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Supplementary Materials for

SARS-CoV-2 within-host diversity and transmission

Katrina A. Lythgoe^{1,2*}†, Matthew Hall^{1*}†, Luca Ferretti¹, Mariateresa de Cesare^{1,3}, George MacIntyre-Cockett^{1,3}, Amy Trebes³, Monique Andersson^{4,5}, Newton Otecko¹, Emma L. Wise^{6,7}, Nathan Moore⁶, Jessica Lynch⁶, Stephen Kidd⁶, Nicholas Cortes^{6,8}, Matilde Mori⁹, Rebecca Williams⁶, Gabrielle Vernet⁶, Anita Justice⁴, Angie Green³, Samuel M. Nicholls¹⁰, M. Azim Ansari¹¹, Lucie Abeler-Dörner¹, Catrin E. Moore¹, Timothy E. A. Peto^{4,12}, David W. Eyre^{4,13}, Robert Shaw⁴, Peter Simmonds¹¹, David Buck³, John A. Todd³ on behalf of the Oxford Virus Sequencing Analysis Group (OVSG)‡, Thomas R. Connor^{14,15}, Shirin Ashraf¹⁶, Ana da Silva Filipe¹⁶, James Shepherd¹⁶, Emma C. Thomson¹⁶, The COVID-19 Genomics UK (COG-UK) Consortium§, David Bonsall^{1,3,4}, Christophe Fraser^{1,3,17}, Tanya Golubchik^{1,2†}

*These authors contributed equally to this work.

†Corresponding author. Email: Tanya.Golubchik@bdi.ox.ac.uk (T.G.);

Katrina.Lythgoe@bdi.ox.ac.uk (K.A.L.); Matthew.Hall@bdi.ox.ac.uk (M.H.)

‡The full list of the OVSG members is provided in the supplementary materials.

§The full list of names and affiliations of COG-UK members is provided in the supplementary materials.

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This PDF file includes:

- Figs. S1 to S8
- Tables S1 to S5
- List of OVSG members
- List of COG-UK consortium names and affiliations

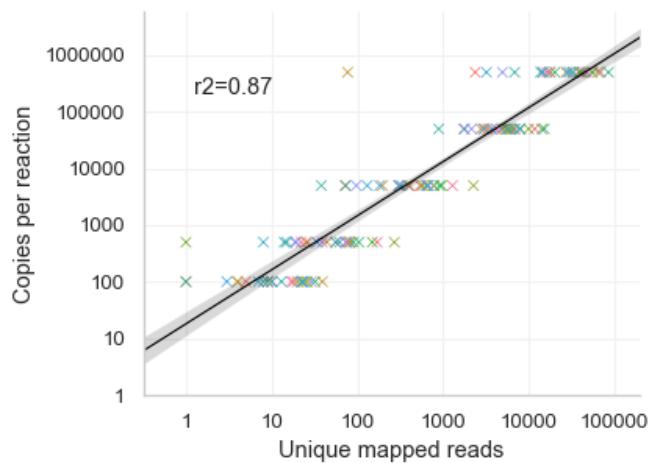
Other Supplementary Material for this manuscript includes the following:

(available at science.scienmag.org/cgi/content/full/science.abg0821/DC1)

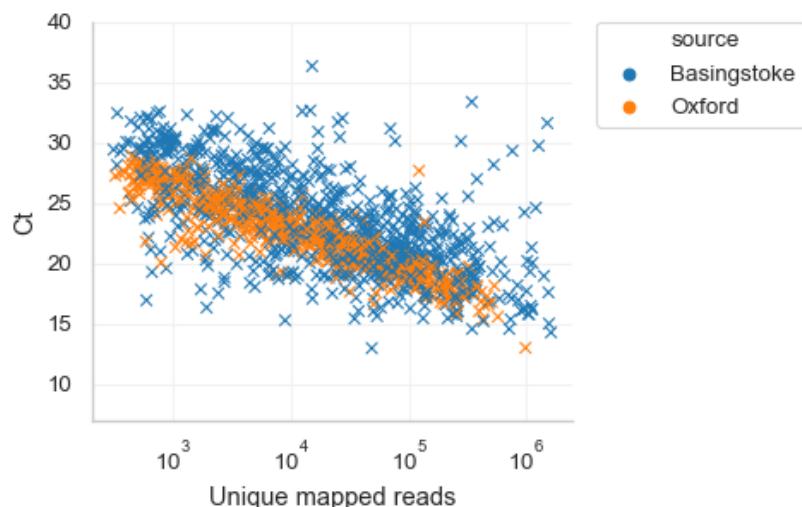
[MDAR Reproducibility Checklist \(PDF\)](#)

Figures S1-S8

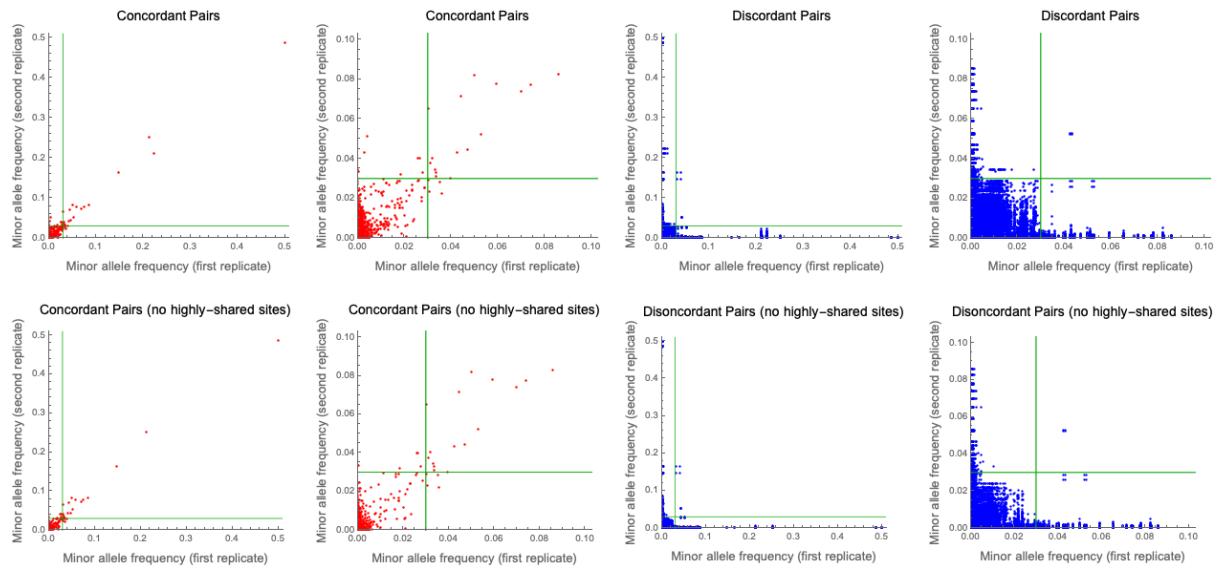
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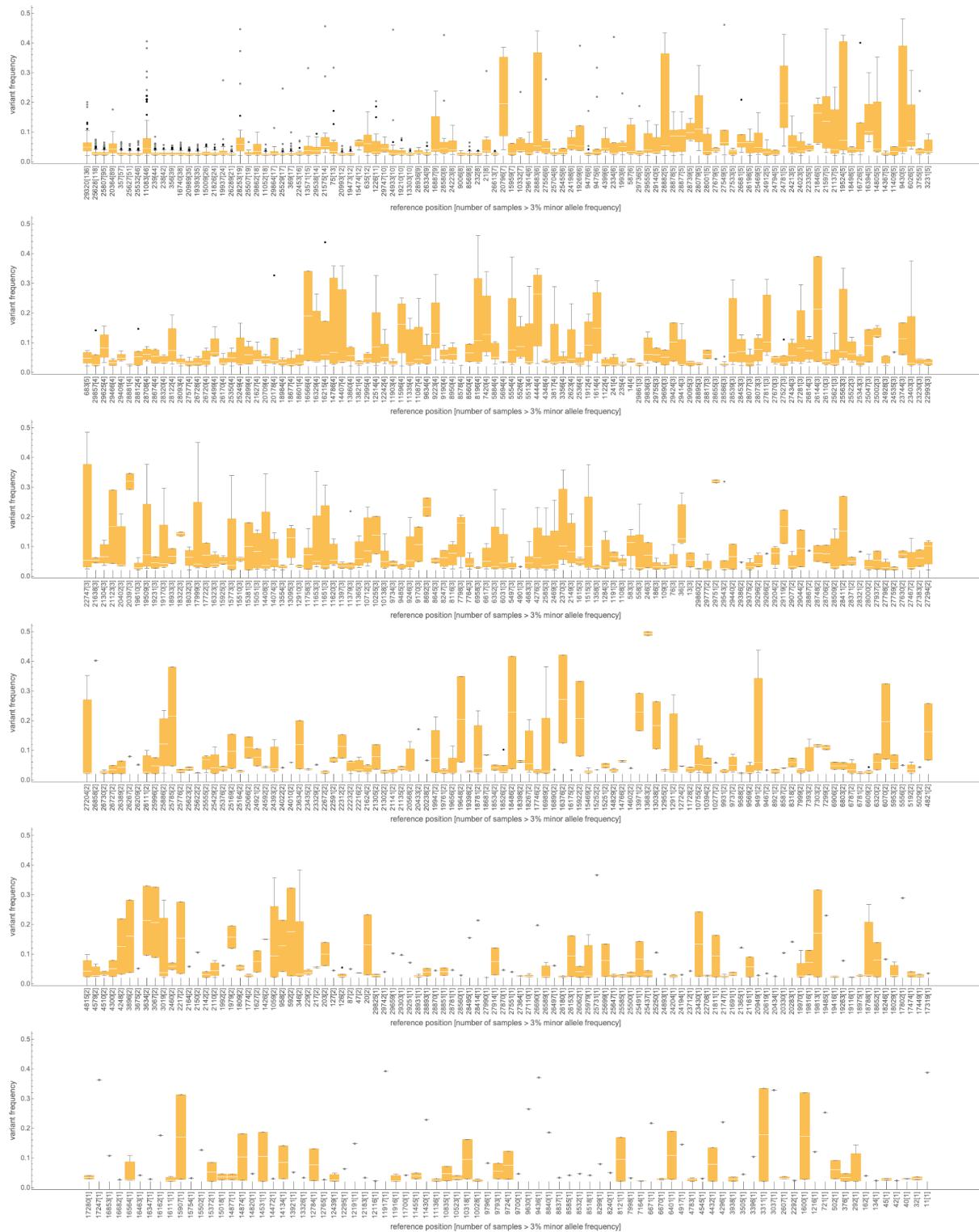
b



