

Northumbria Research Link

Citation: Stephenson, Kimberly, Wilson, Byron, Taylor, Michael, McLaren, Kurt, van Veen, Rick, Kunna, John and Campbell, Jayaka (2022) Modelling Climate Change Impacts on Tropical Dry Forest Fauna. Sustainability, 14 (8). p. 4760. ISSN 2071-1050

Published by: MDPI

URL: <https://doi.org/10.3390/su14084760> <<https://doi.org/10.3390/su14084760>>

This version was downloaded from Northumbria Research Link:
<http://nrl.northumbria.ac.uk/id/eprint/49155/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)

Article

Modelling Climate Change Impacts on Tropical Dry Forest Fauna

Kimberly Stephenson ^{1,2,3,*}, Byron Wilson ³, Michael Taylor ², Kurt McLaren ^{3,4}, Rick van Veen ³, John Kunna ³ and Jayaka Campbell ² 

- ¹ School of Environmental Sciences, University of Guelph, 50 Stone Rd E., Guelph, ON N1G 2W1, Canada
² Climate Studies Group Mona (CSGM), Department of Physics, University of the West Indies, Kingston 7, Jamaica; michael.taylor@uwimona.edu.jm (M.T.); jayaka.campbell02@uwimona.edu.jm (J.C.)
³ Department of Life Sciences, University of the West Indies, Kingston 7, Jamaica; grahami1919@gmail.com (B.W.); kurt.mclaren@northumbria.ac.uk (K.M.); rick_vanveen@hotmail.com (R.v.V.); john.kunna@gmail.com (J.K.)
⁴ Department of Geography and Environmental Sciences, Northumbria University, Ellison Place, Newcastle upon Tyne NE1 8ST, UK
* Correspondence: kimannstephenson@gmail.com

Abstract: Tropical dry forests are among the most threatened ecosystems in the world, and those occurring in the insular Caribbean are particularly vulnerable. Climate change represents a significant threat for the Caribbean region and for small islands like Jamaica. Using the Hellshire Hills protected area in Jamaica, a simple model was developed to project future abundance of arthropods and lizards based on current sensitivities to climate variables derived from rainfall and temperature records. The abundances of 20 modelled taxa were predicted more often by rainfall variables than temperature, but both were found to have strong impacts on arthropod and lizard abundance. Most taxa were projected to decrease in abundance by the end of the century under drier and warmer conditions. Where an increase in abundance was projected under a low emissions scenario, this change was reduced or reversed under a high emissions climate change scenario. The validation process showed that, even for a small population, there was reasonable skill in predicting its annual variability. Results of this study show that this simple model can be used to identify the vulnerability of similar sites to the effects of shifting climate and, by extension, their conservation needs.

Keywords: tropical dry forest; biodiversity; bioclimatic modelling; Caribbean



Citation: Stephenson, K.; Wilson, B.; Taylor, M.; McLaren, K.; van Veen, R.; Kunna, J.; Campbell, J. Modelling Climate Change Impacts on Tropical Dry Forest Fauna. *Sustainability* **2022**, *14*, 4760. <https://doi.org/10.3390/su14084760>

Academic Editor: Liubov Volkova

Received: 21 January 2022

Accepted: 13 April 2022

Published: 15 April 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Climate change is an important issue for the small island states of the Caribbean due to an inherent climate sensitivity attributable to small size, location in the Atlantic hurricane belt, and other factors [1]. The Caribbean's climate is already changing, as seen by rising temperatures, increasingly variable rainfall, increasing drought and flood frequency, rising sea levels, and intensifying storms [1–4]. Modelling studies project that, by the end of the current century, the region will experience an increase in intensity of rainfall events; a decrease in total annual rainfall of up to 40% in some places and particularly during the climatologically wettest period of May to October; warming of up to 4 °C, with night-time temperatures increasing at a faster rate than daytime; more intense hurricanes; more frequent floods and droughts; and sea level rise of up to 1 m [2,5–9]. Being a tropical region, the Caribbean is also projected to experience a climate departure characterized by extreme climate shifts far sooner than other regions [10]. For example, by the mid-2020s it is projected that annual minimum temperatures in some years may be higher than historical maximum temperatures [10].

Although over 9% of neotropical dry forests occur in the insular Caribbean, approximately 66% of that dry forest cover has been lost and much of what remains is at least

somewhat fragmented or degraded [11]. Tropical dry forests (TDFs) are highly sensitive to shifts in climate, but have been the focus of very few studies on the ecological impacts of climate change [12]. In these ecosystems, rainfall has a strong influence on seedling survival, species composition, and above-ground biomass, which can be significantly changed or impaired by drought occurrence [13–17]. Hurricanes also significantly impact survival of mature vegetation and species composition of mature TDFs. In 2015, Hurricane Patricia made landfall as a category 4 cyclone on the Pacific coast of Mexico. In assessing the damage to the Chamela-Cuixmala TDF, Jimenez-Rodríguez et al. (2018) identified that more severe damage occurred in old growth areas than in secondary forest, and the forest composition changed to favor smaller plants that could regenerate quickly [18]. Therefore, climate change will likely result in a shift in the forest composition, ecosystem dynamics, and extinction risk of many species in TDFs, which are already the most threatened ecosystems in the world [12,19]. Characterizing the impacts of climate change on dry forests and other biodiverse Caribbean ecosystems is a necessary first step for planning and promoting actions aimed at the conservation of the region's biodiversity, particularly in the minimally studied dry forests of the region [12,19,20].

Despite being a hotspot for biodiversity, the impact of climate change on Caribbean ecosystems has not been extensively studied. This may be due to several reasons including (i) the low importance placed on biodiversity, (ii) data limitations, (iii) the lack of pristine study sites, and (iv) the lack of suitable/well developed methodologies [21]. Few studies have focused on the region's terrestrial faunal species [20]. Bioclimatic modelling, in particular, has been applied only minimally in the insular Caribbean, as the small sizes of the islands limit their accurate representation within many of the existing models [20]. Additionally, only limited work has previously examined vulnerability to climatic influence (in particular rainfall and temperature) as well as survival potential and extinction risk in important tropical dry forest habitats such as the Hellshire Hills, Jamaica or forests like it. These include studies examining (i) rainfall as a predictor of *Anolis lineatopus* hatchling numbers and juvenile growth [22]; (ii) the link between lizard extinction and temperature [23]; and (iii) seedling dynamics and climate variability [24].

Previous studies have shown that climate is a strong modulator of ectotherm populations, which further emphasizes the need to characterize potential impacts of climate change in these sensitive hotspots. Rainfall and relative humidity are important factors in the life cycles of arthropods and small lizards. Low moisture availability can retard the life cycle of the organism by impairing oviposition, juvenile recruitment, growth and development, and physical function, while high relative humidity can increase pathogen infection [25–27]. Studies assessing the variability of arthropod populations as a function of rainfall seasonality in Grenada in 1977 showed that arthropod populations were 2.3 times larger and 3.1 times greater in biomass during the wet season (June–December) than during the dry season (January–May) [28]. The influence of rainfall on arthropod abundance also impacts food availability for lizards, which has been found to be a limitation of juvenile growth in Jamaican *Anolis lineatopus* [22]. Moisture is also critical to anoles for egg laying and development, and has been identified as a determining factor in the geographical variation in morphology of *Anolis lineatopus* and *A. grahmi* [22,25,29,30]. These factors indicate that rainfall is linked to anole abundance both directly through provision of water and indirectly through its modulation of prey availability. Water availability plays a large part in microhabitat choice in invertebrates and lizards, which tend to occupy shaded areas during dry periods in order to minimize water loss and thermal stress [31–33].

Tropical arthropods and lizards also exhibit high thermal sensitivity that is not minimized by their tolerance of warm habitats. Warmer temperatures alter the rate of development for many arthropods, which can result in fluctuating population size due to changes in breeding patterns, survival, and food availability [27,34,35]. There is, however, a temperature threshold for growth and development, which may be different for adults and nymphs of the same species [36]. Experiments conducted on *Nilaparvata lugens*, one of the most potent rice pests globally, showed that adults consistently had higher critical thermal

maximum (C_{max}), upper lethal temperature (ULT), and heat coma temperature (HCT) than nymphs [37]. Given recent global temperature trends, particularly in the tropics where the species is most prevalent, nymphs are currently very close to exceeding their optimal temperatures [37]. For tropical lizards, studies of thermal vulnerability have found that living in a warm habitat does not remove the possibility of intolerance to high temperatures, given their sensitivity to extreme heating due to their thermoconforming nature [30,38]. Thermal responses are more dependent on general behavior of the species, as basking species living in open habitats tend to maintain high body and operative temperatures relative to shaded habitat non-basking species, which are often active at lower temperatures that are also closer to ambient temperatures [30,38]. Also, the relatively narrow annual temperature range to which tropical groups are accustomed adds to their vulnerability to overheating and reduced activity under substantive rises in ambient temperature outside of their preferred temperature ranges, particularly during summer months [30,38].

Historical relationships between ectotherms and climate show that climate change will lead to evolution in physiology, shifts in climate niches, migration, range expansion or contraction, and alteration of trophic interactions and predator-prey relationships [30,34,39–42]. Some tropical species do, however, have natural physiological responses that confer adaptive capacity, such as the heat-hardening capacity in lizards that allows for survival in extreme temperatures [40,43]. It must be noted, however, that heat hardening varies geographically and does not negate negative impacts of extreme heat on locomotor function [43].

The overall aim of this study is to investigate the potential role of climate in determining faunal abundance in a small dry forest located in Hellshire Hills, Jamaica. We characterize the climate of the Hellshire Hills dry forest, and investigate whether there are discernible links among climate, arthropod, and lizard abundance, and whether the relationships found can be used to develop a simple bioclimatic model representative of the region. The validated simple model is then used to infer information about the impact of projected future changes in the climate of the Caribbean on the biodiversity of the region. Given the dearth of studies examining climatic impacts on regional fauna, this will be a useful addition to regional conservation efforts.

2. Materials and Methods

2.1. Study Site

The study area is the dry limestone forest of the Hellshire Hills in Jamaica. The remote location and rugged topography of the region limit human intrusion and disturbances, and potentially enhance the likelihood of identifying a climate signal in the biodiversity data. The Hellshire Hills dry forest is located on the south-eastern coast of Jamaica in the parish of St. Catherine, to the west of the capital city of Kingston (Figure 1). The area is heavily karstified and covered by tropical dry forest with very little soil cover that is concentrated in sporadic depressions [16]. Jamaica's dry forests are areas of high endemism and serve as a habitat for many of the island's endangered species. For example, the Hellshire Hills harbors the only wild population of the Critically Endangered Jamaican Iguana (*Cyclura collei*), and also supports the rare Jamaican blue-tailed galliwasp (*Celestus duquesneyi*) [44]. The dry forest is largely undisturbed in central and western sections [45]. It is this relatively undisturbed characteristic that makes the area suitable for the study being undertaken.

The area is located in the driest part of the island. It receives less than 1000 mm per year, 85% of which occurs in the main rainy season between August and November. Daily temperatures climb to an average of 27 °C [45]. Vegetation in the area is adapted to the arid conditions and limited soil cover, and is composed primarily of dry evergreens, with mangroves along the coast. The forest comprises both canopy and sub-canopy species of varying growth rates, with seedling survival being heavily modulated by both rainfall and canopy coverage [15,16].

Of significance, the Hellshire Hills form part of the Portland Bight Protected Area (Figure 1). It is considered 'an irreplaceable site of environmental importance' in the

Caribbean and a ‘hotspot within a hotspot’, because it is the largest remaining intact dry limestone forest in the region, a site of international significance, and a key biodiversity hotspot within the larger Caribbean hotspot [46–48]. The Hellshire Hills forest has been the subject of extensive efforts to document the abundance of species, spearheaded by the Jamaican Iguana Recovery Group [16,44,45].

