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Citation: Harrington, David, Guy, Jonathan, Robinson, Karen and Sparagano, Olivier (2010) Comparison of synthetic membranes in the development of an in vitro feeding system for *Dermanyssus gallinae*. *Bulletin of Entomological Research*, 100 (02). pp. 127-132. ISSN 0007-4853

Published by: Cambridge University Press

URL: <http://dx.doi.org/10.1017/s0007485309006865>
<<http://dx.doi.org/10.1017/s0007485309006865>>

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Comparison of synthetic membranes in the development of an *in vitro* feeding system for *Dermanyssus gallinae*

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Abstract

Although artificial feeding models for the poultry red mite (*Dermanyssus gallinae*) most frequently use biological membranes consisting of day-old chick skin, there are ethical considerations associated with the use of skin. The few studies reported in the literature that have investigated the use of synthetic membranes to feed *D. gallinae in vitro* have reported limited success. The current study describes an investigation into the use of synthetic membranes made from either Nescofilm[®] or rayon and silicone, used either alone or in combination with different feather or skin extracts, as well as the use of capillary tubes. In all, 12 different treatments were used, and the feeding rate of *D. gallinae* was compared to that of day-old chick skin. Allowing mites to feed on a membrane consisting of Nescofilm with a skin extract resulted in the highest proportion of mites feeding (32.3%), which was not significantly different to the feeding rate of mites on day-old chick skin (38.8%). This study confirms that synthetic membranes can be used to feed *D. gallinae* artificially. Further optimization of the membrane and mite storage conditions is still necessary, but the study demonstrates a proof of concept.

Keywords: *Dermanyssus gallinae*, synthetic membranes, *in vitro* feeding, arthropods

(Accepted 13 February 2009)

Introduction

The poultry red mite, *Dermanyssus gallinae*, is a haematophagous mite from the order Mesostigmata. Whilst the poultry red mite has a broad host range and has been found associated with gerbils and horses, its main association is with birds (Lucky *et al.*, 2001; Mignon & Losson, 2008). *D. gallinae* is considered the most significant ectoparasite of egg-laying poultry in Europe (Chauve, 1998) although it has a worldwide distribution. Deleterious effects on the avian

host can include mild or fatal anaemia, loss in egg production and elevated stress levels (Kowalski *et al.*, 2006). Substantial *D. gallinae* infestations have been reported to impact upon poultry workers by causing dermatitis (Rosen *et al.*, 2002). *D. gallinae* is currently controlled using a regime of hygiene and application of synthetic chemicals, but reports of increasing acaricide resistance suggest alternative approaches are needed. New approaches under investigation by our group include vaccines (Arkle *et al.*, 2008) and plant-derived products (George *et al.*, 2008), approaches which often require the maintenance of populations of *D. gallinae* under laboratory conditions.

Populations of *D. gallinae* can be maintained in the laboratory by allowing mites to feed periodically on live hens (Tucci, 1997), a method which has inherent welfare

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Table 1. Experimental treatment groups

Group	Treatment	Treatment acronym
1	Day-old chick skin	SK
2	Nescofilm	NF
3	Rayon/70% silicone	R/70S
4	Rayon/50% silicone	R/50S
5	Nescofilm + skin extract	NF + SE
6	Rayon/70% silicone + skin extract	R/70S + SE
7	Rayon/50% silicone + skin extract	R/50S + SE
8	Nescofilm + feather extract	NF + FE
9	Rayon/70% silicone + feather extract	R/70S + FE
10	Rayon/50% silicone + feather extract	R/50S + FE
11	Nescofilm + ethyl acetate	NF + EA
12	Capillary tube	CT