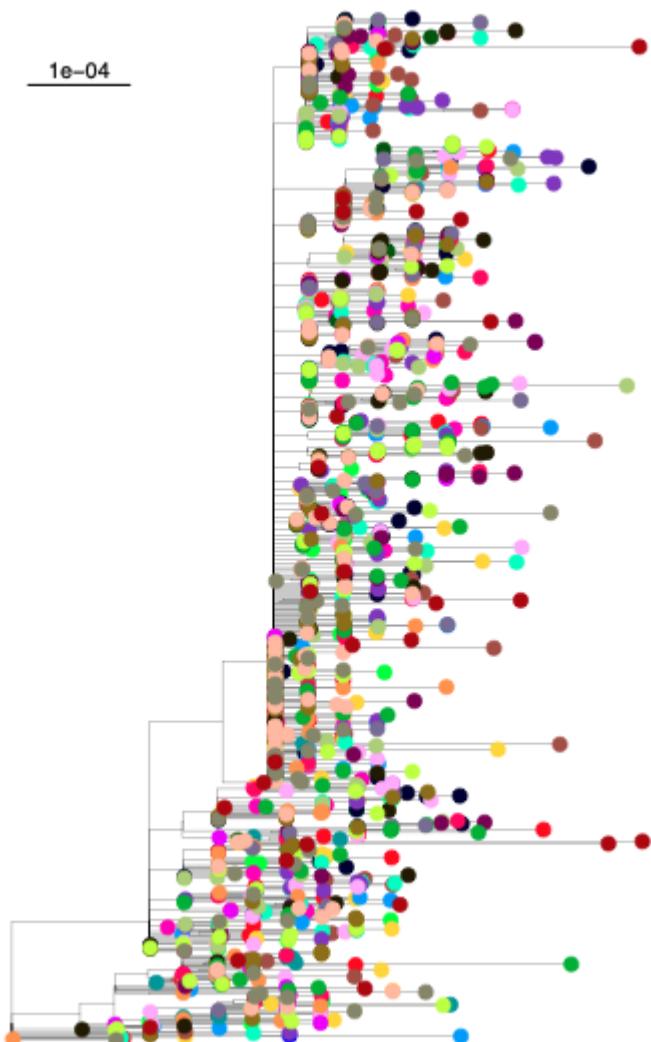
Supplementary Figure 1. (a) Correlation between number of SARS-CoV-2 unique reads and RNA copies/ml for within-batch standard curves for dilution series of positive control RNA. Colour indicates batch. Synthetic SARS-CoV-2 RNA (generated by in vitro transcription by Twist Bioscience) was serially diluted into Universal Human Reference RNA (UHRR) to a final concentration of SARS-CoV-2 RNA of 500,000, 50,000, 5,000, 500, 100 and 0 copies/reaction. Controls were processed and sequenced alongside each batch of samples (batches 3-27). Batches 1 and 2 were processed prior to controls being available and did not have a standard curve. (b) Correlation between nearest available cycle threshold (Ct) value for sequenced clinical samples, as reported by the collecting laboratory, and the number of unique mapped reads. Due to variation in qPCR methodology, Ct values varied substantially between laboratories and over time ($r^2 = 0.43$ Basingstoke, $r^2=0.81$ Oxford, overall $r^2=0.50$) Slightly lower Ct values were observed for Basingstoke samples for the same viral load (estimated from the standard curve), consistent with laboratory variation in Ct estimates.



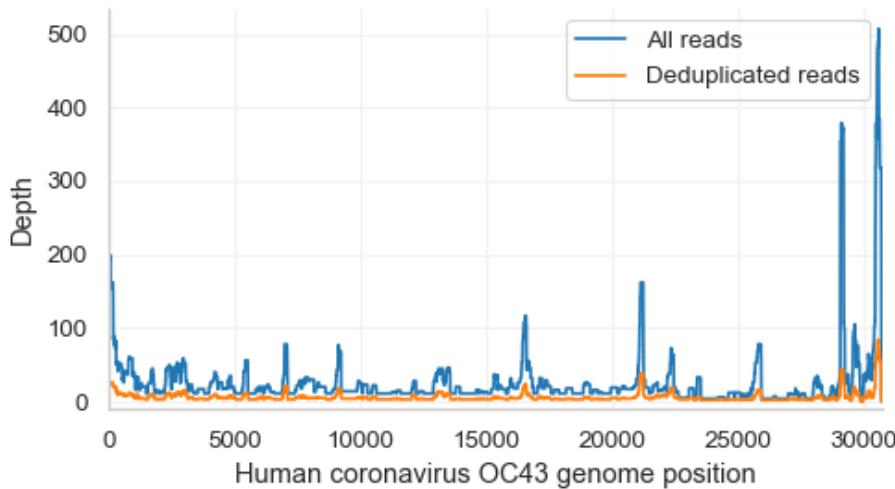
Supplementary Figure 2. Comparison of minor allele frequencies among replicate samples. Data is only included for the 27 replicate pairs where both replicates had more than 50,000 unique mapped reads, and for all sites where minor allele frequency (MAF) $\geq 2\%$ and depth ≥ 100 in at least one of the 54 replicates. For MAFs $> 3\%$, and excluding highly-shared sites, MAFs are highly reproducible. Concordant pairs: The points represent the MAFs in each of the replicate pairs, for all 27 replicate pairs for all identified sites. If MAFs are reproducible, we expect a positive correlation. Discordant pairs: The points represent the MAFs for all pairwise permutations of replicates for all identified sites, excluding concordant replicates. Unless variants are present in multiple samples, the expectation is for points to be positioned along the axes. Top row includes all sites, whereas the bottom row exclude highly-shared sites (those observed at MAF $\geq 3\%$ in 20 or more samples across the entire dataset). The blue points in the upper-right quadrants represent site 28580, which is present in phylogenetically linked individuals, with two of these included in the 27 replicate pairs. The green line shows MAF 3%.