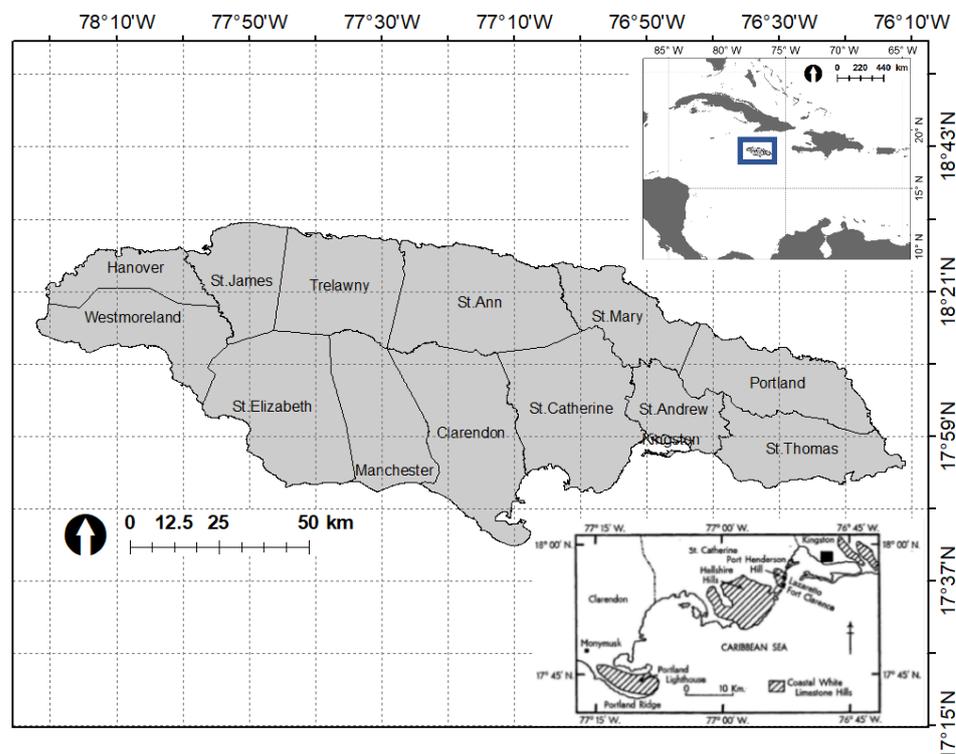


Figure 1. Map of Jamaica with insets showing Jamaica’s position in the Caribbean (top right) and the Hellshire Hills, which is located ~19 km west of the Palisadoes (bottom right). Source of bottom right inset: [49].

2.2. Abundance Data

Collection of abundance data was carried out using an existing system of live (dry) pitfall traps established in 1997 which is detailed in Wilson and Vogel (2000) [45]. The traps were established as a part of the wider experiment mounted by the Jamaican Iguana Recovery Group at the University of the West Indies to survey the herpetofauna of the Hellshire Hills [45]. The traps were established in four grids, each with eight pairs of buckets submerged in the ground and connected by drift fences. Of these four grids, two were experimental plots (that is, $2 \times E$ grids) in which box trapping was used to control invasive predators such as mongooses and feral cats, and two were control plots (C grids) in which no IAS (invasive alien species) trapping was conducted. Data from the control plots (that is, $2 \times C$ grids) were initially analysed separately from experimental plots (E grids) due to the differences in predator treatment. These were later combined for model creation, as no significant difference was found between the patterns of variability of the C and E grids, despite there being a difference in number of animals captured in each grid. As with any trapping technique, there are inherent biases in pitfall trapping, such as the affects of habitat structure, behavior of individual species related to likelihood of trapping and/or potential for escape from pitfall buckets, in addition to the risk of in-trap predation. We acknowledge that some species may be more or less terrestrial or arboreal, and not unrelated, that some species may be more or less able to jump or climb out of the pitfall buckets. However, we regard these potential sources of trap bias as negligible, especially in regard to the temporal patterns that are the focus of the present study.

Genera and species designations are given when available in Table 1. Genera and common names are provided where available, along with codes used for ease of documentation. These codes were also used when building the simple models. All focal lizard species in this study are endemic to Jamaica [50]. The abundance data collected are not available for public use. Statistical analyses and modelling were performed using the R programming language [51].

Table 1. Morphotaxon codes used throughout analysis (genera are provided where available).

Arthropods	
dbcrick	<i>Ceuthophilus</i> sp.
dhrick	Orthoptera: Gryllidae
cent	Chilopoda
hc	<i>Coenobita clypeatus</i>
jsf	Collembola
pb	<i>Armadillidium vulgare</i>
rapb	<i>Dysdercus andreae</i>
bws	<i>Phrynus</i> sp.
Scorp	Scorpiones
sf	Lepismatidae
solpu	<i>Ammotrechula pilosa</i>
va	<i>Dasymutilla militaris</i>
1bbug	<i>Pangaeus bilineatus</i>
2bbug	<i>Periplaneta</i> sp.
1bb	<i>Tarpela mutabilis</i>
2bb	<i>Branchus jamaicensis</i>
sbb	Coleoptera: Chrysomelidae
Lizards	
al	<i>Anolis lineatopus</i>
ag	<i>Anolis grahami</i>
cc	<i>Celestus cruscus</i>
cd	<i>Celestus duquesneyi</i>
mm	<i>Spondylurus fulgidus</i>

2.3. Climate Data

To characterize the climate of the Hellshire Hills region, data from an onsite daily weather station over the period 2010–2012 were aggregated into monthly data and an annual climatology. The weather station was installed at field station ‘South Camp’ at a height of 1.8 m and at an elevation of 126 m, at coordinates 17.87° N 76.96° W.

Because of the limited length of the Hellshire Hills station data, weather station data from a nearby site with similar climate characteristics were also used. The closest station with records of appreciable length was located at the Norman Manley International Airport approximately 19 km away. Daily rainfall and temperature data for the period 1997–2013 were obtained from the Meteorological Service of Jamaica. This period overlapped with the abundance data and allowed for a lag in linkages to be explored.

Norman Manley was established as a proxy for the Hellshire Hills (hereafter referred to as NM and HH, respectively) using Pearson correlations and generalized additive mixed modelling (GAMM) for the overlapping periods of the two datasets (2010–2012). GAMM was applied to daily rainfall and temperature anomalies using the ‘mgcv’ package [52] in R, implementing a gaussian distribution with an identity link function and autoregressive moving average (ARMA) error structures to account for autocorrelation in the timeseries. To determine whether the HH and NM relationships identified by the GAMM runs would be retained, ARMA and model parameters were checked to ensure that 95% confidence limits did not include zero, and autocorrelation function (ACF) and partial autocorrelation function (PACF) were used to assess whether model residuals were random and not autocorrelated. No model parameters were found to pass through zero, and ACF and PACF

were found to be within an acceptable range of variation, thereby verifying good model fit and validating the use of NM as a proxy for HH in this study.

A suite of relevant local climate indices was derived from the NM data for use in the HH model (Appendix A). Climate indices are known to best capture potential patterns of modulation at the local scale [4,53,54]. Abundance is also known to show stronger similarities in patterns of variation with extreme indices and seasonal variables rather than monthly or daily climate variables [55]. Climate seasonality was analyzed in anticipation that species abundance may be more related to climatic variations in one part of the year (e.g., the early or late rainy season), than in another part of the year [22,56]. The seasonality of Jamaica is often defined in terms of rainfall abundance and is commonly noted as consisting of (i) two 3-month dry periods occurring in November–December–January (NDJ) and February–March–April (FMA); and (ii) two wet periods occurring in May–June–July (MJJ) (early) and August–September–October (ASO) (late) wet seasons [57]. This study shifted these seasons slightly, in keeping with linkages previously identified in the literature for the focal taxa and site, such as the inclusion of November in the late wet season influence and the effect of the December–January period on feeding and survival [22]. The shift, however, maintains the overall pattern of variability (i.e., dry versus wet seasons). The seasons studied were MJJ (early wet season), ASON (late wet season) and DJF (dry season).

It is important to note that, although the climate of Jamaica is known to be linked to global climatic fluctuations such as ENSO [58], indices representing these large-scale drivers were not used as predictors in the model being created. Firstly, it is assumed that the influence of the global predictors will be through a change in local climate variables such as rainfall or temperature (see for example [58–60]), Secondly, the goal is to use the simple model being created to infer the influence of future climate changes on the abundance, and there remain challenges with respect to projecting characteristics of these large scale fluctuations in the future (see for example [61]).

2.4. Model Creation

The strength of the climate signal in arthropod and lizard abundance was evaluated using NM data by means of graphs, Spearman's rank correlation coefficients, and backward regression [22,62]. Using the same methods, relationships among all species (i.e., species-species relationships) were also evaluated. The evaluations served as the primary guidance for the creation of the simple model for the HH region by (i) establishing that relationships exist between climate and species, and between species, and (ii) providing guidance for potential predictor variables (climate or species) for each taxon.

A simple model of climate-species and species-species relationships for the Hellshire Hills was created based on the strength of statistical relationships identified. Model creation for each arthropod and lizard species was conducted using abundance data for the period 1997–2009 and climate data for the 13-year period of 1996–2008. To overcome some of the challenges associated with low numbers, such as high error rates, the control and experimental data for taxa were combined before model creation was attempted as there was no statistical difference between the results of the two treatments. A generalized linear model (GLM) with a Poisson distribution and log link function was implemented using the 'bestglm' package and was used as a preliminary filter to identify the most suitable predictors for each predictand [63]. The bestglm function returns the best subset of predictor inputs for GLMs using several selection methods that are available [64]. For this study, the bestglm returned a ranking of the best combination of predictors selected based on their AIC values. The best combination of predictors was then assessed using a GLM with a quasipoisson distribution and a log link function to account for overdispersion and species with inflated zero abundances [65] and because the 'bestglm' function did not allow for the inclusion of the quasipoisson distribution. The pool of predictors for each predictand was narrowed by finding variables that were significant (p -value < 0.05) when they were included (in combination) with other variables. Specifically, if >2 predictors that were identified by 'bestglm' as the best combination were included together in a model, and one

or more was/were not significant, the single non-significant variable was dropped or for >1 non-significant variables, they were dropped in turn (one dropped and the other retained and vice-versa) and the model was then run again. The final model was not accepted until all predictors were significant. If there were two remaining predictors and one of the two was not significant, then the predictor with the highest pseudo R-squared value was used to build the final models. This was also used to select models where two or more predictors were non-significant and when they were dropped in turn all the predictors were significant (resulting in >1 possible final models). The pseudo R-squared was calculated using the following formula:

$$100 \times ((\text{model null.deviance} - \text{model deviance})/\text{model\$null.deviance}).$$

It was used for model selection because the AIC and R-squared cannot be calculated for a GLM with a quasipoisson distribution. Thus, the ideal combination of significant predictors for a predictand was identified as that which gave the higher pseudo R-squared value, and model fit was assessed using the marginal plots and distribution of Pearson's and deviance residuals. This process was carried out for taxa that had both linear and polynomial trends. Predictors were centered where necessary.

The predictors used for each model included all 22 taxa, 20 climate (including seasonal) indices, and seasonal variables, for a total of 42 predictor variables. The climate variables used also lagged the abundance data by one year and, in the case of seasons that approached the trapping period (e.g., DJF), were terminated at the start of said period. Taxa were retained as predictors of other taxa only in cases of known or suspected interaction. Trophic relationships, specifically lizards as predators of invertebrate prey were based on the dietary information for the relevant lizard species [66]. The final model allowed no feedback between predictands (they could not simultaneously predict each other), and negative values were reported <0.

Final equations were obtained by means of a generalized linear mixed model (GLMM) to allow for the inclusion of both fixed effects and random effects. The random effects that were included were the grids (2) nested within the treatments (2)—the abundance values for each year ($\times 13$) were nested within each grid (giving a total of 26 abundance values for each species). The `glmer` function from the 'lme4' package with a poisson distribution and log link function [65] was used to implement the GLMM if there was no over or under dispersion. If there was over or under dispersion or if the `glmer` could not converge for a particular taxon, the `glmmPQL` function from the 'mass' package was used with a quasipoisson distribution and log link function [67].