disadvantages. Depending upon the number of mites feeding, the bird can experience stress and general discomfort during the procedure. Kilpinen *et al.* (2005) infested groups of laying hens with *D. gallinae* and observed both their behavior and health in comparison to non-infested controls. The study found that following introduction of *D. gallinae* into the floor pen on two occasions, the mite populations ranged from approximately 3000 to 12,000 mites per 100 ml litter at the peak of infestation, equating to approximately 200–800 mites per bird. Significantly increased self-grooming behaviour was noted in infested birds, as well as increased feather pecking, reduced packed cell volume (PCV) and mortality due to anaemia. Instead of using a live host, a number of authors have shown that *D. gallinae* can also be fed using chicken skin under artificial conditions (Kirkwood, 1968; Zeman, 1988; Bruneau *et al.*, 2001; McDevitt *et al.*, 2006). However, the conditions to feed *D. gallinae* in the laboratory are quite specific. Despite being cosmopolitan in its choice of host *in vivo*, *D. gallinae* typically will only feed under *in vitro* conditions when the feeding model consists of day-old chick skin. The use of day-old chick skin clearly has an ethical and welfare disadvantage in that it involves the euthanasia of young animals. A reduction in the number of animals used for either live feeding or provision of skin can contribute to the overall aim of national guidelines on reduction and refinement of procedures used on experimental animals. Artificial feeding models are attractive for a second reason, namely they can reduce intra-assay variability, especially where variability due to individual host-parasite relationships can have an effect (Kröber & Guerin, 2007a). Such lower-cost models can also allow for an increased number of replicates for a given treatment, thereby increasing the power of statistical analyses. Artificial feeding models have a wide range of applications, including the investigation of arthropod transmitted pathogens, identification of biologically active arthropod molecules and testing of antibodies against arthropod antigens and insecticide/acaricide testing (Kröber & Guerin, 2007a). A range of other methods, such as wool soaked with serum, capillary tubes and silicone membranes, have been successfully used in *in vitro* feeding systems of other arthropod ectoparasites (Kocan *et al.*, 2005; Thind & Ford, 2007) whilst synthetic membranes have been used in tick feeding models to test acaricide efficacy against *Ixodes ricinus* (Kröber & Guerin, 2007b).

However, the use of synthetic membranes to feed *D. gallinae* has met with limited success. Zeman (1988) and

Bruneau *et al.* (2001) both reported that *D. gallinae* would not feed on Parafilm 'M' or a synthetic skin substitute. Zeman (1988) identified the role of kairomones (a sub-class of pheromones that benefit the receiver) in skin and feathers of poultry as an attractant to *D. gallinae* and, consequently, produced the only report of successfully feeding *D. gallinae* via synthetic membranes.

Therefore, the aim of this experiment was to investigate the use of alternative synthetic membranes and capillary tubes to feed *D. gallinae in vitro* by combining the principles of skin and feather compound extraction of Zeman (1988) with the production of silicon membranes described by Kröber & Guerin (2007b).

Material and methods

Experimental design

The experiment comprised 12 experimental treatment groups as shown in table 1 and was undertaken in two blocks. Treatments consisted of a feeding chamber with one of 12 different membranes: day-old chick skin (SK); capillary tube (CT); or combinations of Nescofilm (NF) or silicone and rayon membranes of differing silicone concentrations (R/S) and ethyl acetate either alone or in combination with skin or feather extract (EA, SE and FE, respectively). Five replicates per block were used in the first block and four replicates were used in the second block (due to a reduced number of available unfed *D. gallinae*) with the exception of treatments 11 and 12 where five replicates were used in block two only.

Manufacture of synthetic membranes and preparation of skin

Synthetic membranes were constructed from Nescofilm[®] (Nippon Shoji Kaisha Ltd, Osaka, Japan), a sealing film for use in laboratories, or a combination of rayon and silicone sealant. Membranes using Nescofilm (NF) were made by stretching the film as thin as possible without tearing it. Silicone membranes (R/S) were made following an adaption of the method of Kröber & Guerin (2007b). Silicone sealant Ceresit 10B (Henkel Loctite Adhesives Ltd, Winsford, UK) with a shore A hardness of 25° (a measure of the relative hardness of an elastic material; the lower the value, the softer the material) was mixed with 30% DC 200 silicone oil (Sigma-Aldrich Ltd, Poole, UK) in order to achieve either a 50:50 silicone:silicone oil mix or 70:30 silicone:silicone oil mix. Hexane was then added at a rate of 1.5 g per 1 g silicone mixture to render the mixture temporarily more fluid to assist application. Pieces of rayon (regenerated cellulose; Lens Saver[®] lens cloth, Hear Saver Ltd, Stoney Creek, Canada), approximately 70 × 60 mm, were laid on a layer of plastic film, and the silicone mixture was applied using a silicone spatula. The membranes were allowed to air dry for a minimum of seven days before use. All membranes were examined under 10 × magnification to ensure there were no holes in them prior to use. Day-old skin was harvested from euthanased day-old layer chicks, which was plucked, scraped of fat and stored frozen at –20°C. Skin was thawed prior to use.