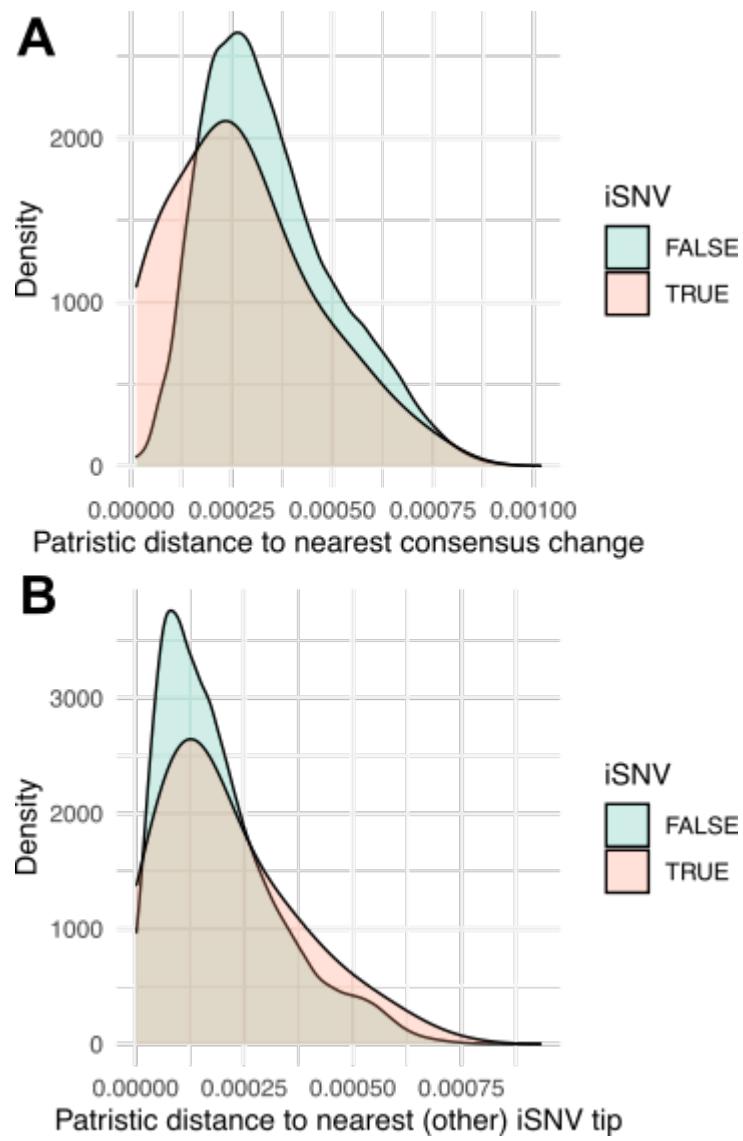
Supplementary Figure 3. Box-whisker plots showing distribution of variant frequencies. All sites variable at minor allele frequency (MAF) > 3% in at least one high-depth sample (>50,000 mapped reads) are included. All variants with MAF > 2% are plotted. The x-axis gives the nucleotide position, with the number of samples MAF > 3% in brackets. All sites are included except the poly A tail, and sites diverse in synthetic controls. The first 18 ‘highly-shared’ sites are either ubiquitous, or only ever seen at low frequencies, suggesting these are non-genomic variants.



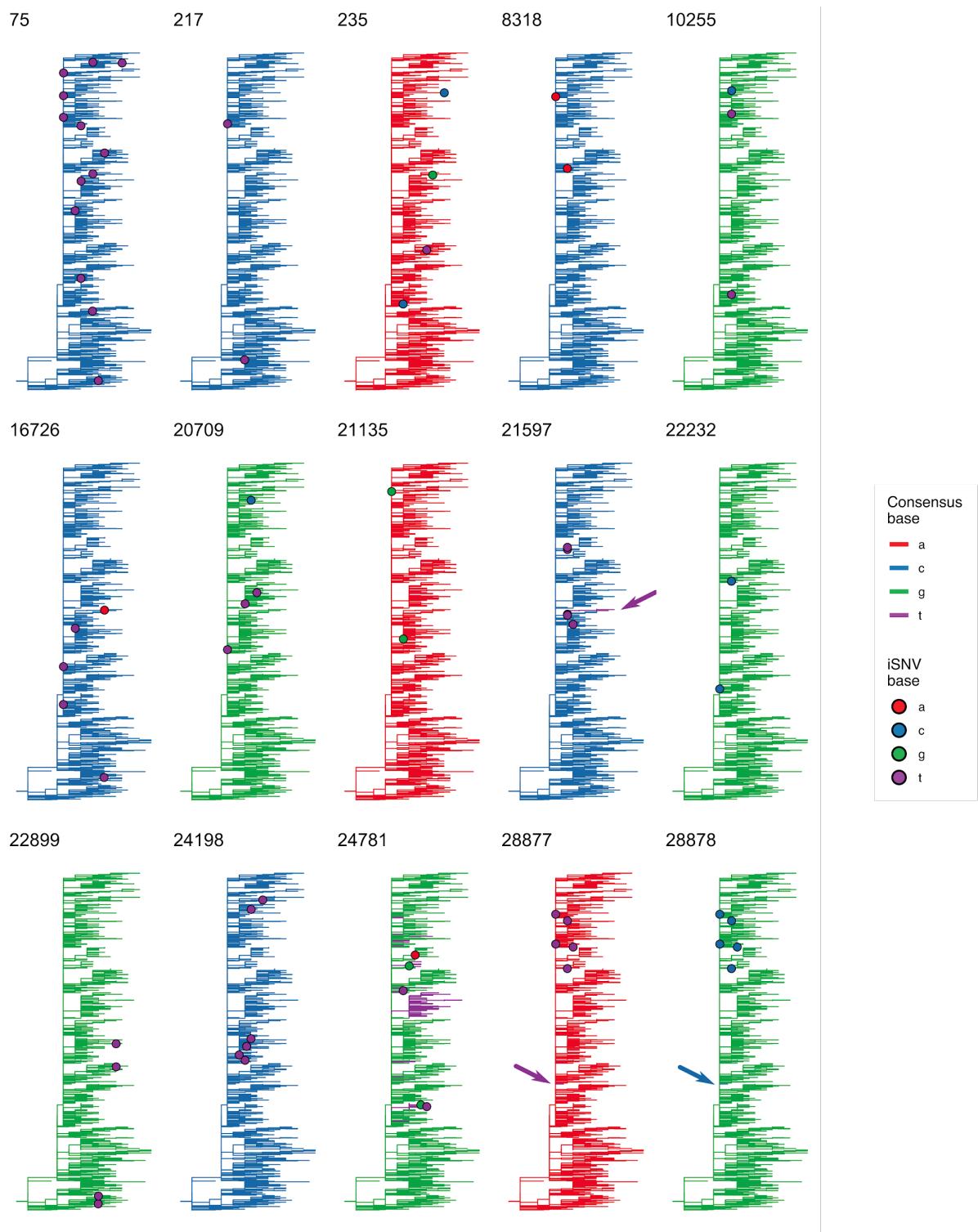
Supplemental Figure 4 - Consensus phylogeny of all 1390 Oxford and Basingstoke samples. Tips are coloured by sequencing batch.



Supplementary Figure 5. Genome coverage for co-infecting human coronavirus OC43 in a SARS-CoV-2-positive sample, OXON-AEC3D. A single co-infection with a non-SARS-CoV-2 circulating coronavirus was detected among a subset of 111 samples analysed with both SARS-CoV-2-specific probes and the Castanet metagenomic respiratory probe panel. Shown in blue are positions of the 2953 proper read pairs mapping to the Castanet reference for OC43, with unique (deduplicated) read depth in orange.

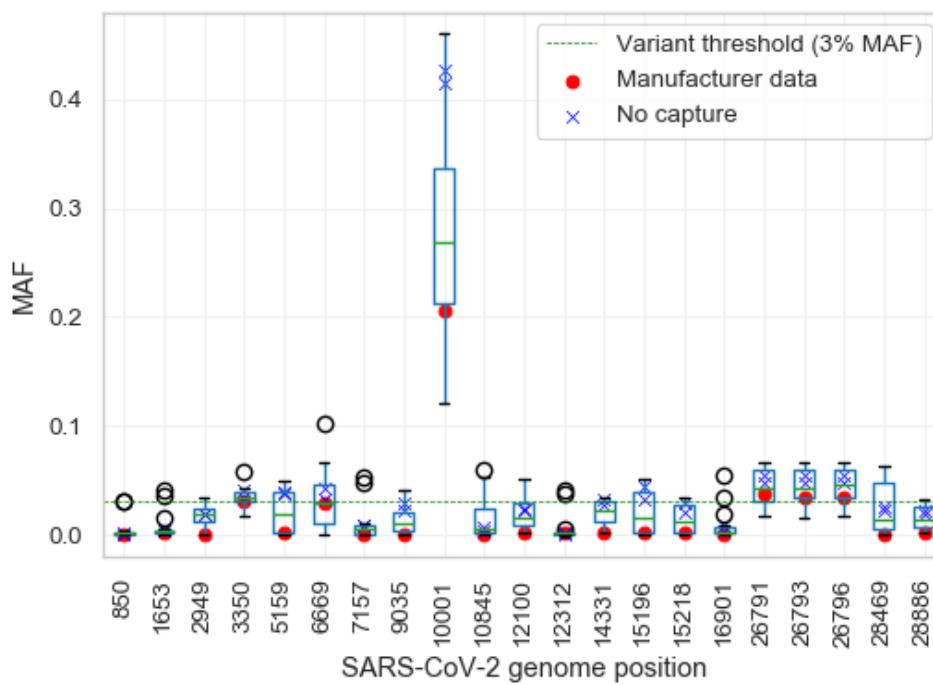


Supplementary Figure 6 - A Across all iSNVs that reach consensus, kernel density plot of the patristic distances from iSNV tips (orange) and other tips (green) to the nearest consensus branch change of the nucleotides involved. **B** Across all iSNVs that do not reach consensus and occur at least twice, kernel density plot of the patristic distances from iSNV tips and other tips to the nearest iSNV tip (other than the tip itself).