A GLM was used to identify candidate predictors for the final models because the preferred and more accurate GLMM function (`glmer`) did not always converge, and it cannot include a quasipoisson distribution. Furthermore, the `glmmPQL`, which was more robust, did not return an AIC, hence a GLM was deemed more appropriate for predictor screening. Before the final models were accepted, model fit was assessed using the marginal plots and distribution of Pearson's and deviance residuals. Additionally, the autocorrelation function (ACF) and partial autocorrelation function (PACF) were used to determine whether model residuals were random and not temporally autocorrelated. No model was found to be temporally autocorrelated. The marginal and conditional R-squared, which were calculated using the 'MuMIn' package, were reported for the final models [68]. Also, validation of each model was conducted using two methods—prediction of an independent dataset, and the use of the package 'spTimer' in R. Firstly, each model was validated by predicting an independent subset of data. The abundance dataset was split into two for training and validation. Model training was conducted using the period 1997–2009, and validation conducted for the period 2010–2013. Mean historical abundance was compared graphically to predicted data for that period to evaluate the deviation of the model predictions from recorded data. Secondly, the package 'spTimer' was used to assess a range of mean square error estimations over the modelling period [69].

The ultimate aim was the creation of a simple model for the Hellshire Hills that would take into account all statistically significant relationships (whether climate-species or species-species) but was primarily centered on taxa that required only climate as their predictors. The latter constraint was imposed in order to maintain focus on deducing a mechanism for evaluating the variability and extinction risk of small populations in the face of a changing climate. In this case, then, only future climate data would be required as input to the simple model after each constituent model creation and validation. It is recognized that the simple model proposed has limitations, such as the use of climate and taxon abundance as the only inputs, an increase in experiment-wise error rate in proportion to an increase in the number of statistical tests, and the eventual focus on a subset of the arthropod community of the area. Notwithstanding, it is felt that it is sufficiently indicative to draw some conclusions about the influence of climate on the species of the Hellshire Hills. The final bioclimatic model is termed the Hellshire Hills model or hereafter the HH model.

2.5. Future Projections of Abundance

Using the HH model, projections of climate impacts were generated by inputting climate data consistent with future climate scenarios. End-of-century data from the regional climate model (RCM) PRECIS were used as input for the HH model. The PRECIS RCM was selected because it has already been validated and extensively used to generate projections for Jamaica and the wider Caribbean region [2,7,70]. The model output used in this study included (i) simulated baseline rainfall and temperature data for the period 1961–1999, and (ii) simulated future rainfall and temperature data under the A2 (high emissions) and B2 (medium emissions) SRES scenarios for the period 2071–2099. During the time frame of this study, RCM projections from PRECIS were only available for Jamaica at 50 km resolution. The scale is much larger than that of the region being considered and this is noted as a limitation of the methodology. The projected end-of-century change in climatic variables with respect to the simulated baseline for the grid box situated over the Hellshire Hills was determined and extracted from the model data.

Projected climate data were used as input in the equations obtained through GLMM to generate future absolute abundance of each taxon represented in the model. Mean change in abundance for the end of the century (2071–2100) with respect to baseline abundance (1997–2013) was calculated for both scenarios. The end result was used to form a profile of the changing faunal community under differing climate scenarios, including those in changing climate environments.

3. Results

3.1. Climate of the Hellshire Hills

The Hellshire Hills climatology was determined using data obtained from the weather station installed at the site (Figure 2). The climatology suggests a pattern of mean temperatures that are coolest in the northern hemisphere winter months (minimum in December) and warmest in the summer months (peak in August). The Hellshire Hills exhibit an overall warmer climate than both Norman Manley and the Jamaican average, with wider variation between maximum and minimum temperatures throughout the year. The region is generally hot all year round, with comparatively little monthly variation (Figure 2A). The annual ranges for mean, maximum and minimum temperatures for the Hellshire Hills region were only 3.3, 3.4, and 3.2 °C respectively. Daily temperature range was typically smaller during the warmer summer months than it was during colder periods. The available data also indicated that the maximum mean daytime temperature for the Hellshire Hills varied between 31 and 33 °C, and was consistently high throughout the year. In comparison, minimum night-time temperature varied from a low of 21 °C in December–March to as high as 25 °C in June, while mean temperatures varied between 24 and 28 °C, with a peak in August. The diurnal temperature variation appeared to be greater for Hellshire Hills than Norman Manley for the same period, with generally cooler nights and warmer days.

A maximum summer daytime temperature of 39 °C was recorded in the Hellshire Hills on 31 July 2012, and a minimum winter night-time temperature of 17 °C was recorded on 15 December 2010. In the summer of 2011, 13 days between June and August exceeded 35 °C. This rose to 25 days in the summer of 2012, indicating an increase in the number of hot days.

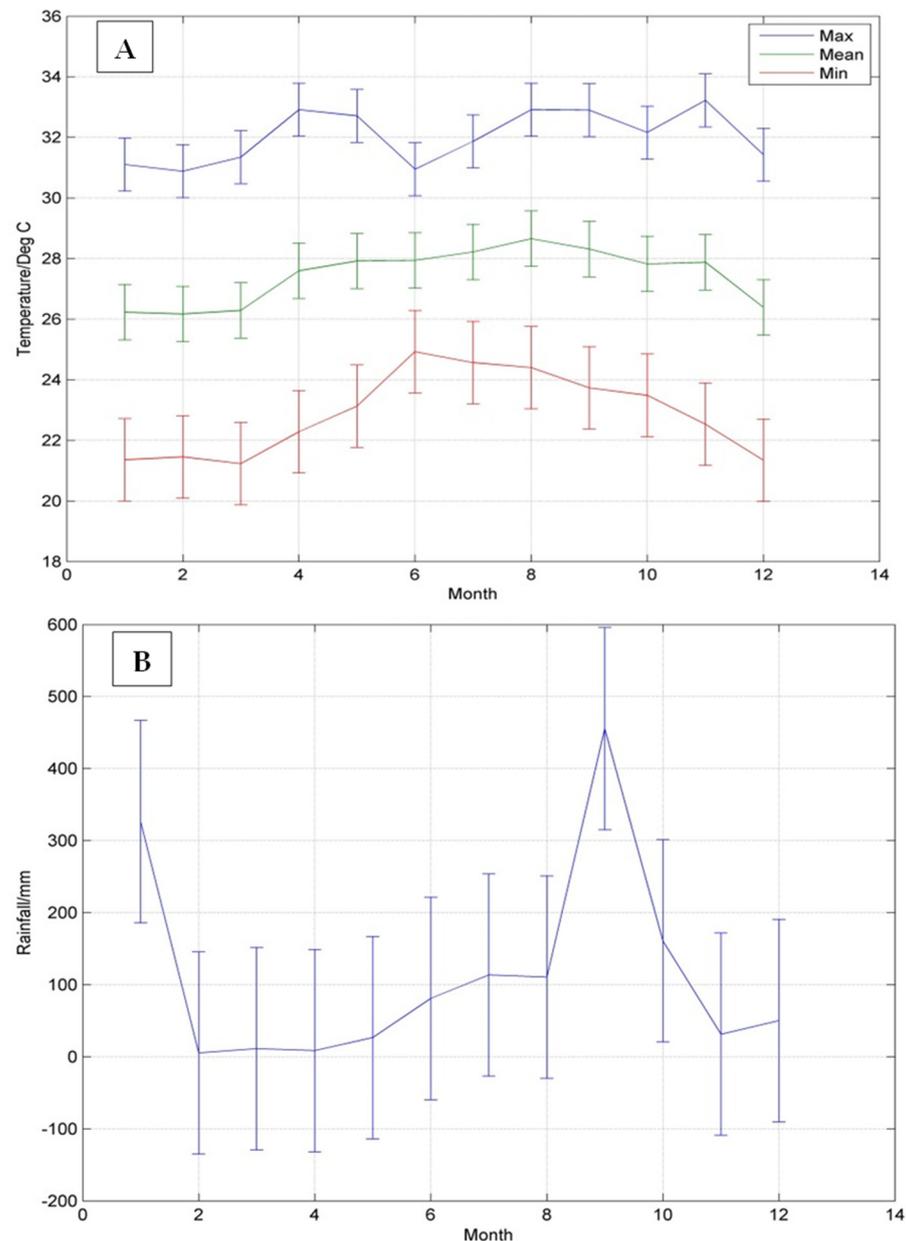


Figure 2. Temperature (A) and rainfall (B) climatologies of the Hellshire Hills (HH) for the period 2010–2012. Maximum temperatures are presented in blue, minimum temperatures in red, and mean temperatures in green. Temperatures represent the monthly averages calculated from the respective datasets, and error bars represent one standard deviation. Rainfall represents the monthly averages calculated from the respective datasets, and error bars represent one standard deviation (Norman Manley and Jamaica Data Source: Meteorological Service of Jamaica).

The site is by comparison very dry when compared with rainfall received at Norman Manley or the island average. Year by year analysis of the HH station data suggested a general mirroring of Jamaica’s bimodal rainfall pattern, i.e., with a rainy season between May and November interrupted by a short dry spell in August. However, the peak in May

was not distinct in all three years of data, resulting in the derived climatology displaying a more unimodal distribution over the rainfall season (Figure 2B). The early months (February to March) were also particularly dry, with almost no rainfall registered during the period (Figure 2B). Typically, January–February represents the peak of the Hellshire Hills dry season. Figure 2B also shows that nine months of the year received approximately 100 mm or less of rainfall. However, we note the influence of the drought of 2010, which may have skewed the data, and the influence of Tropical Storm Nicole and Hurricane Sandy in 2010 and 2012, respectively. Notwithstanding, the general pattern that emerges is of a region with very little rainfall except for a few months of the year.

Projections from the PRECIS RCM for the grid box over the Hellshire Hills indicate that the region is projected to warm by between 2.8 °C (B2) and 4.3 °C (A2) (Table 2). Warming is most pronounced during the summer months (June–July) under both scenarios, reaching as high as 5.2 °C under the A2 scenario and 3.0 °C under the B2 scenario. The traditionally cooler months between November and February show smaller magnitude warming (2–3 °C). Model projections also suggest an increase in the frequency of very warm days and a corresponding decrease in the frequency of very cool days irrespective of scenario. Projected percentage changes in rainfall indicate between 23% and 40% less rainfall in the annual mean occurring in the Hellshire Hills under the B2 and A2 scenarios (respectively) by the end of the century. That is, an already significantly dry region will become even drier. Drying is most significant in the summer months under both scenarios.

Table 2. Projections of mean percentage changes (%) in monthly rainfall and mean absolute changes in monthly temperature (°C) under A2 (high emissions) and B2 (low emissions) scenarios for the period 2071–2099.

Month	Rainfall		Temperature	
	A2	B2	A2	B2
January	−39.1	−6.6	3.4	2.3
February	−18.7	−0.8	4.2	2.5
March	16.8	−33.8	4.3	3.6
April	10.9	8.7	3.5	2.8
May	−52.4	−12.4	4.5	3.0
June	−65.7	−30.0	5.2	3.1
July	−57.4	−31.0	5.2	3.1
August	−51.8	−30.7	4.9	3.0
September	−61.5	−43.3	5.0	3.1
October	−67.7	−47.3	4.7	2.8
November	−43.0	−18.5	3.6	2.4
December	−53.9	−28.7	3.2	2.4

In general, present-day seasonality will still be retained by the end of the century, i.e., the summer months will still be the warmest and wettest periods in the far future, while February through March will remain the driest and coolest months. However, there will be an increase in annual temperature range of 2.5–3 °C along with the possibility that under the more severe emissions scenario rain in January may be comparable to peak rain received in September due to the drastic reduction in the latter month's totals. In tandem the temperature and rainfall projections present an end of century profile which suggests that the Hellshire Hills will become even warmer and drier. Importantly, the changes projected are larger than the range of variability currently seen, as revealed by the trends in the 17-year present-day dataset. This suggests 'unprecedented' changes to the climatic envelope of the Hellshire Hills.

3.2. Variation in Abundance and Climate Linkages

A significant linear trend in abundance was not identified for any taxon over the 17-year period under investigation. Notwithstanding, for 14 of the 22 species, the trend

in abundance over time is slightly negative. The taxa with the largest magnitude slope (whether positive or negative) were stink bugs (*Pangaeus bilineatus* +6.19) and beetles (*Branchus jamaicensis* −5.05, Chrysomelidae −75.01). This is consistent with previously identified significant negative slopes for temperature variables and positive slopes for rainfall [71]. Interannual variability is also evident in almost all the species. This is very reminiscent of the rainfall trends (annual and seasonal) for which periodicity analysis revealed a dominant three-year signal that may also be linked to the variability evident in annual abundance. This suggests that rainfall is a significant influencing factor [71].

