-D) Close-up views of individual microbial grains showing the EPS and white bioclastic grains embedded (arrow).

Figure 7. A) Close-up view of different microbial structures including thrombolites and stromatolites. Note the detached rim and the domal and banded thrombolites, surrounded by organic ooze and gravel-sized grains of microbial origin. Note also the aluminium can overgrown by microbial communities and partially covered by organic ooze. B) Close-up
view of an individual thrombolite showing its components: brown thrombolite, bioclasts and algae growth. Upper growth boundary of these algae indicates the minimum water depth in the pool, which corresponds to a depth of ~20 cm as also calculated from the water level logger data. See Fig. 3 for image location in the studied pool.

Figure 8. A) Cross-sectional view of an individual domal thrombolite, showing a clotted texture with internal crude radial laminations, based on differences in colour. Bioclasts enclosed by microbial EPS indicate outward growth. Coarse laminae are indicated by dashed lines. B) Hand specimen of a stromatolite from the pool margin. Microbial laminations preserved through lithification are clearly visible (between white arrows). Purple and green colours correspond to pigments of living cyanobacterial communities and are unrelated to the primary processes that lead to the development of the stromatolite.

Figure 9. Oblique subaqueous view of the partially undercut margin at the north-west side of the pool - see Fig. 3 for image location in the studied pool. Note the laminated stromatolites at the centre (white arrow).

Figure 10. Overview photograph showing the transition from the pool, via a rim overgrown with algae, into the surrounding polygonal microbial mat zone. Note that the polygonal microbial mat in the transition zone appears to be degraded. See Fig. 3 for image location in relation to the studied pool.

Figure 11. A) An area of recently eroded microbial mat exposing the underlying unconsolidated sediment. Note the incoming tide in the bottom left of the image (person is 185 cm tall). B) An erosive scour in the microbial mat zone. C) Schematic cross-section through the scour.

Figure 12. Thin section photomicrographs of stromatolites and thrombolites from the pool margin. A) Lithoclasts embedded in a stromatolite are shown surrounded by microbial organic-rich rims (arrow). B) Mouldic porosity in a thrombolitic area filled by acicular aragonite cements (arrows). C, D) Acicular aragonite cements filling thrombolite intragranular pore space. E, F) Discontinuous organic seams define the microbial laminations observed in hand specimen (see Fig. 8B) (arrows).

Figure 13. Scanning electron microscope micrographs of stromatolites and thrombolites. A, B) Extra-polymeric bacterial substance covering some areas of stromatolite specimen from the pool margin. Note bacterial filaments (white arrows). C) Crystal showing a very high Mg content indicative of proto-dolomite (arrow). The crystal is surrounded by aragonite cement and microbial EPS. D) Group of proto-dolomite crystals with very high Mg contents, surrounded by bacterial filaments, EPS and aragonite. E) Mineral precipitates in the microbialites showing cyanobacterial tubes (arrows). F) Acicular aragonite covered by a layer of EPS (arrows).

Figure 15. Schematic overview of the process that leads to permanent water coverage of the pool. During highest tides the pool is inundated by ~ 1m of water. During low tide the ephemeral ponds system slowly empties its excess water into the pool. Since this process takes much longer than one tidal cycle, the pool remains filled with water. At the margin, cascading water results in degassing, promoting the precipitation of aragonite cements. (MHW = mean high water)
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Seaward

Thrombolite bands

Thrombolite domes

Person is 175 cm in height

Ephemeral ponds

Fig. 9

Fig. 10

Fig. 7

Fig. 4

Aeolian sand (Pleistocene to Early Holocene)

Bioclastic rudstone

Thrombolites, crudely laminated

Organic ooze

Microbial ‘gravel’

Minimum water depth

Lithified sill

Coccoid cyanobacterial mat

Undercutting

Filamentous cyanobacterial mat (polygonal surface)

Bioclastic rudstone (unlithified)

20-30 cm

Marine hardground (Middle to Late Holocene)

Aeolian sand (Pleistocene to Early Holocene)
Ephemeral Pond

A

Ephemeral Pond

Lithified sill

Microbial communities

10 cm

A'

Pool

B

Landward

Seaward

Ephemeral Pond

Lithified sill

Low water

MHW

Pool

Low water
Organic ooze

Stromatolite & thrombolite band

Domal thrombolite

Microbial grains

Aluminium can

Organic ooze

Lithified margin

Microbial communities

Biclasts

Algae
A  Detrital sediment fill between polygons

B  Newly formed pool

C  Cementation and erosion

D  Margin-parallel band formation

E  Stabilisation

F  Isolated thrombolites