Supplementary Figure 7 - The consensus phylogeny coloured by SNP and iSNV for sites mentioned in the main text. Where consensus changes are hard to see they are indicated with an appropriately coloured arrow. Sites 21597, 24781, 28877 and 28878 are the remaining positions where a statistically significant association of iSNV tips with branches with a consensus base change was identified (along with 20796 and 28580). For some of these, coloured arrows indicate the presence of branches with consensus SNPs where this is difficult to see. Sites 22899 and 24198 are Spike variants shown to exhibit reduced sensitivity to convalescent sera (44). The remaining 9 subfigures are for iSNVs

which never reach consensus but show a $p<0.025$ for phylogenetic association of iSNV tips using the association index or the mean patristic distance between iSNV tips. While we lack the power to identify these once the Benjamini-Hochberg adjustment is applied, the patterns remain suggestive of transmission of iSNVs by eye.



Supplementary Figure 8. Within-sample diversity assessed in control RNA (Twist Bioscience). Within-sample diversity was assessed in RNA controls sequenced with each sequencing batch (0.5 mln copies per reaction). At all sites where at least 2 replicates had a minor variant with minimum 3% MAF (boxplot), diversity was compared against a set of NGS reads obtained from Twist Bioscience for the ancestral stock of the *in vitro* transcribed RNA used in this study (red circles). Six variants were consistently recovered from both the manufacturer data and the in-batch controls, at positions 3350, 6669, 10001, 26791, 26793, 26796. To check whether the remaining within-host variants arose during the SMARTer library prep or during probe capture, we additionally resequenced two replicates of the Twist RNA without capture (blue crosses), by diluting neat RNA 50:50 v/v in Universal Human Reference RNA (UHRR) and taking a proportion for sequencing, to yield approximately 50,000 copies of the Twist control RNA per sample. We generated SMARTer libraries from these replicates, and sequenced these alongside other samples in separate batches. The two capture-free replicates had the same range of intra-sample variants as were observed in our routinely sequenced controls, implying that any differences from the manufacturer data cannot be explained by probe capture and must be the result of the SMARTer library protocol and/or stochastic variation between our laboratory aliquot and the ancestral RNA stock sequenced by Twist.

Tables S1-S5

Table S1. Baseline characteristics of SARS-CoV-2 samples in our dataset collected by participating hospitals in Oxford and Basingstoke, UK, between 8 March and 10 June 2020. Total number of SARS-CoV-2 genomes includes replicate sequences from the same starting material (swab); number of samples corresponds to unique nasopharyngeal swabs. Participant age and sex excludes the 92 samples from anonymous participants in Oxford. Lineages were determined using Pangolin v2.1.7 (1) on 26 January 2021.

	Collecting laboratory	
	Oxford	Basingstoke
SARS-CoV-2 genomes, n(%)	552 (39.7)	838 (60.3)
Samples (swabs), n(%)	539 (41.0)	774 (58.9)
Participants, n(%)	539* (43.0)	727 (57.0)
Proportion female	0.60	0.61
Age, median	48	49
(min - max)	(0 - 98)	(0 - 100)
Sampling date, median	10-Apr-20	09-Apr-20
(min - max)	(16-Mar-2020 - 06-May-2020)	(06-Mar-2020 - 10-Jun-2020)
Ct value, median	22.2	23.2
(min - max)	(13.0 3 - 28.89)	(13.0 - 36.3)
SARS-CoV-2 lineage, n(%) - first sample per participant:		
B.1.1.119	216 (40.07)	261 (35.90)
B.1	73 (13.54)	84 (11.55)
B.1.1.194	1 (0.19)	142 (19.53)
B.1.1.1	38 (7.05)	22 (3.03)
B.1.1.220	3 (0.56)	33 (4.54)
B.1.1.175	20 (3.71)	15 (2.06)
B.40	11 (2.04)	22 (3.03)
B	15 (2.78)	12 (1.65)
B.1.1.10	4 (0.74)	20 (2.75)
B.39	2 (0.37)	21 (2.89)
B.1.1.114	15 (2.78)	2 (0.28)
B.1.1.257	15 (2.78)	1 (0.14)

B.1.1.51	1 (0.19)	14 (1.93)
B.1.13	4 (0.74)	10 (1.38)
B.1.1.289	13 (2.41)	0 (0.00)
B.28	8 (1.48)	4 (0.55)
B.1.391	9 (1.67)	3 (0.41)
B.1.1.217	7 (1.30)	5 (0.69)
B.1.1.269	4 (0.74)	6 (0.83)
B.3	4 (0.74)	5 (0.69)
B.1.1.237	7 (1.30)	0 (0.00)
B.1.321	7 (1.30)	0 (0.00)
B.1.1.270	7 (1.30)	0 (0.00)
B.1.1.104	0 (0.00)	7 (0.96)
B.1.1.164	2 (0.37)	4 (0.55)
B.1.231	6 (1.11)	0 (0.00)
B.1.1.261	5 (0.93)	0 (0.00)
B.1.1.304	5 (0.93)	0 (0.00)
B.1.1.64	4 (0.74)	1 (0.14)
B.1.249	3 (0.56)	0 (0.00)
B.1.379	3 (0.56)	0 (0.00)
B.1.93	2 (0.37)	1 (0.14)
B.1.1.277	0 (0.00)	3 (0.41)
B.1.1.123	2 (0.37)	1 (0.14)
B.31	0 (0.00)	3 (0.41)
B.1.1.90	2 (0.37)	1 (0.14)
B.29	0 (0.00)	2 (0.28)
B.1.147	2 (0.37)	0 (0.00)
B.1.104	2 (0.37)	0 (0.00)
B.1.91	0 (0.00)	2 (0.28)
B.54	2 (0.37)	0 (0.00)
B.1.1.10.2 (L.2)	0 (0.00)	2 (0.28)
B.1.1.273	2 (0.37)	0 (0.00)

B.1.1.208	1 (0.19)	1 (0.14)
B.1.1.49	0 (0.00)	1 (0.14)
B.1.1.211	0 (0.00)	1 (0.14)
B.47	0 (0.00)	1 (0.14)
B.45	1 (0.19)	0 (0.00)
B.35	0 (0.00)	1 (0.14)
B.1.1.106	1 (0.19)	0 (0.00)
B.1.1.127	0 (0.00)	1 (0.14)
B.1.1.166	1 (0.19)	0 (0.00)
B.1.1.180	0 (0.00)	1 (0.14)
B.23	1 (0.19)	0 (0.00)
B.1.98	1 (0.19)	0 (0.00)
B.1.1.183	0 (0.00)	1 (0.14)
B.1.1.216	0 (0.00)	1 (0.14)
B.1.1.71	0 (0.00)	1 (0.14)
B.1.1.256	1 (0.19)	0 (0.00)
B.1.1.264	0 (0.00)	1 (0.14)
B.1.229	1 (0.19)	0 (0.00)
B.1.222	1 (0.19)	0 (0.00)
B.1.199	0 (0.00)	1 (0.14)
B.1.182	0 (0.00)	1 (0.14)
B.1.153	0 (0.00)	1 (0.14)
B.1.1.292	0 (0.00)	1 (0.14)
B.1.1.31	1 (0.19)	0 (0.00)
B.1.1.4	1 (0.19)	0 (0.00)
B.1.1.89	0 (0.00)	1 (0.14)
B.1.1.8	1 (0.19)	0 (0.00)
A.1	0 (0.00)	1 (0.14)
A.2	1 (0.19)	0 (0.00)
A.2.2	0 (0.00)	1 (0.14)

* Includes 92 swabs from anonymous participants

Table S2. Sites masked due to sequenced diversity in Twist synthetic controls (112) or as “highly shared” sites with low-level within-host diversity in more than 20 samples (18). <https://github.com/katrinalythgoe/COVIDdiversity> (70).

Table S3. Identified within-host variable sites.