The following observations are therefore noted from abundance data in both control and experimental plots: (i) there is no significant linear trend, (ii) there is strong interannual variability, and (iii) there are common linkages with rainfall across numerous taxa, in particular for lizards.

All trap areas showed similar trends in overall captures throughout the study period, and experimental plots had consistently higher capture rates than control plots. Correlations were generally high and significant (i) between control grids; (ii) between experimental grids, and (iii) between the total captures for the control and experimental plots combined. This suggests that, in general, the factors (e.g., climate) that are driving variability of the trapping at one control or experimental plot are common to the other control or experimental plot.

3.3. Model Creation

Final taxon models were obtained for 20 of the 22 taxa (Figure 3; Table 3). Significant models were not obtained for springtails (Collembola) and Jamaican blue-tailed galliwasps (*Celestus duquesneyi*). Three taxa, namely crickets and silverfish (*Ceuthophilus* sp., Gryllidae, Lepismatidae), were found to have non-linear (polynomial) relationships with their predictors, but all others were found to be linear. It is noted that 16 of 20 models retained only one predictor. The models had exclusively either only climate or another species as predictors, except for leaf beetles (Chrysomelidae), which had both Jamaican gray anoles (*Anolis lineatopus*) and a climate variable (rainfall during the mid-summer dry spell (JA) and dry season (DJF)) as predictors (Figure 3; Table 3). The predominant predictors were climate indices, more so rainfall than temperature. Thirteen of the 20 models included a climate variable as a predictor, with 12 having only climate predictors (Figure 3; Table 3). The rainfall predictors retained by the models were total annual rainfall and seasonal rainfall (Figure 3; Table 3). Among the seasonal rainfall variables retained, the dry season (DJF) was most frequently identified as a significant predictor, and the late wet season (ASON) identified least frequently (Figure 3; Table 3). Three of the four lizard taxa for which relationships were obtained were strongly related to rainfall, particularly total annual rainfall and dry season rainfall (DJF) (Figure 3; Table 3).

Extreme temperature indices emerged as the temperature predictors retained, i.e., as opposed to mean temperatures (Figure 3; Table 3). The percentage of days when maximum or minimum temperature exceeded the 90th percentile (TX90P and TN90P) explained five of the six temperature related taxa (Table 3). Diurnal temperature range (DTR) was the other temperature variable retained for one taxon, centipedes (Chilopoda).

Seven taxa were predicted solely by abundance of another species (Figure 3; Table 3). In all cases, the species predictors were themselves all predicted solely by a climate variable, suggesting an indirect climate influence. Predator-prey association was apparent in the eight taxa that had abundance of other species as predictors. The largest explained variance among all models occurred in velvet ants (*Dasymutilla militaris*) ($mR^2 = 59\%$, $cR^2 = 68.6\%$) predicted by TX90P and rainfall during JA. Lowest explained variance for any model ($mR^2 = 8.1\%$, $cR^2 = 8.1\%$) was for stink bugs (*Pangaeus* sp.) predicted by TX90P. The three polynomial models yielded among the highest explained variances of all models with cR^2 ranging from 47.7% for crickets (Gryllidae) to 65.5% silverfish (Lepismatidae).

It is noted that the influence of temperature was most often positive. In cases of apparent predator-prey relationships, prey species were positively influenced by their

predictors up to a point, after which a further increase in predator abundance became a negative influence, and predators increased in abundance with an increase in their prey.

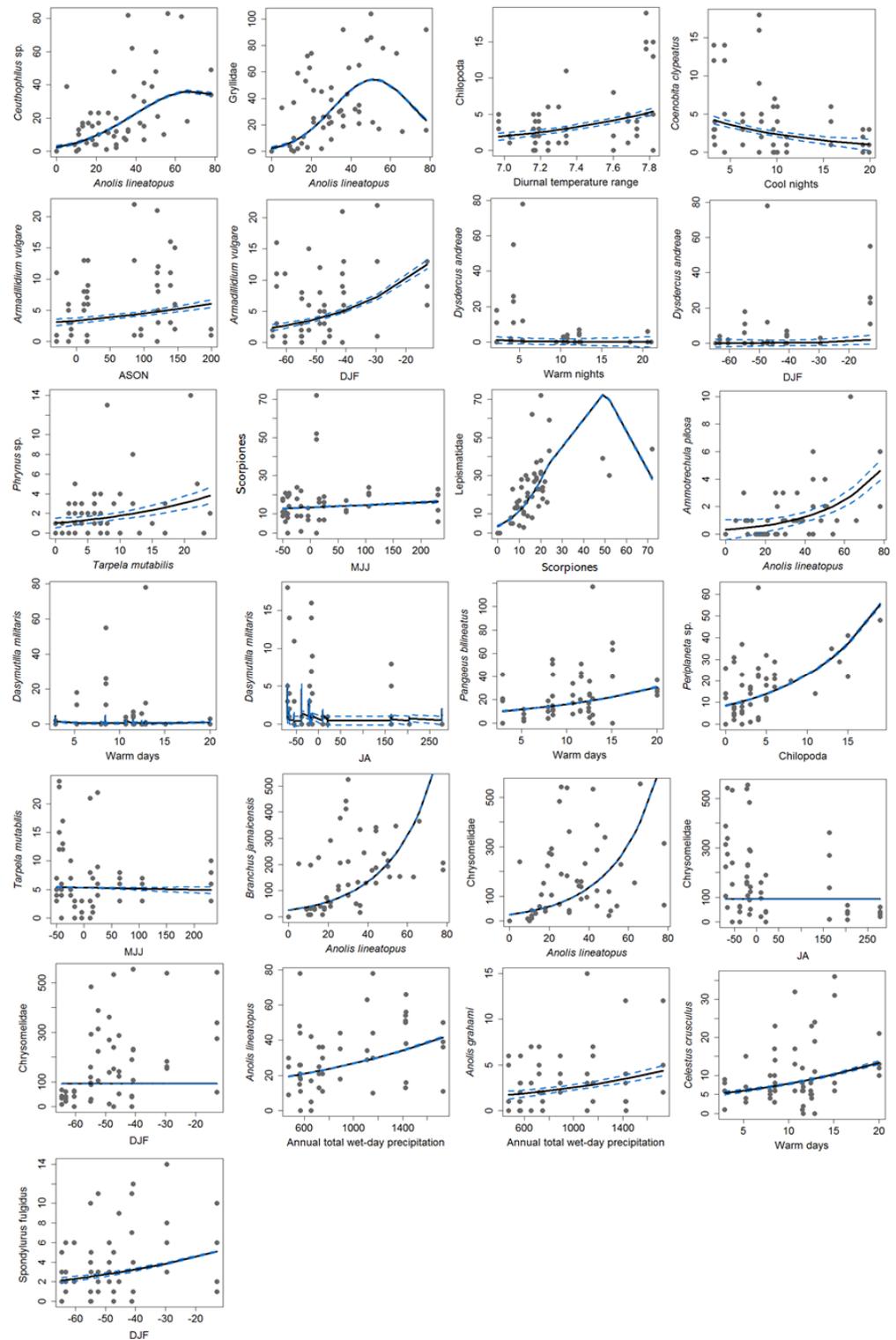


Figure 3. Observed (dots) and fitted (solid black line) values of significant models for all modelled taxa. On each graph, abundance in number of individuals is presented on the *y*-axis and one predictor on the *x*-axis. For predictors that are also species, *x*-axis represents abundance in number of individuals. For climate predictors, descriptions and units can be found in Table A1. Blue dashed lines indicate 95% confidence intervals.

Table 3. Results of the final generalized linear mixed models (GLMMs) for each taxon, where C—data centered; poly—polynomial GLMM; SE—standard error; Z—Z distribution for GLMM; Pr(>|z|)—Probability; mR²—marginal R-squared; and cR²—conditional R-squared. Continued on next page.

Taxa	Parameters	Estimate	SE	Z	Pr(> z)	mR ²	cR ²
<i>Ceuthophilus</i> sp.	(Intercept)	0.903	0.454	1.988	0.047	39.4	51.9
	poly(al, degree = 2)1	0.078	0.023	3.485	<0.001		
	poly(al, degree = 2)2	−0.001	0.000	−2.047	0.041		
Gryllidae	(Intercept)	0.816	0.508	1.605	0.108	43.0	47.7
	poly(al, degree = 2)1	0.124	0.028	4.457	<0.001		
	poly(al, degree = 2)2	−0.001	0.000	−3.496	<0.001		
Chilopoda	(Intercept)	−7.913	3.026	−2.615	0.009	14.9	17.7
	dtr	1.227	0.407	3.011	0.003		
<i>Coenobita clypeatus</i>	(Intercept)	1.718	0.333	5.154	<0.001	15.0	15.0
	tn10p	−0.086	0.031	−2.748	0.006		
<i>Armadillidium vulgare</i>	(Intercept)	2.693	0.455	5.913	<0.001	24.3	26.1
	ASON	0.005	0.002	2.508	0.012		
	DJF	0.036	0.010	3.650	<0.001		
<i>Dysdercus andreae</i>	(Intercept)	4.272	1.665	2.566	0.010	28.2	28.2
	tn90p	−0.242	0.098	−2.473	0.013		
	DJF	0.076	0.035	2.167	0.030		
<i>Phrynus</i> sp.	(Intercept)	0.018	0.255	0.070	0.944	9.3	9.3
	1bb	0.055	0.024	2.294	0.022		
Scorpiones	(Intercept)	2.650	0.090	29.451	<0.001	6.4	6.4
	MJJ	−0.002	0.001	−1.817	0.069		
Lepismatidae	(Intercept)	1.284	0.257	4.994	<0.001	58.7	65.5
	poly(scorp, degree = 2)1	0.130	0.019	6.891	<0.001		
	poly(scorp, degree = 2)2	−0.001	0.000	−5.270	<0.001		
<i>Ammotrechula pilosa</i>	(Intercept)	−1.147	0.376	−3.047	0.002	26.1	26.1
	al	0.034	0.008	4.139	<0.001		
<i>Dasymutilla militaris</i>	(Intercept)	3.172	0.757	4.191	<0.001	59.0	68.6
	tx90p	−0.286	0.061	−4.681	<0.001		
	JA	−0.015	0.003	−5.055	<0.001		
<i>Pangaeus bilineatus</i>	(Intercept)	2.129	0.362	5.886	<0.001	8.1	8.1
	tx90p	0.065	0.031	2.085	0.037		
<i>Periplaneta</i> sp.	(Intercept)	2.146	0.167	12.830	<0.001	21.0	21.0
	cent	0.098	0.026	3.710	<0.001		
<i>Tarpela mutabilis</i>	(Intercept)	1.758	0.110	15.938	<0.001	32.1	32.1
	MJJ	−0.007	0.002	−4.044	<0.001		
<i>Branchus jamaicensis</i>	(Intercept)	3.226	0.303	10.663	<0.001	34.2	34.2
	al	0.042	0.008	5.043	<0.001		
Chrysomelidae	(Intercept)	4.397	0.633	6.948	<0.001	48.9	53.4
	al	0.044	0.008	5.440	<0.001		
	JA	−0.005	0.002	−3.226	0.001		
	DJF	0.024	0.011	2.128	0.033		
<i>Anolis lineatopus</i>	(Intercept)	2.677	0.283	9.473	<0.001	11.3	11.3
	prcptot	0.001	0.000	2.074	0.038		
<i>Anolis grahami</i>	(Intercept)	0.175	0.360	0.486	0.627	9.1	9.1
	prcptot	0.001	0.000	2.244	0.025		

Table 3. Cont.

Taxa	Parameters	Estimate	SE	Z	Pr(> z)	mR ²	cR ²
<i>Celestus cruscus</i>	(Intercept)	1.530	0.288	5.320	<0.001	8.5	8.5
	tx90p	0.053	0.024	2.170	0.030		
<i>Spondylurus fulgidus</i>	(Intercept)	1.846	0.002	825.700	<0.001	6.3	12.1
	DJF	0.017	0.002	8.400	<0.001		

3.4. The HH Model

Using the relationships identified from the individual models, a simple schematic diagram compiling all taxon models for the Hellshire Hills (the HH model) was created (Figure 4). In the figure, the predictor-predictand relationships of the individual models are represented by the arrow directions, i.e., from predictors to predictand. It is not, however, implied that the relationship suggested by each arrow explains the same amount of variability.