Skin and feather extracts

Feather and skin extracts for use with the membranes were prepared following the method of Zeman (1988). Skin

and feathers were collected from 18 week-old laying hens euthanased by cervical dislocation. Skin (5.5 g) was finely chopped and added to 11 ml 99.5% ethyl acetate. The mixture was placed at 2–8°C for five days, and then the aqueous phase was removed and stored at –20°C and the skin was discarded. Approximately 2 g of feathers were finely chopped and added to 10 ml 99.5% ethyl acetate. The sample was allowed to sit at room temperature for 30 min and then spun at 13,000×*g* for 15 min. The supernatant was removed and stored, and the feathers were spun again for 15 min at 13,000×*g*. The supernatant was removed and added to the first supernatant sample collected. The supernatant was stored at –20°C and the feathers were discarded. Where appropriate, 10 µl of feather or skin extracts were added to membranes of groups 5–10. Group 11 received 10 µl ethyl acetate only. Following the addition of ethyl acetate or extract, membranes were air dried for 45 min before use in the chambers.

Capillary tube

Blood (see subsequent section) was drawn up into heparinised capillary tubes (Hawksley & Sons Ltd, Lancing, England), and the tube was sealed at one end using Cristaseal clay (Hawksley & Sons Ltd, Lancing, England).

Source of *D. gallinae*

D. gallinae were obtained from a commercial poultry farm and kept at 22°C in the dark in flat-bottomed tissue flasks sealed with a fine gauze for 7–10 days to allow digestion of the blood meal. Mites were then kept at 4°C in the dark for a minimum of four weeks to ensure that they were starved.

Source of blood

Nine laying hens (42 days old) with no known exposure to *D. gallinae* were euthanased by cervical dislocation, and blood was collected by cardiac puncture into glass vacutainers containing 200 units sodium heparin (Sigma-Aldrich Ltd, Poole, UK). Blood was stored at 2–8°C and used within two days of collection. Prior to use in the feeding model, blood was allowed to warm to room temperature and the blood samples pooled to provide the required volume of blood for testing synthetic membranes on that occasion.

Design and assembly of feeding chamber

The feeding apparatus used has been described previously by Harrington *et al.* (2009). Briefly, it comprised a clear glass vial with a modified silicon/poly(tetrafluoroethylene) PTFE ring (a disc, with the centre cross cut removed) placed inside a snap ring plastic cap with a hole in the centre. The membrane to be tested was fitted to the ring, cut to size and placed between the ring and cap. A 0.5-ml tube with the end removed was secured to the top of the lid using modeling clay to act as a blood reservoir. For day-old chick skin, the skin was stretched across the ring such that the outer surface of the skin was facing into the vial. Capillary tubes were used in the feeding chamber by using unmodified silicon/PTFE discs; the tube was pushed through the cross cut and held in place using modeling clay. As described earlier, synthetic membranes were cut to size and placed in the chamber.

Unfed mites (20 females) were collected into the vial and the vial sealed with the ring/membrane/cap. The reservoir was filled with 200 µl fresh heparinised blood. Mite chambers were kept in an incubator (WTB Binder, Tuttlingen, Germany) in the dark at 37°C and 75–85% RH. Mites were examined 17 h after being allowed to feed, and the number of mites which had fed was observed. Mites were defined to have fed if they were engorged and/or had changed colour from pale grey/brown to bright red (McDevitt *et al.*, 2006).

Statistical analysis

To evaluate feeding rates on the synthetic membranes, data were transformed to conform to a normal distribution and analysed by analysis of variance (ANOVA) using the general linear model (GLM) command fitting treatment and treatment as factors. Tukey's *t*-test was used for pairwise comparisons between treatments. Treatment 12 (CT) was excluded from statistical analysis since no mites fed in this group and inclusion of the group created a highly skewed data distribution. Statistical significance was declared at $P \leq 0.05$. Data are presented as arithmetic mean ± standard error (SE) and the *F* statistic is shown. The total number of mites in each chamber was the denominator in all calculations of the mean. All statistical analyses were performed using Minitab 15.1.2 for Windows (Minitab Inc., College, USA).