Sites with at least one minor allele at frequency $\geq 3\%$ at depth of at least 100 reads, in a sample depth $\geq 50,000$ unique mapped reads. Throughout, “samples” refers to all sequencing runs, and therefore includes replicates in the totals. n_notPopConsensus refers to the number of samples in which the minor variant is not the population-level consensus (most common consensus allele); n_SNPs gives the number of SNPs on the tree; homoplasy is “TRUE” if a homoplasy exists on the tree; maf_median is the median minor allele frequency (MAF) for all samples with $>2\%$ MAF; maf_IQR is the inter-quartile range of minor allele frequencies for all samples with $>2\%$ MAF.

<https://github.com/katrinalythgoe/COVIDdiversity> (70).

Table S4. Summary data for the household transmission analysis. Number of intra-host nucleotide variants sites (iSNVs) in source and recipient pairs (minor allele frequency $>3\%$), and number of mapped reads. Stochastic sampling effects elevate the true number of iSNVs in low viral load samples ($<50,000$ mapped reads).

Direction inferred from collection dates (at least 7 days difference)

Household number	iSNVs (source)	iSNVs (recipient)	Mapped reads (source)	Mapped reads (recipient)
1	28	58	[10000,1e+05)	[100,1000)
3	45	0	[100,1000)	[10000,1e+05)
4	6	4	[100,1000)	[1000,10000)
8	0	3	[1e+06,1e+07)	[1e+05,1e+06)
9	1	37	[1e+05,1e+06)	[10000,1e+05)
13	3	2	[1e+05,1e+06)	[10000,1e+05)
14	2	43	[10000,1e+05)	[100,1000)

No direction inferred

Household number	iSNVs (patient 1)	iSNVs (patient 2)	Mapped reads (patient 1)	Mapped reads (patient 2)
2	2	4	[1e+05,1e+06)	[1e+05,1e+06)
5	31	35	[1000,10000)	[10000,1e+05)
6	3	0	[1e+06,1e+07)	[1e+06,1e+07)
7	34	2	[1000,10000)	[100,1000)
10	0	5	[10000,1e+05)	[100,1000)
11	19	6	[10000,1e+05)	[10000,1e+05)
12	1	1	[10000,1e+05)	[1e+05,1e+06)
16	1	4	[1e+05,1e+06)	[10000,1e+05)

Table S5. iSNVs and dn/ds by gene and over the whole genome, counting each variant only once regardless of frequency. All genome positions are relative to the Wuhan-Hu-1 reference sequence. iSNVs at the 18 “highly shared” sites and those identified from the synthetic controls are excluded, as are those in the poly-A tail (positions 29865-29903). Note that due to gene overlap and non-coding intergenic regions, the total number of iSNVs (781) cannot be obtained as the sum of any column in this table, even if the rows for nonstructural proteins in ORF1ab are excluded. * nsp12 overlaps the boundary between ORF1a and ORF1b. ** Intergenic regions are excluded from this row.

Gene	Start	End	Length	Unique iSNVs			Alternative dn/ds (95% CI)
				Total	NS	S	
5'UTR	1	265	265	40	-	-	-
ORF1a	266	13483	13218	205	125	80	0.44 (0.33, 0.58)
nsp1	266	805	540	12	5	7	0.22 (0.06, 0.68)
nsp2	806	2719	1914	36	21	15	0.4 (0.21, 0.79)
nsp3	2720	8554	5835	74	46	28	0.45 (0.29, 0.74)
nsp4	8555	10054	1500	35	21	14	0.43 (0.22, 0.87)
nsp5A	10055	10972	918	10	9	1	2.55 (0.48, 46.97)
nsp6	10973	11842	870	17	10	7	0.39 (0.15, 1.09)
nsp7	11843	12091	249	3	1	2	0.14 (0.01, 1.48)
nsp8	12092	12685	594	7	4	3	0.36 (0.08, 1.83)
nsp9	12686	13024	339	7	4	3	0.41 (0.09, 2.08)
nsp10	13025	13441	417	4	4	0	∞ (0.46, ∞)
nsp12*	13442	16236	2795	42	23	19	0.32 (0.17, 0.59)
ORF1b	13468	21555	8088	127	73	54	0.37 (0.26, 0.52)
nsp13	16237	18039	1803	25	14	11	0.37 (0.17, 0.85)
nsp14	18040	19620	1581	30	16	14	0.3 (0.15, 0.63)
nsp15	19621	20658	1038	17	12	5	0.65 (0.24, 2.04)
nsp16	20659	21552	894	13	8	5	0.43 (0.14, 1.43)
S	21563	25384	3822	56	37	19	0.55 (0.32, 0.98)
ORF3a	25393	26220	828	33	28	5	1.6 (0.68, 4.72)
E	26245	26472	228	3	2	1	0.68 (0.07, 14.66)
M	26523	27191	669	11	7	4	0.54 (0.16, 2.05)
ORF6	27202	27387	186	5	4	1	0.97 (0.14, 18.97)
ORF7a	27394	27759	366	12	10	2	1.47 (0.39, 9.58)
ORF7b	27756	27887	132	4	4	0	∞ (0.41, ∞)
ORF8	27894	28259	366	11	7	4	0.47 (0.14, 1.78)
N	28274	29533	1260	50	33	17	0.58 (0.33, 1.06)
ORF10	29558	29674	117	3	2	1	0.53 (0.05, 11.28)
3'UTR	29675	29903	229	26	-	-	-
All coding regions**	266	29674	29256	519	331	188	0.49 (0.41, 0.59)
Full genome	1	29903	22903	781	-	-	-

Oxford Virus Sequencing Group (OVSG) Analysis Group membership

John A Todd, Tanya Golubchik, David Bonsall, Christophe Fraser, Derrick Crook, Tim Peto, Monique Andersson, Katie Jeffery, David Eyre, Timothy Walker, Robert Shaw, Peter Simmonds, Katrina Lythgoe, Luca Ferretti, Matthew Hall, Mariateresa de Cesare, Paolo Piazza, David Buck, Richard Cornall.

COG-UK Full list of consortium names and affiliations

Funding acquisition, leadership, supervision, metadata curation, project administration, samples, logistics, Sequencing, analysis, and Software and analysis tools:

Thomas R Connor ^{33, 34}, and Nicholas J Loman ¹⁵.

Leadership, supervision, sequencing, analysis, funding acquisition, metadata curation, project administration, samples, logistics, and visualisation:

Samuel C Robson ⁶⁸.

Leadership, supervision, project administration, visualisation, samples, logistics, metadata curation and software and analysis tools:

Tanya Golubchik ²⁷.

Leadership, supervision, metadata curation, project administration, samples, logistics sequencing and analysis:

M. Estee Torok ^{8, 10}.

Project administration, metadata curation, samples, logistics, sequencing, analysis, and software and analysis tools:

William L Hamilton ^{8, 10}.

Leadership, supervision, samples logistics, project administration, funding acquisition sequencing and analysis:

David Bonsall ²⁷.

Leadership and supervision, sequencing, analysis, funding acquisition, visualisation and software and analysis tools:

Ali R Awan ⁷⁴.

Leadership and supervision, funding acquisition, sequencing, analysis, metadata curation, samples and logistics:

Sally Corden³³.

Leadership supervision, sequencing analysis, samples, logistics, and metadata curation:
Ian Goodfellow ¹¹.

Leadership, supervision, sequencing, analysis, samples, logistics, and Project administration:
Darren L Smith ^{60, 61}.

Project administration, metadata curation, samples, logistics, sequencing and analysis:
Martin D Curran ¹⁴, and Surendra Parmar ¹⁴.

Samples, logistics, metadata curation, project administration sequencing and analysis:
James G Shepherd ²¹.

Sequencing, analysis, project administration, metadata curation and software and analysis tools:
Matthew D Parker ³⁸ and Dinesh Aggarwal ^{1, 2, 3}.

Leadership, supervision, funding acquisition, samples, logistics, and metadata curation:
Catherine Moore ³³ .

Leadership, supervision, metadata curation, samples, logistics, sequencing and analysis:
Derek J Fairley^{6, 88}, Matthew W Loose ⁵⁴, and Joanne Watkins ³³.

Metadata curation, sequencing, analysis, leadership, supervision and software and analysis tools:
Matthew Bull ³³ , and Sam Nicholls ¹⁵ .

Leadership, supervision, visualisation, sequencing, analysis and software and analysis tools:
David M Aanensen ^{1, 30}.

Sequencing, analysis, samples, logistics, metadata curation, and visualisation:
Sharon Glaysher ⁷⁰ .

Metadata curation, sequencing, analysis, visualisation, software and analysis tools:
Matthew Bashton ⁶⁰, and Nicole Pacchiarini ³³.