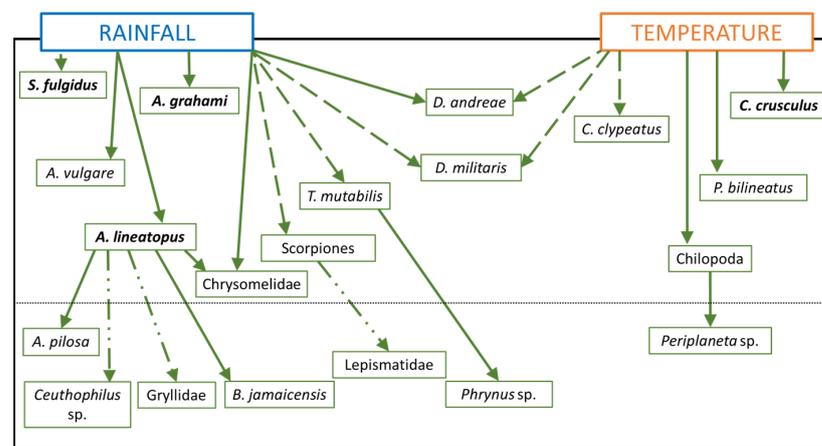


Figure 4. Simple Hellshire Hills (HH) model schematic diagram showing lizard (bold names) and arthropod models in the Hellshire Hills ecosystem. Arrows indicate connections among model predictors (start of arrow) and predictands (end of arrow). Solid arrows represent positive relationships, broken arrows represent negative relationships, and long dash-dot-dot arrows represent polynomial relationships. Horizontal dotted black line separates taxa that are predicted by at least one climate variable (above) from those that are only predicted by other taxa (below). Diagram does not imply order of prediction.

The HH model is simple, comprehensive, and climate-based. Although it captures climate-species and species-species relationships, it is guided solely by statistical relationships and does not capture any feedback mechanisms. The HH model has only climate variables (rainfall and temperature) as inputs, which are then used to estimate taxon abundance. It is this latter property of the HH model that facilitates a determination of how changes in climate may potentially influence the abundance of the species present in the Hellshire Hills.

3.5. Model Validation

The models varied in performance. However, they generally capture magnitudes and patterns. Magnitudes and trends in abundance during the independent period were also captured reasonably by most models. However, these predictions tended to be more accurate in the last two to three years (2011–2013) than in the initial year (2010). Six models showed little to no predicted trend during the independent period, namely crickets, centipedes, hermit crabs, red cotton stainers, boxy-whip scorpions, and cockroaches (*Ceuthophilus* sp., Chilopoda, *Coenobita clypeatus*, *Dysdercus andreae*, *Phrynus* sp.,

and *Periplaneta* sp.), despite trends being evident in observed data. Population size was not necessarily a hindrance to prediction, as is evident in the accuracy of predictions for solpugids (*Ammotrechula pilosa*) and Jamaican skinks (*Spondylurus fulgidus*) during the independent period.

Mean squared error, root mean squared error, and mean absolute error (see Table 4) were lowest in models for which trends seemed to be best captured. RMSE estimates for these models all fell below 2.0. These were followed by eight models that had RMSE values below 10.0. High error estimates for remaining models, particularly beetles (*Branchus jamaicensis* and Chrysomelidae), indicate divergence between observed and predicted abundance.

Table 4. Error estimates for each model. MSE = mean squared error, RMSE = root mean squared error, and MAE = mean absolute error.

Taxon	MSE	RMSE	MAE
<i>Ceuthophilus</i> sp.	221.62	14.89	13.31
Gryllidae	341.46	18.48	15.46
Chilopoda	7.00	2.65	1.77
<i>Coenobita clypeatus</i>	8.62	2.94	2.00
<i>Armadillidium vulgare</i>	12.08	3.48	2.54
<i>Dysdercus andreae</i>	69.69	8.35	4.31
<i>Phrynus</i> sp.	3.62	1.90	1.00
Scorpiones	63.54	7.97	4.31
Lepismatidae	874.69	29.58	29.46
<i>Ammotrechula pilosa</i>	1.92	1.39	1.00
<i>Dasymutilla militaris</i>	2.77	1.66	1.23
<i>Pangaeus bilineatus</i>	160.54	12.67	9.62
<i>Periplaneta</i> sp.	76.46	8.74	7.23
<i>Tarpela mutabilis</i>	16.38	4.05	2.69
<i>Branchus jamaicensis</i>	36,259.77	190.42	119.92
Chrysomelidae	20,542.15	143.33	104.31
<i>Anolis lineatopus</i>	273.69	16.54	11.38
<i>Anolis grahami</i>	2.38	1.54	1.15
<i>Celestus cruscus</i>	21.62	4.65	3.46
<i>Spondylurus fulgidus</i>	2.69	1.64	1.31

3.6. Model Projections

Projected mean change factors estimated for each taxon under each scenario for the end of the century varied (Table 5), i.e., they indicated a decrease or no change in many taxa, but also an increase in others, with some clear differences dependent on the scenario considered.

Under the A2 scenario, eight taxa are projected to decrease, including three of the five lizards assessed. These groups are pill bugs, red cotton stainers, solpugids, beetles, gray anoles, turquoise anoles, and skinks (*Armadillidium vulgare*, *Dysdercus andreae*, *Ammotrechula pilosa*, *Branchus jamaicensis*, Chrysomelidae, *Anolis lineatopus*, *Anolis grahami*, and *Spondylurus fulgidus*). It is notable that this group consists entirely of species predicted by total or seasonal rainfall, or species predicted by other species that were in turn predicted by rainfall. Four taxa are projected to experience no discernible change in abundance, three of which were predicted by temperature indices. Eight taxa were projected to increase in abundance. These groups were crickets, scorpions, silverfish, velvet ants, stink bugs, beetles, and common galliwasps (*Ceuthophilus* sp., Gryllidae, Scorpiones, Lepismatidae, *Dasymutilla militaris*, *Pangaeus bilineatus*, *Tarpela mutabilis*, and *Celestus cruscus*). The magnitudes of the changes are, however, on a much smaller scale than the losses estimated for the eight taxa that decreased. The predictors also represent a ‘mixed bag’ including rainfall, temperature and other species. Of most interest was the fact that two of the taxa that increased (cricket taxa *Ceuthophilus* sp. and Gryllidae) are predicted by gray anoles

(*Anolis lineatopus*), for which abundance decreased under the significant drying. The latter is known to be a predator of the former.

Table 5. Projected mean absolute changes in abundance towards end of century under A2 (high emissions) and B2 (low emissions) scenarios.

Scenario	A2	B2
<i>Ceuthophilus</i> sp.	16	<0
Gryllidae	30	<0
Chilopoda	0	<0
<i>Coenobita clypeatus</i>	0	<0
Collembola	-	-
<i>Armadillidium vulgare</i>	<0	10
<i>Dysdercus andreae</i>	<0	129
<i>Phrynus</i> sp.	0	<0
Scorpiones	1	6
Lepismatidae	2	46
<i>Ammotrechula pilosa</i>	<0	<0
<i>Dasymutilla militaris</i>	22	<0
<i>Pangaeus bilineatus</i>	5	<0
<i>Periplaneta</i> sp.	0	3
<i>Tarpela mutabilis</i>	1	<0
<i>Branchus jamaicensis</i>	<0	398
Chrysomelidae	<0	2007
<i>Anolis lineatopus</i>	<0	56
<i>Anolis grahami</i>	<0	5
<i>Celestus cruscus</i>	1	8
<i>Celestus duquesneyi</i>	-	-
<i>Spondylurus fulgidus</i>	<0	6

Under the B2 scenario, an increase was projected for 11 taxa, including all lizards. These were pill bugs, red cotton stainers, scorpions, silverfish, cockroaches, leaf beetles, gray anoles, turquoise anoles, common galliwasp, and skinks (*Armadillidium vulgare*, *Dysdercus andreae*, Scorpiones, Lepismatidae, *Periplaneta* sp., *Branchus jamaicensis*, Chrysomelidae, *Anolis lineatopus*, *Anolis grahami*, *Celestus cruscus*, and *Spondylurus fulgidus*). Interestingly, all but two of these 11 taxa are predicted by rainfall directly or indirectly. The suggestion is that the taxa modelled may be sensitive to the magnitude of change, particularly with respect to rainfall. Decreased abundance was projected for nine arthropods and no lizards. These taxa were crickets, centipedes, hermit crabs, boxy whip scorpions, solpugids, velvet ants, stink bugs, and beetles (*Ceuthophilus* sp., Gryllidae, Chilopoda, *Coenobita clypeatus*, *Phrynus* sp., *Ammotrechula pilosa*, *Dasymutilla militaris*, *Pangaeus bilineatus* and *Tarpela mutabilis*).

In comparing the projections for the two scenarios, eight taxa shifted from a pattern of increase or no change under A2 to one of decrease under B2. The reverse was noted in seven. For taxa that increased under both scenarios, that change was larger under the B2 scenario.

A2 and B2 generally were both dry scenarios, with A2 indicating severe drying. There appears to be sensitivity to the magnitude of drying, particularly among lizard taxa and those taxa predicted by the lizards. The effect of increased temperatures is not entirely evident from the models created.

4. Discussion

The assessment of the historical and future climate of the Hellshire Hills indicates a warming and drying dry forest. Results of this study support the idea of climate sensitivity among the fauna of the site and provide indications of how that sensitivity is being manifested. For example, (i) some of the strongest correlations were with seasonal totals for rainfall and derived temperature indices rather than for annual totals; (ii) in general,

covariation tended to be stronger with rainfall (especially for the lizard species), and (iii) not all taxa exhibited a direct climate-species link but indirect climate linkages were suggested, i.e., even though some species did not exhibit a strong correlation with climate, they were significantly correlated with another species that did. The simple HH model offers insight into the equilibrium among species as aspects of their habitat change over time.

The models produced predominantly used rainfall variables as predictors, but also included temperature predictors for a number of taxa. Dry season rainfall was identified as the predictor of Jamaican skinks (*Spondylurus fulgidus*, formerly *Mabuya mabouya*). This can feasibly be linked to the strong wet-dry seasonality of skink reproduction, particularly given that birth of young and peak testis size have been known to occur in at least three other species of tropical skinks (*Mabuya frenata*, *M. heathi* and *M. nigropunctata*) in the late dry season or early wet season [72]. Number of warm days was identified as a predictor of common galliwasps (*Celestus cruscus*), indicating that warmer ambient temperatures are preferable for maintaining optimal body temperature as has been seen in other actively thermoregulating anguids [73].

Five arthropod taxa were predicted by gray anoles (*Anolis lineatopus*). These were of the orders Solpugida (solpugids), Orthoptera (crickets), and Coleoptera (beetles). Whereas crickets and beetles are known prey choices of regional anoles [22,74,75], solpugids may be linked to anoles as prey or competition, as has been seen in previous studies of interactions between anoles and arachnids [76,77]. Solpugids have been found to be consumed by lizards such as side-blotched lizards and geckos [78,79] and are also known to prey on lizards and larger prey [80], but no studies directly showing interaction between solpugids and West Indian anoles were found during this study. Two additional likely predator-prey interactions were noted, in which boxy-whip scorpions (*Phrynus* sp.) was predicted by beetles (*Tarpela mutabilis*) and scorpions (Scorpiones) predicted silverfish (Lepismatidae). Though these interactions have not been directly studied in the region, *Phrynus* sp. and Scorpiones have been found to opportunistically prey on a wide range of arthropods (and vertebrates), particularly under food stress [76,80–83]. In both instances, the predictor was one that was modulated by rainfall.

While temperature is a contributing factor, seasonal rainfall and soil moisture have been found in previous studies to be particularly critical to the survival of pill bugs (*Armadillidium vulgare*) [84,85]. Prior studies assessing *A. vulgare* have noted desiccation during periods of drought due to reduced moisture in leaf litter, and strong linkages between population density and summer rainfall of the previous year [84].