Results

Assessment of feeding rate

No mites fed on blood presented by CT. In the first block, feeding rates in all treatments ranged from 0.0 to 33.8%, whilst in the second block feeding rates ranged from 0.0 to 45.1% (table 2). The highest feeding rates were recorded for SK.

Overall, proportion of fed mites was highest in the treatment using SK, where 38.8% mites fed, whilst the use of NF+SE resulted in the numerically highest percentage of mites feeding compared to all other groups using synthetic membranes, 32.3% (table 2). The membranes R/70S resulted in the lowest numerical proportion of fed mites on any of the synthetic membranes with just 5.3% of mites feeding. There was a highly significant effect of treatment upon the mean proportion of fed mites ($F_{10,79} = 4.38$, $P < 0.001$). A significantly higher proportion ($P < 0.01$) of mites fed on SK than R/70S, R/50S, R/50S+SE and R/70+FE. The proportion of fed mites was significantly higher ($P < 0.05$) on NF+SE than R/70S, R/50S and R/50S+SE.

There was no significant effect of block or extract type when all proportion of fed mites on all synthetic membranes were compared. Mean feeding rate of both experiments for all synthetic membranes without extract, all synthetic membranes with SE and all synthetic membranes with FE was 8.8, 19.2 and 11.8%, respectively. Similarly, when data for silicone-based membranes only (R/70S, R/50S) were analysed for effect of extract, there was no significant effect of block or extract. However, analysis of the effect of extract on the proportion of fed mites on NF membranes, showed that significantly more mites fed on NF+SE than NF alone or NF+FE ($F_{2,23} = 7.39$, $P < 0.05$).

Table 2. Total number of mites alive and fed (F), alive and unfed (UF), dead and fed (DF) and dead unfed (DUF) for each experiment and mean percentage (\pm SE) of fed mites in each treatment for both blocks and the mean of blocks 1 and 2 (except treatment 11).

Treatment	Total number of mites for all replicates								Mean percentage total fed mites*			
	Experiment 1 ($n=100$)				Experiment 2 ($n=80^{**}$)				Experiment		Mean	
	F	UF	DF	DUF	F	UF	DF	DUF	1 ($n=5$)	2 ($n=4$)		
1 SK	29	53	5	13	32	5	33	10	33.8 (± 3.7)	45.1 (± 19.7)	38.8 ^a	
2 NF	5	85	3	7	6	9	48	17	8.2 (± 2.1)	17.6 (± 7.7)	12.4 ^{a,c}	
3 R/70S	4	90	2	4	3	2	57	18	4.7 (± 3.9)	6.1 (± 0.9)	5.3 ^{b,c}	
4 R/50S	3	89	1	7	9	3	52	16	3.3 (± 2.5)	15.2 (± 6.7)	8.6 ^{b,c}	
5 NF+SE	29	49	4	18	17	6	32	25	34.6 (± 8.1)	29.5 (± 13.2)	32.3 ^a	
6 R/70S+SE	32	2	42	24	2	2	53	23	30.0 (± 14.7)	5.0 (± 2.0)	18.9 ^{a,c}	
7 R/50S+SE	2	75	5	18	1	3	31	45	7.5 (± 4.1)	5.0 (± 3.5)	6.4 ^{b,c}	
8 NF+FE	11	66	1	22	6	0	54	20	11.4 (± 3.2)	7.9 (± 2.4)	9.9 ^{a,c}	
9 R/70S+FE	17	5	60	18	3	1	52	24	20.6 (± 13.2)	4.8 (± 3.4)	12.7 ^{b,c}	
10 R/50S+FE	8	78	1	13	8	5	52	15	8.9 (± 2.2)	15.6 (± 2.0)	13.6 ^{a,c}	
11 NF+EA	–	–	–	–	11	8	53	28	–	–	19.6 (± 5.8)	19.6 ^{a,c}

Within a column, values with different superscripts are significantly different at $P \leq 0.05$. * Mean percentage of total fed mites and total number of fed mites differ due to rounding errors from means of replicates. ** Except treatment 11, $n=100$.