Sequencing, analysis, visualisation, metadata curation, and software and analysis tools:
Anthony P Underwood ^{1, 30}.

Funding acquisition, leadership, supervision and project administration:
Thushan I de Silva ³⁸, and Dennis Wang ³⁸.

Project administration, samples, logistics, leadership and supervision:
Monique Andersson²⁸, Anoop J Chauhan ⁷⁰, Mariateresa de Cesare ²⁶, Catherine Ludden ^{1,3}, and Tabitha W Mahungu ⁹¹.

Sequencing, analysis, project administration and metadata curation:
Rebecca Dewar ²⁰, and Martin P McHugh ²⁰.

Samples, logistics, metadata curation and project administration:
Natasha G Jesudason ²¹, Kathy K Li MBBCh ²¹, Rajiv N Shah ²¹, and Yusri Taha ⁶⁶.

Leadership, supervision, funding acquisition and metadata curation:
Kate E Templeton ²⁰.

Leadership, supervision, funding acquisition, sequencing and analysis:
Simon Cottrell ³³, Justin O'Grady ⁵¹, Andrew Rambaut ¹⁹, and Colin P Smith⁹³.

Leadership, supervision, metadata curation, sequencing and analysis:
Matthew T.G. Holden ⁸⁷, and Emma C Thomson ²¹.

Leadership, supervision, samples, logistics and metadata curation:
Samuel Moses ^{81, 82}.

Sequencing, analysis, leadership, supervision, samples and logistics:
Meera Chand ⁷, Chrystala Constantinidou ⁷¹, Alistair C Darby ⁴⁶, Julian A Hiscox ⁴⁶, Steve Paterson ⁴⁶, and Meera Unnikrishnan ⁷¹.

Sequencing, analysis, leadership and supervision and software and analysis tools:
Andrew J Page ⁵¹, and Erik M Volz ⁹⁶.

Samples, logistics, sequencing, analysis and metadata curation:
Charlotte J Houldcroft ⁸, Aminu S Jahun ¹¹, James P McKenna ⁸⁸, Luke W Meredith ¹¹, Andrew Nelson ⁶¹, Sarojini Pandey ⁷², and Gregory R Young ⁶⁰.

Sequencing, analysis, metadata curation, and software and analysis tools:
Anna Price ³⁴, Sara Rey ³³, Sunando Roy ⁴¹, Ben Temperton⁴⁹, and Matthew Wyles ³⁸.

Sequencing, analysis, metadata curation and visualisation:
Stefan Rooke¹⁹, and Sharif Shaaban ⁸⁷.

Visualisation, sequencing, analysis and software and analysis tools:

Helen Adams³⁵, Yann Bourgeois⁶⁹, Katie F Loveson⁶⁸, Áine O'Toole¹⁹, and Richard Stark⁷¹.

Project administration, leadership and supervision:

Ewan M Harrison^{1,3}, David Heyburn³³, and Sharon J Peacock^{2,3}

Project administration and funding acquisition:

David Buck²⁶, and Michaela John³⁶

Sequencing, analysis and project administration:

Dorota Jamrozy¹, and Joshua Quick¹⁵

Samples, logistics, and project administration:

Rahul Batra⁷⁸, Katherine L Bellis^{1,3}, Beth Blane³, Sophia T Grgis³, Angie Green²⁶, Anita Justice²⁸, Mark Kristiansen⁴¹, and Rachel J Williams⁴¹.

Project administration, software and analysis tools:

Radoslaw Poplawski¹⁵.

Project administration and visualisation:

Garry P Scarlett⁶⁹.

Leadership, supervision, and funding acquisition:

John A Todd²⁶, Christophe Fraser²⁷, Judith Breuer^{40,41}, Sergi Castellano⁴¹, Stephen L Michell⁴⁹, Dimitris Gramatopoulos⁷³, and Jonathan Edgeworth⁷⁸.

Leadership, supervision and metadata curation:

Gemma L Kay⁵¹.

Leadership, supervision, sequencing and analysis:

Ana da Silva Filipe²¹, Aaron R Jeffries⁴⁹, Sascha Ott⁷¹, Oliver Pybus²⁴, David L Robertson²¹, David A Simpson⁶, and Chris Williams³³.

Samples, logistics, leadership and supervision:

Cressida Auckland⁵⁰, John Boyes⁸³, Samir Dervisevic⁵², Sian Ellard^{49,50}, Sonia Goncalves¹, Emma J Meader⁵¹, Peter Muir², Husam Osman⁹⁵, Reenesh Prakash⁵², Venkat Sivaprakasam¹⁸, and Ian B Vipond².

Leadership, supervision and visualisation

Jane AH Masoli^{49,50}.

Sequencing, analysis and metadata curation

Nabil-Fareed Alikhan ⁵¹, Matthew Carlile ⁵⁴, Noel Craine ³³, Sam T Haldenby ⁴⁶, Nadine Holmes ⁵⁴, Ronan A Lyons ³⁷, Christopher Moore ⁵⁴, Malorie Perry ³³, Ben Warne ⁸⁰, and Thomas Williams ¹⁹.

Samples, logistics and metadata curation:

Lisa Berry ⁷², Andrew Bosworth ⁹⁵, Julianne Rose Brown ⁴⁰, Sharon Campbell ⁶⁷, Anna Casey ¹⁷, Gemma Clark ⁵⁶, Jennifer Collins ⁶⁶, Alison Cox ^{43, 44}, Thomas Davis ⁸⁴, Gary Eltringham ⁶⁶, Cariad Evans ^{38, 39}, Clive Graham ⁶⁴, Fenella Halstead ¹⁸, Kathryn Ann Harris ⁴⁰, Christopher Holmes ⁵⁸, Stephanie Hutchings ², Miren Iturriza-Gomara ⁴⁶, Kate Johnson ^{38, 39}, Katie Jones ⁷², Alexander J Keeley ³⁸, Bridget A Knight ^{49, 50}, Cherian Koshy ⁹⁰, Steven Liggett ⁶³, Hannah Lowe ⁸¹, Anita O Lucaci ⁴⁶, Jessica Lynch ^{25, 29}, Patrick C McClure ⁵⁵, Nathan Moore ³¹, Matilde Mori ^{25, 29, 32}, David G Partridge ^{38, 39}, Pinglawathee Madona ^{43, 44}, Hannah M Pymont ², Paul Anthony Randell ^{43, 44}, Mohammad Raza ^{38, 39}, Felicity Ryan ⁸¹, Robert Shaw ²⁸, Tim J Sloan ⁵⁷, and Emma Swindells ⁶⁵.

Sequencing, analysis, Samples and logistics:

Alexander Adams ³³, Hibo Asad ³³, Alec Birchley ³³, Tony Thomas Brooks ⁴¹, Giselda Bucca ⁹³, Ethan Butcher ⁷⁰, Sarah L Caddy ¹³, Laura G Caller ^{2, 3, 12}, Yasmin Chaudhry ¹¹, Jason Coombes ³³, Michelle Cronin ³³, Patricia L Dyal ⁴¹, Johnathan M Evans ³³, Laia Fina ³³, Bree Gatica-Wilcox ³³, Iliana Georgana ¹¹, Lauren Gilbert ³³, Lee Graham ³³, Danielle C Groves ³⁸, Grant Hall ¹¹, Ember Hilvers ³³, Myra Hosmillo ¹¹, Hannah Jones ³³, Sophie Jones ³³, Fahad A Khokhar ¹³, Sara Kumziene-Summerhayes ³³, George MacIntyre-Cockett ²⁶, Rocio T Martinez Nunez ⁹⁴, Caoimhe McKerr ³³, Claire McMurray ¹⁵, Richard Myers ⁷, Yasmin Nicole Panchbhaya ⁴¹, Malte L Pinckert ¹¹, Amy Plimmer ³³, Joanne Stockton ¹⁵, Sarah Taylor ³³, Alicia Thornton ⁷, Amy Trebes ²⁶, Alexander J Trotter ⁵¹, Helena Jane Tutil ⁴¹, Charlotte A Williams ⁴¹, Anna Yakovleva ¹¹ and Wen C Yew ⁶².

Sequencing, analysis and software and analysis tools:

Mohammad T Alam ⁷¹, Laura Baxter ⁷¹, Olivia Boyd ⁹⁶, Fabricia F. Nascimento ⁹⁶, Timothy M Freeman ³⁸, Lily Geidelberg ⁹⁶, Joseph Hughes ²¹, David Jorgensen ⁹⁶, Benjamin B Lindsey ³⁸, Richard J Orton ²¹, Manon Ragonnet-Cronin ⁹⁶ Joel Southgate ^{33, 34}, and Sreenu Vattipally ²¹.