Combinations of temperature and rainfall predictors were identified for red cotton stainers and velvet ants (*Dysdercus andreae* and *Dasymutilla militaris*). The red cotton stainer, *Dysdercus andreae*, was predicted by a negative association with number of warm nights and a positive association with dry season rainfall. However, higher rainfall has been linked to lower abundance in *D. andreae* [86] and higher temperature to greater growth and reproduction in a similar species, *D. fasciatus* [87]. It should be noted, therefore, that the climate influences found by the model were accurate, but in the opposite direction of those previously identified. The velvet ant *Dasymutilla militaris* was predicted by rainfall during the midsummer dry spell (JA) and number of warm days, both inverse linkages. This temperature relationship is expected in species of Mutillidae, as they are most active during the early morning and late afternoon when temperatures are cooler, and least active during warmer hours, indicating that warming would be detrimental to feeding and reproduction and may impact patterns of behavior [88]. Similarly, it has been argued that dry conditions are more favorable to velvet ants, though this has not been proven [89].

Four taxa were predicted solely by temperature variables. Diurnal temperature range was identified as a predictor of the centipedes (Chilopoda), which thereby predicted a known prey species of the group, cockroaches (*Periplaneta* sp.) [90,91]. Though centipedes in some parts of the Caribbean region have been relatively well identified [92,93], the interactions of centipedes with their environments are not generally well known. However, in a study of preferences of 12 species of Chilopoda in a river valley that varied in

habitat types, xerothermic conditions were most frequently chosen over wetter and cooler habitat types [94]. Given the highly significant importance of air and soil temperatures to Chilopoda [95], it follows that diurnal temperature range is a reasonable influence of abundance. In very cold conditions, it is typical of the burrowing stinkbug (*Pangaeus bilineatus*) to occupy deeper layers of soil in suspended development [96]. The increase in temperature associated with the transition from winter towards summer months, however, results in individuals increasingly occupying soil closer to the surface and becoming reproductively viable [96], indicating that an increase in the number of warm days as seasons change has strong influence on activity of *P. bilineatus*. Assessments of a similar Cydnidae species, *Cyrtomenus bergi*, found temperature to be a significant limiting factor to development [97]. In the study, a temperature of 26 °C was found to be optimal for growth, and immature developmental stages were more resilient under extreme temperatures, provided that temperatures did not exceed 31 °C [97]. Haemolymph composition and alkalinity of the terrestrial hermit crab (*Coenobita clypeatus*) is temperature regulated [98]. A decrease in temperature is associated with an increase in alkalinity and decrease in the partial pressure of carbon dioxide of haemolymph, and also results in reduced activity, as is characteristic of ectotherms [98], indicating that an increase in the number of cool nights can feasibly have a negative effect on hermit crab abundance as the model suggests. Larval development of tropical *Coenobita* sp. may also be affected by changes in temperature [99].

Bioclimatic modelling has been used minimally in the Caribbean to investigate climate change impacts such as vegetative distribution in Trinidad [20], geographic range shifts of orchids and vectors of parasites in Colombia [100,101], and climate suitability for coffee in Puerto Rico [102]. The results of this study agree with previous studies, showing that tropical organisms are vulnerable to climatic shifts, will vary in their responses depending on taxon specific tolerance, and are likely to decrease in abundance or distribution due to climate change. Notwithstanding their utility, bioclimatic modelling has not widely been undertaken for small scales such as insular tropical dry forest ecosystems, but is more frequently implemented for continental or large island scales. Accordingly, the application of modelling techniques to smaller systems such as islands and small forested areas, which are often centres of endemism and thus high conservation priorities, is urgently needed [20].

The species models' individual abilities reflected varied performances using the metrics of validation, i.e., RSME, patterns of prediction for the training and independent period, and explained variability. The validation process suggested that the models generally captured magnitudes and patterns, particularly for the training period, with notable peaks being represented in most cases. Magnitudes and trends in abundance during the independent period tended, however, to be less reasonably captured by most models. Where an indirect climate link was being captured by a model, the potential for divergence from observed values increased due to the propagation of errors. Nonetheless, the presence of some skill for each model suggested that the simple HH model can be used for the indicative purpose of determining the influence of future climate on faunal abundance in the Hellshire Hills and other dry forests on the small islands of the Caribbean region.

When future climate scenarios were used in the simple HH model, changes in end of century faunal abundance were strongly dependent on the future climate scenario. Given that many of the relationships identified were closely linked to seasonal rainfall totals, it seems that there is a sensitivity in both arthropods and lizards to the magnitude of change in rainfall, which was different (particularly for the dry season) under the two scenarios.

The implications of these results are not only evident for the Hellshire Hills, but can be extrapolated to other small dry forests in the region. Firstly, there is an indisputable link between Caribbean dry forest fauna and climatic change, whether through direct modulation by rainfall and temperature, or through indirect effects among predators and prey. These linkages are expected to be non-linear, and driven by patterns of change rather than magnitude. Secondly, studies on these climatic influences must be specific to these forests and the taxa that occupy them. In providing a simple methodology, the HH model illustrates that this is entirely achievable once there are ecological and climatological datasets

available. It allows for expansion of the pool of climate predictors used to determine with even greater clarity the exact nature of the climate influence in any area. It also emphasizes the value of existing ecological and climatological datasets across the insular Caribbean, and the necessity of improving our understanding of terrestrial and marine ecosystems through ongoing field assessments. Thirdly, climate conservation will become critical to small islands as climate change continues to progress. Evaluation of extinction risks due to climate change will be a necessary component of any ecological conservation program in the Caribbean, as will climate change mitigation and adaptation measures.

This study has demonstrated that, even where they are well preserved, dry forest ecosystems are not immune to the influence of climate change. There is, therefore, a clear need for mainstreaming biodiversity monitoring and implementing local targeted climate mitigation and adaptation measures. The creation and enforcement of protected areas and conservation zones become even more relevant for vulnerable areas like the Hellshire Hills [44]. Additionally, there is a need for greater research into dry forest carbon storage capacity and ecosystem services, as well as identification of appropriate management tools and strategies based on unique characteristics of particular dry forests [19].

To improve the techniques developed here, knowledge gaps must be filled and data acquired for both relatively undisturbed and degraded habitats. This will facilitate an evaluation of the future climate risks to the fauna of these ecosystems, and will require the collaboration of environmental organisations and climate scientists across the region.

5. Conclusions

This study has shown that a simple model can be used to assess climate impacts on fauna in the Hellshire Hills and other Caribbean dry forests despite the small scale relative to areas assessed in previous bioclimatic studies. This allows for estimation of conservation needs in these climate sensitive ecosystems.

Author Contributions: Conceptualization, K.S., B.W. and M.T.; Methodology, K.S., B.W., M.T. and K.M.; Software, K.S., K.M. and J.C.; Validation, K.S.; Formal Analysis, K.S.; Investigation, K.S., B.W., M.T., K.M., R.v.V., J.K. and J.C.; Writing—Original Draft Preparation, K.S.; Writing—Review & Editing, K.S., B.W., M.T. and K.M.; Visualization, K.S.; Supervision, B.W., M.T. and K.M.; Funding Acquisition, B.W. and M.T. All authors have read and agreed to the published version of the manuscript.

Funding: Funders include the International Iguana Foundation, Conservation International, Disney Wildlife Conservation Fund, Dutch Iguana Foundation, Mohamed bin Zayed Conservation Fund, Global Environmental Facility, National Environment and Planning Agency, Centre for Agriculture and Bioscience International, Government of Jamaica, Beneficia Foundation, Environmental Foundation of Jamaica, Woodland Park Zoo Conservation Fund, and several anonymous donors. Funding was also received from The University of the West Indies through the Research and Publications Grant, Principal's New Initiative Fund, and Mona Campus Research Fellowship Programme. Additional support was received from a number of zoos including the Hope Zoo, Fort Worth Zoo, San Diego Zoo, Audubon Zoo, and Miami Metrozoo.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Restrictions apply to the availability of the ecological data, and requests for access must be directed to Professor Byron Wilson. Climate projection data are publicly available datasets and can be accessed at <http://clearinghouse.caribbeanclimate.bz/> accessed on 18 January 2022. Norman Manley International Airport weather data were provided by the Meteorological Office of Jamaica and can be accessed through request.

Acknowledgments: This study was conducted through the Jamaican Iguana Recovery Group and Climate Studies Group, Mona, and was supported by The University of the West Indies, Mona Campus.

Conflicts of Interest: The authors declare no conflict of interest.

Appendix A

Table A1. Climate predictors used in model creation.

	Variable	Definition	Unit
prcptot	Annual total wet-day precipitation	Annual total PRCP in wet days ($RR \geq 1$ mm)	mm
cdd	Consecutive dry days	Maximum number of consecutive days with $RR < 1$ mm	Days
cwd	Consecutive wet days	Maximum number of consecutive days with $RR \geq 1$ mm	Days
R10	Number of heavy precipitation days	Annual count of days when $PRCP \geq 10$ mm	Days
R20	Number of very heavy precipitation days	Annual count of days when $PRCP \geq 20$ mm	Days
r95p	Very wet days	Annual total PRCP when $RR > 95$ th percentile	mm
r99p	Extremely wet days	Annual total PRCP when $RR > 99$ th percentile	mm
sdi	Simple daily intensity index	Annual total precipitation divided by the number of wet days (defined as $PRCP \geq 1.0$ mm) in the year	mm/day
dtr	Diurnal temperature range	Monthly mean difference between TX and TN	°C
tmax	Maximum Tmax	Monthly maximum value of daily maximum temp	°C
tmin	Minimum Tmin	Monthly minimum value of daily minimum temp	°C
tmean	Mean Temperature	Monthly mean value of daily mean temp	°C
tn10p	Cool nights	Percentage of days when TN < 10 th percentile	Days
tn90p	Warm nights	Percentage of days when TN > 90 th percentile	Days
tx10p	Cool days	Percentage of days when TX < 10 th percentile	Days
tx90p	Warm days	Percentage of days when TX > 90 th percentile	Days
MJJ		Annual rainfall anomalies for the season May–June–July	mm
JA		Annual rainfall anomalies for the season July–August (mid-summer drought)	mm
SON		Annual rainfall anomalies for the season September–October–November	mm
DJF		Annual rainfall anomalies for the season December–January–February	mm

References

1. Pulwarty, R.S.; Nurse, L.A.; Trotz, U.O. Caribbean islands in a changing climate. *Environment* **2010**, *52*, 16–27. [\[CrossRef\]](#)
2. Campbell, J.D.; Taylor, M.A.; Bezanilla-Morlot, A.; Stephenson, T.S.; Centella-Artola, A.; Clarke, L.A.; Stephenson, K.A. Generating Projections for the Caribbean at 1.5, 2.0, and 2.5 °C from a High-Resolution Ensemble. *Atmosphere* **2021**, *12*, 328. [\[CrossRef\]](#)
3. Climate Studies Group Mona. *The State of the Caribbean Climate*; Produced for the Caribbean Development Bank; The University of the West Indies: Kingston, Jamaica, 2020.
4. Stephenson, T.S.; Vincent, L.A.; Allen, T.; Van Meerbeeck, C.J.; McLean, N.; Peterson, T.C.; Taylor, M.A.; Aaron-Morrison, A.P.; Auguste, T.; Bernard, D.; et al. Changes in extreme temperature and precipitation in the Caribbean region, 1961–2010. *Int. J. Climatol.* **2014**, *34*, 2957–2971. [\[CrossRef\]](#)
5. Torres, R.R.; Michael, N.T. Sea-level trends and interannual variability in the Caribbean Sea. *J. Geophys. Res. Ocean* **2013**, *118*, 2934–2947. [\[CrossRef\]](#)
6. Hall, T.C.; Sealy, A.M.; Stephenson, T.S.; Kusunoki, S.; Taylor, M.A.; Chen, A.A.; Kitoh, A. Future climate of the Caribbean from a super-high-resolution atmospheric general circulation model. *Theor. Appl. Climatol.* **2013**, *113*, 271–287. [\[CrossRef\]](#)