Discussion

This current study was designed to investigate the use of synthetic membranes for *in vitro* feeding of *D. gallinae* in comparison to using day-old chick skin. There is only one report in the literature of synthetic membranes being used successfully for *in vitro* *D. gallinae* feeding, and the use of silicone membranes has not been previously reported for *D. gallinae*. In the current study, the proportion of mites feeding on a synthetic membrane was not significantly different from that observed with skin.

Zeman (1988) has previously demonstrated the importance of kairomones as a stimulus for *D. gallinae* to feed artificially although, when feeding on a live host, other factors, such as temperature and carbon dioxide, have been shown to play a role (Kilpinen 2001, 2005). Using fresh blood stabilized in 2% EDTA, Zeman (1988) found that mites did not feed on Parafilm membranes unless they had been impregnated with a feather or skin extract. Bruneau *et al.* (2001) reported similar findings, where no mites fed on either of the synthetic membranes used in their study (Parafilm 'M' and a synthetic skin substitute, pansement hydrocolloïde 0.3 mm). In contrast, in the current study, *D. gallinae* fed on synthetic membranes that had not been impregnated with either skin or feather extract, although the feeding rates were relatively low, 5.3 to 12.4%. The use of ethyl acetate without skin or feather extract also appeared to have some attractant effect for mite feeding and led to increased feeding rate compared to Nescofilm alone. Mean feeding rates were higher for Nescofilm membranes + ethyl acetate than in all other synthetic membranes with the exception of Nescofilm + skin extract although, in all cases, the difference was not statistically significant. Zeman (1988) did not report data on mite feeding rates on Parafilm + organic solvent only, and our data would suggest that whilst kairomones are important, a contributory factor to their effectiveness *in vitro* is the solvent itself.

Following the impregnation of membranes with feather or skin extract, feeding rates increased in some but not all treatments. Nescofilm impregnated with skin extract resulted in the highest proportion of fed mites of all synthetic membranes (32.3%), which was not significantly different from that seen with skin (38.8%). This is in contrast to the

observation reported by Zeman (1988), where the highest proportion of mites was observed to feed on Parafilm 'M' impregnated with feather extract. In that study, 30.1% mites fed, which was 9.4% lower than that observed with two-week-old chick skin. In the current study, Nescofilm impregnated with feather extract achieved only a 9.9% mean feeding rate, although this was not significantly different from any other treatment. Similarly, when the effect of extract was examined irrespective of synthetic membrane type, membranes impregnated with skin extract had the highest proportion of mite feeding, 19.2%, compared to 8.8% without extract or 11.8% with feather extract. The difference in the effectiveness of feather and skin extract observed between the current study and that of Zeman (1988) could be related to the extraction method used. In the current study, whilst the method for skin extraction was the same as used by Zeman (1988), the method for feather extraction differed. Whilst Zeman (1988) used a combination of extraction in ethyl acetate followed by evaporation of ethyl acetate under vacuum followed by solubilizing the residue in benzene, in the current study only ethyl acetate was used. However, effectiveness of extract is also likely to be related to properties of the membrane, including thickness. Kröber & Guerin (2007a) describe the importance of the thickness of the silicone membrane in feeding assays to account for both species and developmental stage differences in hypostome length, such as a softer and thinner (60–150 μ m) silicone membrane for *Ixodes ricinus*, which have a shorter hypostome than *Amblyomma hebraeum* in ticks. Membrane thickness was not measured in the current study, but given that adult female *D. gallinae* are considerably smaller in size than adult female *I. ricinus* (1 mm and 2–3 mm, respectively), membrane thickness could obviously be a significant factor in mite feeding with smaller hypostomes than ticks. Similarly, the composition of the membrane is important to ensure it is soft enough for the mite's mouthparts to penetrate. The higher feeding rates observed with Nescofilm are most probably a result of the lower thickness of the membrane. Carroll *et al.* (1992) successfully used sacs made of stretched Parafilm to feed the northern fowl mite, *Ornithonyssus sylviarum*, a mite similar in size to *D. gallinae*.