Samples, logistics and software and analysis tools:

Igor Starinskij ²³.

Visualisation and software and analysis tools:

Joshua B Singer ²¹, Khalil Abudahab ^{1, 30}, Leonardo de Oliveira Martins ⁵¹, Thanh Le-Viet ⁵¹, Mirko Menegazzo ³⁰, Ben EW Taylor ^{1, 30}, and Corin A Yeats ³⁰.

Project Administration:

Sophie Palmer ³, Carol M Churcher ³, Alisha Davies ³³, Elen De Lacy ³³, Fatima Downing ³³, Sue Edwards ³³, Nikki Smith ³⁸, Francesc Coll ⁹⁷, Nazreen F Hadjirin ³ and Frances Bolt ^{44, 45}.

Leadership and supervision:

Alex Alderton¹, Matt Berriman¹, Ian G Charles ⁵¹, Nicholas Cortes ³¹, Tanya Curran ⁸⁸, John Danesh¹, Sahar Eldirdiri ⁸⁴, Ngozi Elumogo ⁵², Andrew Hattersley ^{49, 50}, Alison Holmes ^{44, 45}, Robin Howe ³³, Rachel Jones ³³, Anita Kenyon ⁸⁴, Robert A Kingsley ⁵¹, Dominic Kwiatkowski ^{1, 9}, Cordelia Langford¹, Jenifer Mason⁴⁸, Alison E Mather ⁵¹, Lizzie Meadows ⁵¹, Sian Morgan ³⁶, James Price ^{44, 45}, Trevor I Robinson ⁴⁸, Giri Shankar ³³, John Wain ⁵¹, and Mark A Webber ⁵¹.

Metadata curation:

Declan T Bradley ^{5, 6}, Michael R Chapman ^{1, 3, 4}, Derrick Crooke ²⁸, David Eyre ²⁸, Martyn Guest ³⁴, Huw Gulliver ³⁴, Sarah Hoosdally ²⁸, Christine Kitchen ³⁴, Ian Merrick ³⁴, Siddharth Mookerjee ^{44, 45}, Robert Munn ³⁴, Timothy Peto ²⁸, Will Potter ⁵², Dheeraj K Sethi ⁵², Wendy Smith ⁵⁶, Luke B Snell ^{75, 94}, Rachael Stanley ⁵², Claire Stuart ⁵² and Elizabeth Wastenge²⁰.

Sequencing and analysis:

Erwan Acheson ⁶, Safiah Afifi ³⁶, Elias Allara ^{2, 3}, Roberto Amato ¹, Adrienn Angyal ³⁸, Elihu Aranday-Cortes ²¹, Cristina Ariani ¹, Jordan Ashworth ¹⁹, Stephen Attwood ²⁴, Alp Aydin ⁵¹, David J Baker ⁵¹, Carlos E Balcazar ¹⁹, Angela Beckett ⁶⁸, Robert Beer ³⁶, Gilberto Betancor ⁷⁶, Emma Betteridge ¹, David Bibby ⁷, Daniel Bradshaw⁷, Catherine Bresner ³⁴, Hannah E Bridgewater ⁷¹, Alice Broos ²¹, Rebecca Brown ³⁸, Paul E Brown ⁷¹, Kirstyn Brunker ²², Stephen N Carmichael ²¹, Jeffrey K. J. Cheng ⁷¹, Dr Rachel Colquhoun ¹⁹, Gavin Dabrera ⁷, Johnny Debebe ⁵⁴, Eleanor Drury ¹, Louis du Plessis ²⁴, Richard Eccles ⁴⁶, Nicholas Ellaby ⁷, Audrey Farbos ⁴⁹, Ben Farr ¹, Jacqueline Findlay ⁴¹, Chloe L Fisher ⁷⁴, Leysa Marie Forrest ⁴¹, Sarah Francois ²⁴, Lucy R. Frost ⁷¹, William Fuller³⁴, Eileen Gallagher ⁷, Michael D Gallagher ¹⁹, Matthew Gemmell ⁴⁶, Rachel AJ Gilroy ⁵¹, Scott Goodwin ¹, Luke R Green ³⁸, Richard Gregory ⁴⁶, Natalie Groves ⁷, James W Harrison ⁴⁹, Hassan Hartman ⁷, Andrew R Hesketh ⁹³, Verity Hill ¹⁹, Jonathan Hubb ⁷, Margaret Hughes⁴⁶, David K Jackson ¹, Ben Jackson ¹⁹, Keith James ¹, Natasha Johnson ²¹, Ian Johnston ¹, Jon-Paul Keatley ¹, Moritz Kraemer ²⁴, Angie Lackenby ⁷, Mara Lawniczak ¹, David Lee ⁷, Rich Livett ¹, Stephanie Lo ¹, Daniel Mair ²¹, Joshua Maksimovic ³⁶, Nikos Manesis ⁷, Robin Manley ⁴⁹, Carmen Manso ⁷, Angela Marchbank ³⁴, Inigo Martincorena ¹, Tamyo Mbisa ⁷, Kathryn McCluggage ³⁶, JT McCrone ¹⁹, Shahjahan Miah ⁷, Michelle L Michelsen ⁴⁹, Mari Morgan ³³, Gaia Nebbia ⁷⁸, Charlotte Nelson ⁴⁶, Jenna Nichols ²¹, Paola Niola ⁴¹, Kyriaki Nomikou ²¹, Steve Palmer ¹, Naomi Park ¹, Yasmin A Parr ¹, Paul J Parsons ³⁸, Vineet Patel ⁷, Minal Patel ¹, Clare Pearson ^{2, 1}, Steven Platt ⁷, Christoph Puethe ¹, Mike Quail ¹, Jayna Raghwani ²⁴, Lucille Rainbow ⁴⁶, Shavanti Rajatileka ¹, Mary Ramsay ⁷, Paola C Resende Silva ^{41, 42}, Steven Rudder ⁵¹, Chris Ruis ³, Christine M Sambles ⁴⁹, Fei Sang ⁵⁴, Ulf Schaefer⁷, Emily Scher ¹⁹, Carol Scott ¹, Lesley Shirley ¹, Adrian W Signell ⁷⁶, John Sillitoe ¹, Christen Smith ¹, Dr Katherine L Smollett ²¹, Karla Spellman ³⁶

,Thomas D Stanton ¹⁹, David J Studholme ⁴⁹, Grace Taylor-Joyce ⁷¹, Ana P Tedim ⁵¹, Thomas Thompson ⁶, Nicholas M Thomson ⁵¹, Scott Thurston¹, Lily Tong ²¹, Gerry Tonkin-Hill ¹, Rachel M Tucker ³⁸, Edith E Vamos ⁴, Tetyana Vasylyeva²⁴, Joanna Warwick-Dugdale ⁴⁹, Danni Weldon ¹, Mark Whitehead ⁴⁶, David Williams ⁷, Kathleen A Williamson ¹⁹,Harry D Wilson ⁷⁶,Trudy Workman ³⁴, Muhammad Yasir⁵¹, Xiaoyu Yu ¹⁹, and Alex Zarebski ²⁴.

Samples and logistics:

Evelien M Adriaenssens ⁵¹, Shazaad S Y Ahmad ^{2,47} , Adela Alcolea-Medina ^{59, 77}, John Allan ⁶⁰, Patawee Asamaphan ²¹, Laura Atkinson ⁴⁰, Paul Baker ⁶³, Jonathan Ball ⁵⁵, Edward Barton⁶⁴, Mathew A Beale¹, Charlotte Beaver¹, Andrew Beggs ¹⁶, Andrew Bell ⁵¹, Duncan J Berger ¹, Louise Berry. ⁵⁶, Claire M Bewshea ⁴⁹, Kelly Bicknell ⁷⁰, Paul Bird ⁵⁸, Chloe Bishop ⁷ , Tim Boswell ⁵⁶, Cassie Breen ⁴⁸, Sarah K Buddenborg¹, Shirelle Burton-Fanning ⁶⁶ , Vicki Chalker ⁷, Joseph G Chappell ⁵⁵, Themoula Charalampous ^{78, 94}, Claire Cormie³, Nick Cortes²⁹, ²⁵, Lindsay J Coupland ⁵², Angela Cowell ⁴⁸ , Rose K Davidson ⁵³, Joana Dias ³, Maria Diaz ⁵¹ , Thomas Dibling¹, Matthew J Dorman¹, Nichola Duckworth⁵⁷, Scott Elliott⁷⁰, Sarah Essex⁶³, Karlie Fallon ⁵⁸ , Theresa Feltwell ⁸, Vicki M Fleming ⁵⁶, Sally Forrest ³, Luke Foulser¹, Maria V Garcia-Casado¹, Artemis Gavriil ⁴¹, Ryan P George ⁴⁷, Laura Gifford ³³, Harmeet K Gill ³, Jane Greenaway ⁶⁵, Luke Griffith⁵³, Ana Victoria Gutierrez⁵¹, Antony D Hale ⁸⁵, Tanzina Haque ⁹¹, Katherine L Harper ⁸⁵, Ian Harrison ⁷ , Judith Heaney ⁸⁹, Thomas Helmer ⁵⁸, Ellen E Higginson³ , Richard Hopes ², Hannah C Howson-Wells ⁵⁶, Adam D Hunter ¹, Robert Impey ⁷⁰, Dianne Irish-Tavares ⁹¹, David A Jackson¹ , Kathryn A Jackson ⁴⁶, Amelia Joseph ⁵⁶, Leanne Kane ¹, Sally Kay ¹, Leanne M Kermack ³, Manjinder Khakh ⁵⁶, Stephen P Kidd ^{29, 25, 31}, Anastasia Kolyva ⁵¹, Jack CD Lee ⁴⁰, Laura Letchford ¹ , Nick Levene ⁷⁹, Lisa J Levett ⁸⁹, Michelle M Lister ⁵⁶, Allyson Lloyd ⁷⁰, Joshua Loh ⁶⁰ , Louissa R Macfarlane-Smith ⁸⁵, Nicholas W Machin ^{2, 47}, Mailis Maes ³, Samantha McGuigan ¹, Liz McMinn ¹, Lamia Mestek-Boukhibar ⁴¹, Zoltan Molnar ⁶, Lynn Monaghan ⁷⁹, Catrin Moore ²⁷, Plamena Naydenova ³, Alexandra S Neaverson ¹, Rachel Nelson ¹, Marc O Niebel ²¹ , Elaine O'Toole⁴⁸ , Debra Padgett ⁶⁴, Gaurang Patel ¹ , Brendan AI Payne ⁶⁶, Liam Prestwood ¹, Veena Raviprakash ⁶⁷, Nicola Reynolds⁸⁶, Alex Richter ¹⁶, Esther Robinson ⁹⁵, Hazel A Rogers¹, Aileen Rowan ⁹⁶, Garren Scott ⁶⁴, Divya Shah ⁴⁰, Nicola Sheriff ⁶⁷, Graciela Sluga, Emily Souster¹, Michael Spencer-Chapman¹, Sushmita Sridhar ^{1, 3}, Tracey Swingler ⁵³, Julian Tang⁵⁸, Graham P Taylor⁹⁶, Theocharis Tsoleridis ⁵⁵, Lance Turtle⁴⁶, Sarah Walsh ⁵⁷, Michelle Wantoch ⁸⁶, Joanne Watts ⁴⁸ , Sheila Waugh ⁶⁶, Sam Weeks⁴¹, Rebecca Williams³¹, Iona Willingham⁵⁶, Emma L Wise ^{25, 29, 31}, Victoria Wright ⁵⁴ , Sarah Wyllie ⁷⁰, and Jamie Young ³.

Software and analysis tools

Amy Gaskin³³, Will Rowe ¹⁵, and Igor Siveroni ⁹⁶.

Visualisation:

Robert Johnson ⁹⁶.

1 Wellcome Sanger Institute, **2** Public Health England, **3** University of Cambridge, **4** Health Data Research UK, Cambridge, **5** Public Health Agency, Northern Ireland ,**6** Queen's University Belfast **7** Public Health England Colindale, **8** Department of Medicine, University of Cambridge, **9** University of Oxford, **10** Departments of Infectious Diseases and Microbiology, Cambridge University Hospitals NHS Foundation Trust; Cambridge, UK, **11** Division of Virology, Department of Pathology, University of Cambridge, **12** The Francis Crick Institute, **13** Cambridge Institute for Therapeutic Immunology and Infectious Disease, Department of Medicine, **14** Public Health England, Clinical Microbiology and Public Health Laboratory, Cambridge, UK, **15** Institute of Microbiology and Infection, University of Birmingham, **16** University of Birmingham, **17** Queen Elizabeth Hospital, **18** Heartlands Hospital, **19** University of Edinburgh, **20** NHS Lothian, **21** MRC-University of Glasgow Centre for Virus Research, **22** Institute of Biodiversity, Animal Health & Comparative Medicine, University of Glasgow, **23** West of Scotland Specialist Virology Centre, **24** Dept Zoology, University of Oxford, **25** University of Surrey, **26** Wellcome Centre for Human Genetics, Nuffield Department of Medicine, University of Oxford, **27** Big Data Institute, Nuffield Department of Medicine, University of Oxford, **28** Oxford University Hospitals NHS Foundation Trust, **29** Basingstoke Hospital, **30** Centre for Genomic Pathogen Surveillance, University of Oxford, **31** Hampshire Hospitals NHS Foundation Trust, **32** University of Southampton, **33** Public Health Wales NHS Trust, **34** Cardiff University, **35** Betsi Cadwaladr University Health Board, **36** Cardiff and Vale University Health Board, **37** Swansea University, **38** University of Sheffield, **39** Sheffield Teaching Hospitals, **40** Great Ormond Street NHS Foundation Trust, **41** University College London, **42** Oswaldo Cruz Institute, Rio de Janeiro **43** North West London Pathology, **44** Imperial College Healthcare NHS Trust, **45** NIHR Health Protection Research Unit in HCAI and AMR, Imperial College London, **46** University of Liverpool, **47** Manchester University NHS Foundation Trust, **48** Liverpool Clinical Laboratories, **49** University of Exeter, **50** Royal Devon and Exeter NHS Foundation Trust, **51** Quadram Institute Bioscience, University of East Anglia, **52** Norfolk and Norwich University Hospital, **53** University of East Anglia, **54** Deep Seq, School of Life Sciences, Queens Medical Centre, University of Nottingham, **55** Virology, School of Life Sciences, Queens Medical Centre, University of Nottingham, **56** Clinical Microbiology Department, Queens Medical Centre, **57** PathLinks, Northern Lincolnshire & Goole NHS Foundation Trust, **58** Clinical Microbiology, University Hospitals of Leicester NHS Trust, **59** Viapath, **60** Hub for Biotechnology in the Built Environment, Northumbria University, **61** NU-OMICS Northumbria University, **62** Northumbria University, **63** South Tees Hospitals NHS Foundation Trust, **64** North Cumbria Integrated Care NHS Foundation Trust, **65** North Tees and Hartlepool NHS Foundation Trust, **66** Newcastle Hospitals NHS Foundation Trust, **67** County Durham and Darlington NHS Foundation Trust, **68** Centre for Enzyme Innovation, University of Portsmouth, **69** School of Biological Sciences, University of Portsmouth, **70** Portsmouth Hospitals NHS Trust, **71** University of Warwick, **72** University Hospitals Coventry and Warwickshire, **73** Warwick Medical School and Institute of Precision Diagnostics, Pathology, UHCW NHS Trust, **74** Genomics Innovation Unit, Guy's and St. Thomas' NHS Foundation Trust, **75** Centre for Clinical Infection & Diagnostics Research, St. Thomas' Hospital and Kings College London, **76** Department of Infectious Diseases, King's College London, **77** Guy's and St. Thomas' Hospitals NHS Foundation Trust, **78** Centre for Clinical Infection and Diagnostics Research, Department of Infectious Diseases, Guy's and St Thomas' NHS Foundation Trust, **79** Princess Alexandra Hospital Microbiology Dept. , **80** Cambridge University Hospitals NHS Foundation Trust, **81** East Kent Hospitals University NHS Foundation Trust, **82** University of Kent, **83** Gloucestershire Hospitals NHS Foundation Trust, **84** Department of Microbiology, Kettering General Hospital, **85** National Infection Service, PHE and Leeds Teaching Hospitals Trust, **86** Cambridge Stem Cell Institute, University of Cambridge, **87** Public Health Scotland, **88** Belfast Health & Social Care Trust, **89** Health Services Laboratories, **90** Barking, Havering and Redbridge University Hospitals NHS Trust, **91** Royal Free NHS Trust, **92** Maidstone and Tunbridge Wells NHS Trust, **93** University of Brighton, **94** Kings College London, **95** PHE Heartlands, **96** Imperial College London, **97** Department of Infection Biology, London School of Hygiene and Tropical Medicine.

References and Notes

1. A. Rambaut, E. C. Holmes, Á. O'Toole, V. Hill, J. T. McCrone, C. Ruis, L. du Plessis, O. G. Pybus, A dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. *Nat. Microbiol.* **5**, 1403–1407 (2020). [doi:10.1038/s41564-020-0770-5](https://doi.org/10.1038/s41564-020-0770-5) Medline
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