7. Karmalkar, A.V.; Taylor, M.A.; Campbell, J.; Stephenson, T.; New, M.; Centella, A.; Benzanilla, A.; Charlery, J. A review of observed and projected changes in climate for the islands in the Caribbean. *Atmósfera* **2013**, *26*, 283–309. [[CrossRef](#)]
8. Bender, M.A.; Knutson, T.R.; Tuleya, R.E.; Sirutis, J.J.; Vecchi, G.A.; Garner, S.T.; Held, I.M. Modeled impact of anthropogenic warming on the frequency of intense Atlantic hurricanes. *Science* **2010**, *327*, 454–458. [[CrossRef](#)]
9. Knutson, T.R.; McBride, J.L.; Chan, J.; Emanuel, K.; Holland, G.; Landsea, C.; Held, I.; Kossin, J.P.; Srivastava, A.K.; Sugi, M. Tropical cyclones and climate change. *Nat. Geosci.* **2010**, *3*, 157–163. [[CrossRef](#)]
10. Mora, C.; Frazier, A.G.; Longman, R.J.; Dacks, R.S.; Walton, M.M.; Tong, E.J.; Sanchez, J.J.; Kaiser, L.R.; Stender, Y.O.; Anderson, J.M.; et al. The projected timing of climate departure from recent variability. *Nature* **2013**, *502*, 183–187. [[CrossRef](#)]
11. Portillo-Quintero, C.A.; Sánchez-Azofeifa, G.A. Extent and conservation of tropical dry forests in the Americas. *Biol. Conserv.* **2010**, *143*, 144–155. [[CrossRef](#)]
12. Nelson, H.P.; Devenish-Nelson, E.S.; Rusk, B.L.; Geary, M.; Lawrence, A.J. A call to action for climate change research on Caribbean dry forests. *Reg. Environ. Change* **2018**, *18*, 1337–1342. [[CrossRef](#)]
13. Becknell, J.M.; Kucek, L.K.; Powers, J.S. Aboveground biomass in mature and secondary seasonally dry tropical forests: A literature review and global synthesis. *For. Ecol. Manag.* **2012**, *276*, 88–95. [[CrossRef](#)]
14. Brandeis, T.J.; Helmer, E.H.; Marcano-Vega, H.; Lugo, A.E. Climate shapes the novel plant communities that form after deforestation in Puerto Rico and the US Virgin Islands. *For. Ecol. Manag.* **2009**, *258*, 1704–1718. [[CrossRef](#)]
15. McLaren, K.P.; McDonald, M.A. The effects of moisture and shade on seed germination and seedling survival in a tropical dry forest in Jamaica. *For. Ecol. Manag.* **2003**, *183*, 61–75. [[CrossRef](#)]
16. McLaren, K.P.; Lévesque, M.; Sharma, C.; Wilson, B.; McDonald, M.A. From seedlings to trees: Using ontogenetic models of growth and survivorship to assess long-term (>100 years) dynamics of a neotropical dry forest. *For. Ecol. Manag.* **2011**, *262*, 916–930. [[CrossRef](#)]
17. Rojas-Sandoval, J.; Meléndez-Ackerman, E. Reproductive phenology of the Caribbean cactus *Harrisia portoricensis*: Rainfall and temperature associations. *Botany* **2011**, *89*, 861–871. [[CrossRef](#)]
18. Jimenez-Rodríguez, D.L.; Alvarez-Añorve, M.Y.; Pineda-Cortes, M.; Flores-Puerto, J.I.; Benítez-Malvido, J.; Oyama, K.; Avila-Cabadilla, L.D. Structural and functional traits predict short term response of tropical dry forests to a high intensity hurricane. *For. Ecol. Manag.* **2018**, *426*, 101–114. [[CrossRef](#)]
19. Blackie, R.; Baldauf, C.; Gautier, D.; Gumbo, D.; Kassa, H.; Parthasarathy, N.; Paumgarten, F.; Sola, P.; Pulla, S.; Waeber, P.; et al. *Tropical Dry Forests: The State of Global Knowledge and Recommendations for Future Research*; CIFOR: Bogor, Indonesia, 2014.
20. Maharaj, S.S.; New, M. Modelling individual and collective species responses to climate change within Small Island States. *Biol. Conserv.* **2013**, *167*, 283–291. [[CrossRef](#)]
21. Day, Owen, and Caribbean Natural Resources Institute. *The Impacts of Climate Change on Biodiversity in Caribbean Islands: What We Know, What We Need to Know, and Building Capacity for Effective Adaptation*; Caribbean Natural Resources Institute: San Juan, Trinidad and Tobago, 2009.
22. Vogel, P. Seasonal hatchling recruitment and juvenile growth of the lizard *Anolis lineatopus*. *Copeia* **1984**, *1984*, 747–757. [[CrossRef](#)]
23. Sinervo, B.; Mendez-De-La-Cruz, F.; Miles, D.B.; Heulin, B.; Bastiaans, E.; Villagrán-Santa Cruz, M.; Lara-Resendiz, R.; Martínez-Méndez, N.; Calderón-Espinosa, M.L.; Meza-Lázaro, R.N.; et al. Erosion of lizard diversity by climate change and altered thermal niches. *Science* **2010**, *328*, 894–899. [[CrossRef](#)]
24. McLaren, K.; Monroe, S.; Wilson, B. The Arctic oscillation, climatic variability, and biotic factors influenced seedling dynamics in a Caribbean moist forest. *Ecology* **2016**, *97*, 2416–2435. [[CrossRef](#)]
25. Cox, R.M.; Calsbeek, R. Sexually antagonistic selection, sexual dimorphism, and the resolution of intralocus sexual conflict. *Am. Nat.* **2009**, *173*, 176–187. [[CrossRef](#)]
26. Gorman, G.C.; Licht, P. Seasonality in ovarian cycles among tropical *Anolis* lizards. *Ecology* **1974**, *55*, 360–369. [[CrossRef](#)]
27. Gullan, P.J.; Cranston, P.S. *The Insects: An Outline of Entomology*; John Wiley & Sons: Hoboken, NJ, USA, 2014.
28. Tanaka, L.K.; Tanaka, S.K. Rainfall and seasonal changes in arthropod abundance on a tropical oceanic island. *Biotropica* **1982**, *14*, 114–123. [[CrossRef](#)]
29. Jackman, T.R.; Irschick, D.J.; De Queiroz, K.; Losos, J.B.; Larson, A. Molecular phylogenetic perspective on evolution of lizards of the *Anolis grahami* series. *J. Exp. Zool.* **2002**, *294*, 1–16. [[CrossRef](#)] [[PubMed](#)]
30. Piantoni, C.; Navas, C.A.; Ibarquengoytía, N.R. Vulnerability to climate warming of four genera of New World iguanians based on their thermal ecology. *Anim. Conserv.* **2016**, *19*, 391–400. [[CrossRef](#)]
31. Chown, S.L.; Gaston, K.J. Exploring links between physiology and ecology at macro-scales: The role of respiratory metabolism in insects. *Biol. Rev.* **1999**, *74*, 87–120. [[CrossRef](#)]
32. Claussen, D. Studies of water loss in two species of lizards. *Comp. Biochem. Physiol.* **1967**, *20*, 115–130. [[CrossRef](#)]
33. Ryan, M.J.; Latella, I.M.; Giermakowski, J.T.; Snell, H.; Poe, S.; Pangle, R.E.; Gehres, N.; Pockman, W.T.; McDowell, N.G. Too dry for lizards: Short-term rainfall influence on lizard microhabitat use in an experimental rainfall manipulation within a piñon-juniper. *Funct. Ecol.* **2016**, *30*, 964–973. [[CrossRef](#)]
34. Andrew, N.R.; Hughes, L. Species diversity and structure of phytophagous beetle assemblages along a latitudinal gradient: Predicting the potential impacts of climate change. *Ecol. Entomol.* **2004**, *29*, 527–542. [[CrossRef](#)]
35. Kiritani, K. Predicting impacts of global warming on population dynamics and distribution of arthropods in Japan. *Popul. Ecol.* **2006**, *48*, 5–12. [[CrossRef](#)]

36. Thornthwaite, C.W. An approach toward a rational classification of climate. *Geogr. Rev.* **1948**, *38*, 55–94. [[CrossRef](#)]
37. Piyaphongkul, J.; Pritchard, J.; Bale, J. Can tropical insects stand the heat? A case study with the brown planthopper *Nilaparvata lugens* (Stål). *PLoS ONE* **2012**, *7*, e29409. [[CrossRef](#)] [[PubMed](#)]
38. Huey, R.B.; Deutsch, C.A.; Tewksbury, J.J.; Vitt, L.J.; Hertz, P.E.; Álvarez Pérez, H.J.; Garland Jr, T. Why tropical forest lizards are vulnerable to climate warming. *Proc. R. Soc. B Biol. Sci.* **2009**, *276*, 1939–1948. [[CrossRef](#)] [[PubMed](#)]
39. Algar, A.C.; Mahler, D.L. Area, climate heterogeneity, and the response of climate niches to ecological opportunity in island radiations of *Anolis* lizards. *Glob. Ecol. Biogeogr.* **2016**, *25*, 781–791. [[CrossRef](#)]
40. Kearney, M.; Shine, R.; Porter, W.P. The potential for behavioral thermoregulation to buffer “cold-blooded” animals against climate warming. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 3835–3840. [[CrossRef](#)]
41. Logan, M.L.; Cox, R.M.; Calsbeek, R. Natural selection on thermal performance in a novel thermal environment. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 14165–14169. [[CrossRef](#)]
42. McCluney, K.E.; Sabo, J.L. Water availability directly determines per capita consumption at two trophic levels. *Ecology* **2009**, *90*, 1463–1469. [[CrossRef](#)]
43. Phillips, B.L.; Munoz, M.M.; Hatcher, A.; Macdonald, S.L.; Llewelyn, J.; Lucy, V.; Moritz, C. Heat hardening in a tropical lizard: Geographic variation explained by the predictability and variance in environmental temperatures. *Funct. Ecol.* **2016**, *30*, 1161–1168. [[CrossRef](#)]
44. Wilson, B.; Grant, T.; van Veen, R.; Hudson, R.; Fleuchaus, D.; Robinson, O.; Stephenson, K. 25 Years of Conservation Effort for the Jamaican Iguana. *Herpetol. Conserv. Biol.* **2016**, *11*, 237–254.
45. Wilson, B.S.; Vogel, P. A Survey of the Herpetofauna of the Hellshire Hills, Jamaica, including the Rediscovery of the Blue-tailed Galliwasp (*Celestus duquesneyi* Grant). *Caribb. J. Sci.* **2000**, *36*, 244–249.
46. Lewis, D.S.; van Veen, R.; Wilson, B.S. Conservation implications of small Indian mongoose (*Herpestes auro-punctatus*) predation in a hotspot within a hotspot: The Hellshire Hills, Jamaica. *Biol. Invasions* **2011**, *13*, 25–33. [[CrossRef](#)]
47. Wege, D.C.; Ryan, D.; Varty, N.; Anadón-Irizarry, V.; Pérez-Leroux, A. *Ecosystem Profile: The Caribbean Islands Biodiversity Hotspot*; BirdLife International, Critical Ecosystem Partnership Fund: Washington, DC, USA, 2009.
48. Grant, T.D. Biosphere reserve to transshipment Port: Travesty for Jamaica’s Goat Islands. *IRCF Reptiles Amphib. Conserv. Nat. Hist.* **2014**, *21*, 37–43. [[CrossRef](#)]
49. Loveless, A.R.; Asprey, G.F. The Dry Evergreen Formations of Jamaica: I. The Limestone Hills of the South Coast. *J. Ecol.* **1957**, *45*, 799–822. [[CrossRef](#)]
50. Wilson, B.S. Conservation of Jamaican amphibians and reptiles. In *Conservation of Caribbean Island Herpetofaunas Volume 2: Regional Accounts of the West Indies*; Hailey, A., Wilson, B., Horrocks, J., Eds.; Brill: Leiden, The Netherlands, 2011; Volume 2, pp. 273–310.
51. R Development Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2017.
52. Simon, N.W. *Core Statistics*; Cambridge University Press: Cambridge, UK, 2015.
53. Alexander, L.V.; Zhang, X.; Peterson, T.C.; Caesar, J.; Gleason, B.; Klein Tank, A.M.G.; Haylock, M.; Collins, D.; Trewin, B.; Rahimzadeh, F.; et al. Global observed changes in daily climate extremes of temperature and precipitation. *J. Geophys. Res. Atmos.* **2006**, *111*, D05109. [[CrossRef](#)]
54. Frich, P.; Alexander, L.V.; Della-Marta, P.M.; Gleason, B.; Haylock, M.; Klein Tank, A.M.G.; Peterson, T. Observed coherent changes in Climatic Extremes During the Second Half of the Twentieth Century. *Clim. Res.* **2002**, *19*, 193–212. [[CrossRef](#)]
55. Stenseth, N.C.; Ottersen, G.; Hurrell, J.W.; Mysterud, A.; Lima, M.; Chan, K.S.; Yoccoz, N.G.; Ådlandsvik, B. Studying climate effects on ecology through the use of climate indices: The North Atlantic Oscillation, El Niño Southern Oscillation and beyond. *Proc. R. Soc. Lond. Ser. B Biol. Sci.* **2003**, *270*, 2087–2096. [[CrossRef](#)]
56. Calsbeek, R.; Sinervo, B. Correlational Selection on Lay Date and Life-history traits: Experimental manipulations of territory and Nest Site Quality. *Evolution* **2007**, *61*, 1071–1083. [[CrossRef](#)]
57. Chen, A.A.; Taylor, M.A. Investigating the Link Between Early Season Caribbean Rainfall and the El Niño+1 Year. *Int. J. Climatol.* **2002**, *22*, 87–106. [[CrossRef](#)]
58. Giannini, A.; Kushnir, Y.; Cane, M.A. Interannual variability of Caribbean rainfall, ENSO, and the Atlantic Ocean. *J. Clim.* **2000**, *13*, 297–311. [[CrossRef](#)]
59. Gouirand, I.; Moron, V.; Hu, Z.Z.; Jha, B. Influence of the warm pool and cold tongue El Niños on the following Caribbean rainy season rainfall. *Clim. Dyn.* **2014**, *42*, 919–929. [[CrossRef](#)]
60. Taylor, M.A.; Enfield, D.B.; Chen, A.A. Influence of the tropical Atlantic versus the tropical Pacific on Caribbean rainfall. *J. Geophys. Res. Oceans* **2002**, *107*, 10–11. [[CrossRef](#)]
61. Yun, K.S.; Yeh, S.W.; Ha, K.J. Inter-El Niño variability in CMIP5 models: Model deficiencies and future changes. *J. Geophys. Res. Atmos.* **2016**, *121*, 3894–3906. [[CrossRef](#)]
62. Stephenson, T.S.; Chen, A.A.; Taylor, M.A. Toward the development of prediction models for the primary Caribbean dry season. *Theor. Appl. Climatol.* **2008**, *92*, 87–101. [[CrossRef](#)]
63. McLeod, A.I.; Xu, C.; Lai, Y. *bestglm: Best Subset GLM and Regression Utilities*, R package version 0.37.3; R Foundation for Statistical Computing: Vienna, Austria, 2020; Available online: <https://CRAN.R-project.org/package=bestglm> (accessed on 21 January 2022).