As mentioned previously, both biological and synthetic membranes have been used to present the blood meal to

D. gallinae. In the current study, *D. gallinae* did not feed on blood-filled capillary tubes without membranes. The role of ticks in the transmission of *Anaplasma marginale* has been studied by adhering capillary tubes to the head of the tick, *Dermacentor variabilis* (Kocan *et al.*, 2005). This is impractical for *D. gallinae* since it is a short-term feeder, and the bore of a capillary tube is larger than the mite itself. However, it is possible that the provision of blood to mites using very fine needles or blunt cannula, could overcome the need for membranes, although the lack of the kairomone stimulus would still need to be addressed in an area of further research.

McDevitt *et al.* (2006) suggested the importance of 'conditioning' *D. gallinae* to promote high feeding rates. They indicated that starving mites for seven days at 20°C following by a period of cooling at 1°C for 30 days resulted in 68–72% of mites feeding on blood (treated with the anticoagulant sodium heparin) through day-old chick skin, compared to a feeding rate of 24% in mites kept at 20 ± 3°C ('not conditioned'). In contrast, feeding rate on skin in the current study was 38.8%, despite using similar conditions for mite conditioning as described by McDevitt *et al.* (2006). This suggests that there are additional factors which determine optimal feeding rates of mites in individual laboratories, perhaps due to factors other than just temperature, such as humidity or the nature of the storage vessels for the mites themselves.

Further studies are required to optimize synthetic membranes as successful substitutes to day-old chick skin in *D. gallinae* feeding models to encourage higher mite feeding rates. Statistically, there was no single treatment that resulted in significantly higher *D. gallinae* feeding rates than all others, despite large numerical differences. This is a problem of the artificial assay and an area of further development necessary to reduce the intra-assay variation in feeding rates observed in the current study. Bruneau *et al.* (2001) suggested that mites adapt to the *in vitro* feeding system following high mortality in the first generation. Whilst completion of the mite life cycle was not a parameter studied in the current experiment, maintenance of mite colonies adapted to synthetic membranes could reduce the intra-assay variation observed, although testing to ensure the laboratory mite population was still representative of the 'wild' population would be necessary.

Optimization of the assay should focus on improving liveability of mites in chambers, as well as refining membranes. However, this study does demonstrate that alternative membranes to Nescofilm or Parafilm are possible (with further development). Synthetic membranes or alternative methods of blood meal presentation should be an active area of *D. gallinae* research to address the ethical consideration of reducing animal use in research. Samples for use in skin and feather extracts can be sourced from commercial poultry slaughterhouses, thereby reducing the need to maintain and euthanase birds specifically for this purpose.

Conclusion

D. gallinae were observed to feed through synthetic membranes comprising either Nescofilm or a silicone sealant/silicone oil mixture and rayon. Feeding rates were highest in a membrane consisting of Nescofilm impregnated with skin extract, which gave a statistically similar feeding

rate to that observed with day-old chick skin. However, optimization of the membrane properties and testing conditions is still required.

Acknowledgements

This work was supported by the Biological and Biotechnology Research Council. We would like to thank Dr Robert Shiel for help with data analysis.

References

- Arkle, S., Harrington, D., de Luna, C., George, D., Guy, J. & Sparagano, O.A.E. (2008) Immunological control of poultry red mite: the use of whole mite antigens as a candidate vaccine. *Annals of the New York Academy of Sciences* **1149**, 36–40.
- Bruneau, A., Dernburg, A., Chauve, C. & Zenner, L. (2001) First *in vitro* cycle of the chicken mite, *Dermanyssus gallinae* (DeGeer, 1778), utilizing an artificial feeding device. *Parasitology* **123**, 583–589.
- Carroll, J.F., Young, K.W. & Bruce, W.A. (1992) Simple *in vitro* feeding system for northern fowl mites (Acari: Macronyssidae). *Journal of Economic Entomology* **85**, 848–852.
- Chauve, C. (1998) The poultry red mite *Dermanyssus gallinae* (De Geer, 1778): current situation and future prospects for control. *Veterinary Parasitology* **79**, 239–245.
- George, D.R., Callaghan, K., Guy, J.H. & Sparagano, O.A.E. (2008) Lack of prolonged activity of lavender essential oils as acaricides against the poultry red mite (*Dermanyssus gallinae*) under laboratory conditions. *Research in Veterinary Science* **85**, 540–542.
- Harrington, D., Mohi El Din, H., Guy, J., Robinson, K. & Sparagano, O. (2009) Characterization of the immune response of domestic fowl following immunization with proteins extracted from *Dermanyssus gallinae*. *Veterinary Parasitology*, <http://dx.doi.org/10.1016/j.vetpar.2008.11.004>.
- Kilpinen, O. (2001) Activation of the poultry red mite, *Dermanyssus gallinae* (Acari: Dermanyssidae), by increasing temperatures. *Experimental and Applied Acarology* **25**, 859–867.
- Kilpinen, O. (2005) How to obtain a bloodmeal without being eaten by a host: the case of the poultry red mite, *Dermanyssus gallinae*. *Physiological Entomology* **30**, 232–240.
- Kilpinen, O., Roepstorff, A., Permin, A., Norgaard-Nielsen, G., Lawson, L.G. & Simonsen, H.B. (2005) Influence of *Dermanyssus gallinae* and *Ascaridia galli* infections on behaviour and health of laying hens (*Gallus gallus domesticus*). *British Poultry Science* **46**, 26–34.
- Kirkwood, A.C. (1968) Some observations on the feeding habits of the poultry mites *Dermanyssus gallinae* and *Liponyssus sylviarum*. *Experimental and Applied Entomology* **11**, 315–320.
- Kocan, K.M., Yoshioka, J., Sonenshine, D.E., de la Fuente, J., Ceraul, S.M., Blouin, E.F. & Almazán, C. (2005) Capillary tube feeding system for studying tick-pathogen interactions of *Dermacentor variabilis* (Acari: Ixodidae) and *Anaplasma marginale* (Rickettsiales: Anaplasmataceae). *Journal of Medical Entomology* **42**, 864–874.
- Kowalski, A., Sokół, R. & Jedlińska-Krakowska, M. (2006) Influence of red mite *Dermanyssus gallinae* invasions on corticosterone and haematological levels and immunological indices in egg-laying hens. *Medycyna Weterynaryjna* **62**, 1188–1190.

- Kröber, T. & Guerin, P.M.** (2007a) *In vitro* feeding assays for hard ticks. *Trends in Parasitology* **23**, 445–449.
- Kröber, T. & Guerin, P.M.** (2007b) An *in vitro* feeding assay to test acaricides for control of hard ticks. *Pest Management Science* **63**, 17–22.
- Lucky, A.W., Sayers, C.P., Argus, J.D. & Lucky, A.** (2001) Avian mite bites acquired from a new source-pet gerbils. *Archives of Dermatology* **137**, 167–170.
- McDevitt, R., Nisbet, A.J. & Huntley, J.F.** (2006) Ability of a proteinase inhibitor mixture to kill poultry red mite, *Dermanyssus gallinae* in an *in vitro* feeding system. *Veterinary Parasitology* **141**, 380–385.
- Mignon, B. & Losson, B.** (2008) Dermatitis in a horse associated with the poultry mite (*Dermanyssus gallinae*). *Veterinary Dermatology* **19**, 38–43.
- Rosen, S., Yeruham, I. & Braverman, Y.** (2002) Dermatitis in humans associated with the mites *Pyemotes tritici*, *Dermanyssus gallinae*, *Ornithonyssus bacoti* and *Androlaelaps casalis* in Israel. *Medical and Veterinary Entomology* **16**, 442–444.
- Thind, B.B. & Ford, H.L.** (2007) Maintaining *Psoroptes ovis* (Acari: Psoroptidae) off-host in the laboratory. *Experimental and Applied Acarology* **43**, 205–211.
- Tucci, E.C.** (1997) A laboratory method for the rearing of *Dermanyssus gallinae* (De Geer, 1778) (Acari, Dermanyssidae). *Arquivos do Instituto Biológico* **64**, 1–4.
- Zeman, P.** (1988) Surface skin lipids of birds—a proper host kairomone and feeding inducer in the poultry red mite, *Dermanyssus gallinae*. *Experimental and Applied Acarology* **5**, 163–173.