64. McLeod, A.I.; Xu, C. *bestglm: Best Subset GLM*; R Package Version 0.31; R Foundation for Statistical Computing: Vienna, Austria, 2010; Available online: <https://cran.r-project.org/src/contrib/Archive/bestglm/> (accessed on 21 January 2022).
65. Bates, D.; Maechler, M.; Bolker, B.; Walker, S.; Christensen, R.H.B.; Singmann, H.; Dai, B.; Grothendieck, G.; Green, P.; Bolker, M.B. *Package "lme4"*, R package version 1.1-10; The R Project for Statistical Computing: Vienna, Austria, 2016.
66. Henderson, R.W.; Powell, R. *Natural History of West Indian Reptiles and Amphibians*; University Press of Florida: Gainesville, FL, USA, 2009.
67. Ripley, B.; Venables, B.; Bates, D.M.; Hornik, K.; Gebhardt, A.; Firth, D. Package 'mass'. *Cran R* **2013**, *538*, 113–120.
68. Barton, K. *MuMIn: Multi-Model Inference*, R package version 1.46.0; R Foundation for Statistical Computing: Vienna, Austria, 2022; Available online: <https://CRAN.R-project.org/package=MuMIn> (accessed on 21 January 2022).
69. Bakar, K.S.; Sahu, S.K. spTimer: Spatio-temporal bayesian modelling using R. *J. Stat. Softw.* **2015**, *63*, 1–32. [[CrossRef](#)]
70. Centella-Artola, A.; Taylor, M.A.; Bezanilla-Morlot, A.; Martinez-Castro, D.; Campbell, J.D.; Stephenson, T.S.; Vichot, A. Assessing the effect of domain size over the Caribbean region using the PRECIS regional climate model. *Clim. Dyn.* **2015**, *44*, 1901–1918. [[CrossRef](#)]
71. Stephenson, K. Modelling the Impact of Climate Change on a Dry Forest Fauna. Ph.D. Thesis, The University of the West Indies, Mona Campus, Jamaica, 2017.
72. Vrcibradic, D.; Rocha, C.F.D. Reproductive cycle and life-history traits of the viviparous skink *Mabuia frenata* in southeastern Brazil. *Copeia* **1998**, 612–619. [[CrossRef](#)]
73. Kingsbury, B.A. Thermal constraints and eurythermy in the lizard *Elgaria multicarinata*. *Herpetologica* **1994**, 266–273.
74. Curio, E.; Möbius, H. Versuche zum Nachweis eines Riechvermögens von *Anolis l. lineatopus* (Rept., Iguanidae). *Z. Tierpsychol.* **1978**, *47*, 281–292.
75. Von Brockhusen-Holzer, F.; Curio, E. Ethotypic variation of prey recognition in juvenile *Anolis lineatopus* (Reptilia: Iguanidae). *Ethology* **1990**, *86*, 19–32. [[CrossRef](#)]
76. Polis, G.A.; Myers, C.A.; Holt, R.D. The ecology and evolution of intraguild predation: Potential competitors that eat each other. *Annu. Rev. Ecol. Syst.* **1989**, *20*, 297–330. [[CrossRef](#)]
77. Spiller, D.A.; Schoener, T.W. An experimental study of the effect of lizards on web-spider communities. *Ecol. Monogr.* **1988**, *58*, 57–77. [[CrossRef](#)]
78. Best, T.L.; Gennaro, A.L. Feeding ecology of the lizard, *Uta stansburiana*, in southeastern New Mexico. *J. Herpetol.* **1984**, *18*, 291–301. [[CrossRef](#)]
79. Catenazzi, A.; Brookhart, J.O.; Cushing, P.E. Natural history of coastal Peruvian solifuges with a redescription of *Chinchippus peruvianus* and an additional new species (Arachnida, Solifugae, Ammotrechidae). *J. Arachnol.* **2009**, *37*, 151–159. [[CrossRef](#)]
80. Cloudsley-Thompson, J.L. *Spiders, Scorpions, Centipedes and Mites: The Commonwealth and International Library: Biology Division*; Elsevier: Amsterdam, The Netherlands, 2015.
81. Chapin, K.J.; Hebets, E.A. The behavioral ecology of amblypygids. *J. Arachnol.* **2016**, *44*, 1–14. [[CrossRef](#)]
82. Polis, G.A. *The Biology of Scorpions*; Stanford University Press: Palo Alto, CA, USA, 1990.
83. Yamashita, T. Surface activity, biomass, and phenology of the striped scorpion, *Centruroides vittatus* (Buthidae) in Arkansas, USA. *Euscorpis* **2004**, *17*, 25–33.
84. Miller, R.H.; Cameron, G.N. Effects of temperature and rainfall on populations of *Armadillidium vulgare* (Crustacea: Isopoda) in Texas. *Am. Midl. Nat.* **1987**, *117*, 192–198. [[CrossRef](#)]
85. Warburg, M.R. *Evolutionary Biology of Land Isopods*; Springer Science & Business Media: Berlin/Heidelberg, Germany, 2013.
86. Wolcott, G.N. The Status of Economic Entomology in Peru. *Bull. Entomol. Res.* **1929**, *20*, 225–231. [[CrossRef](#)]
87. Schaefer, C.W.; Ahmad, I. Cotton stainers and their relatives (Pyrrhocoroidea: Pyrrhocoridae and Largidae). In *Heteroptera of Economic Importance*; CRC Press: Boca Raton, FL, USA, 2000; pp. 271–308.
88. Polidori, C.; Beneitez, A.; Asís, J.D.; Tormos, J. Scramble competition by males of the velvet ant *Nemka viduata* (Hymenoptera: Mutillidae). *Behaviour* **2013**, *150*, 23–37.
89. Vieira, C.R.; Pitts, J.; Colli, G.R. Microhabitat changes induced by edge effects impact velvet ant (Hymenoptera: Mutillidae) communities in southeastern Amazonia, Brazil. *J. Insect Conserv.* **2015**, *19*, 849–861. [[CrossRef](#)]
90. Yang, S.; Liu, Z.; Xiao, Y.; Li, Y.; Rong, M.; Liang, S.; Zhang, Z.; Yu, H.; King, G.F.; Lai, R. Chemical punch packed in venoms makes centipedes excellent predators. *Mol. Cell. Proteom.* **2012**, *11*, 640–650. [[CrossRef](#)] [[PubMed](#)]
91. Guizze, S.P.; Knysak, I.; Barbaro, K.C.; Karam-Gemael, M.; Chagas Jr, A. Predatory behavior of three centipede species of the order *Scolopendromorpha* (Arthropoda: Myriapoda: Chilopoda). *Zoologia* **2016**, *33*, e20160026. [[CrossRef](#)]
92. Perez-Gelabert, D.E.; Edgecombe, G.D. Scutigermorph centipedes (Chilopoda: Scutigermorpha) of the Dominican Republic, Hispaniola. *Novit. Caribaea* **2013**, *6*, 36–44. [[CrossRef](#)]
93. Shelley, R.M.; Sikes, D.S. Centipedes and Millipeds (Arthropoda: Diplopoda, Chilopoda) from Saba Island, Lesser Antilles, and a Consolidation of Major References on the Myriapod Fauna of "Lesser" Caribbean Islands. *Insecta Mundi* **2012**, *742*, 1–9.
94. Leśniewska, M.; Jastrzębski, P.; Stańska, M.; Hajdamowicz, I. Centipede (Chilopoda) richness and diversity in the Bug River valley (Eastern Poland). *ZooKeys* **2015**, *510*, 125. [[CrossRef](#)]
95. Bachvarova, D.; Doichinov, A.; Stoev, P.; Kalchev, K. Habitat preferences and effect of environmental factors on the seasonal activity of *Lithobius nigripalpis* L. Koch, 1867 (Chilopoda: Lithobiomorpha: Lithobiidae). In Proceedings of the 16th International Congress of Myriapodology, Olomouc, Czech Republic, 20–25 July 2014; p. 5.

96. Cole, C.L. Stratification and survival of diapausing burrowing bugs. *Southwest. Entomol.* **1988**, *13*, 243–246.
97. Riis, L.; Esbjerg, P.; Bellotti, A.C. Influence of temperature and soil moisture on some population growth parameters of *Cyrtomenus bergi* (Hemiptera: Cydnidae). *Fla. Entomol.* **2005**, *88*, 11–22. [[CrossRef](#)]
98. Wheatly, M.G.; Burggren, W.W.; McMahon, B.R. The effects of temperature and water availability on ion and acid-base balance in hemolymph of the land hermit crab *Coenobita clypeatus*. *Biol. Bull.* **1984**, *166*, 427–445. [[CrossRef](#)]
99. Hamasaki, K.; Kato, S.; Murakami, Y.; Dan, S.; Kitada, S. Larval growth, development and duration in terrestrial hermit crabs. *Sex. Early Dev. Aquat. Org.* **2015**, *1*, 93–107. [[CrossRef](#)]
100. González, C.; Paz, A.; Ferro, C. Predicted altitudinal shifts and reduced spatial distribution of *Leishmania infantum* vector species under climate change scenarios in Colombia. *Acta Trop.* **2014**, *129*, 83–90. [[CrossRef](#)] [[PubMed](#)]
101. Reina-Rodríguez, G.A.; Rubiano Mejía, J.E.; Castro Llanos, F.A.; Soriano, I. Orchid distribution and bioclimatic niches as a strategy to climate change in areas of tropical dry forest in Colombia. *Lankesteriana* **2017**, *17*, 17–47. [[CrossRef](#)]
102. Fain, S.J.; Quiñones, M.; Álvarez-Berrios, N.L.; Parés-Ramos, I.K.; Gould, W.A. Climate change and coffee: Assessing vulnerability by modeling future climate suitability in the Caribbean island of Puerto Rico. *Clim. Change* **2018**, *146*, 175–186. [[CrossRef](#)]